

Circular RNA hsa_circ_0011946 promotes cell growth, migration, and invasion of oral squamous cell carcinoma by upregulating PCNA

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Abstract. – OBJECTIVE: The importance of circular RNAs in malignant tumors has been well concerned nowadays. Oral squamous cell carcinoma (OSCC) is diagnosed prevalent in the world. Our study aims to uncover the potential functions of hsa_circ_0011946 in OSCC development.

PATIENTS AND METHODS: Real Time-quantitative Polymerase Chain Reaction (RT-qPCR) was performed to determine the level of hsa_circ_0011946 in OSCC tissues and cell lines. Hsa_circ_0011946 was knocked down in OSCC cells. Biological functions of hsa_circ_0011946 in OSCC were identified by performing cell proliferation assay, colony formation assay, wound healing assay, and Transwell assay. The underlying mechanism of hsa_circ_0011946 in regulating OSCC progression was explored by RT-qPCR and Western blot assay.

RESULTS: Hsa_circ_0011946 was highly expressed in OSCC tissues compared with adjacent samples. It was also upregulated in OSCC cell lines. The knockdown of hsa_circ_0011946 inhibited cell growth, migration, and invasion in OSCC. The expression of PCNA was reduced via knockdown of hsa_circ_0011946. Furthermore, the expression of PCNA in tumor tissues was positively correlated to the expression of hsa_circ_0011946.

CONCLUSIONS: Hsa_circ_0011946 could promote cell growth, migration, and invasion of OSCC by upregulating PCNA, which may offer a new therapeutic intervention for OSCC patients.

Key Words:

Circular RNA, Hsa_circ_0011946, Oral squamous cell carcinoma, PCNA. Circular RNA Hsa_circ_0011946, Oral squamous cell carcinoma.

Introduction

Oral cancer is the sixth most common cancer worldwide, which is more prevalent in developing countries^{1,2}. Oral squamous cell carcinoma (OSCC) is the major subgroup of oral cancer, accounting for approximately 3% of all newly diagnosed clinical cancer cases³. Due to the invisible changes in the oral cavity, the early detection of OSCC is extremely important to improve the unsatisfied prognosis^{4,5}. Despite development has been made in the treatment and diagnosis for OSCC, the prognosis of OSCC remains poor, with the overall 5-year survival rate below 50%^{6,7}. Distant metastasis or local recurrences of OSCC contribute to the poor outcome. Thus, it is essential and urgent to find out important molecular markers involving in the development of OSCC and figure out new therapeutic treatments for OSCC patients.

As a novel class of noncoding RNAs, circular RNAs (circRNAs) are formed by the junction of the 3' end and 5' end. With the development of high-throughput sequencing technology, circRNAs are indicated to participate in the process of gene expressions. Moreover, increasing evidence has revealed that circRNAs function as important factors in tumorigenesis by sponging miRNAs. In particular, as a miR-1252 sponge, hsa_circ_0043256 inhibits cell proliferation and induces cell apoptosis in non-small cell lung cancer⁸. Circ-ABCB10 facilitates cell proliferation

and tumorigenesis in breast cancer by sponging miR-1271⁹. By regulating miR-1324/FZD5/Wnt/ β -catenin signaling, circ_0067934 facilitates tumor growth and cell migration in hepatocellular carcinoma¹⁰. By competing with microRNA-150-5p, circRNA ZNF609 enhances tumor growth and metastasis of nasopharyngeal carcinoma¹¹. However, the role of hsa_circ_0011946 in OSCC has not been studied.

In our report, hsa_circ_0011946 was upregulated in OSCC samples and cell lines. Moreover, knockdown of hsa_circ_0011946 inhibited tumor proliferation, migration, and invasion of OSCC *in vitro*. PCNA has been reported to participate in the progression of many cancers including OSCC. Furthermore, hsa_circ_0011946 knockdown was able to downregulate PCNA *in vitro*.

Patients and Methods

Tissue Samples

A total of 58 OSCC tissues and para-cancer tissues were obtained at the Fourth Affiliated Hospital, Harbin Medical University. The progression of OSCC patients was analyzed. None of the enrolled OSCC patients had preoperative radiotherapy or chemotherapy. The research was approved by the Ethics Committee of the Fourth Affiliated Hospital, Harbin Medical University. The signed written informed consents were obtained from all participants before the study.

Cell Culture

Human OSCC cell lines Tca8113, TSCCA, CAL-27, SCC9, and normal human oral keratinocytes (HOK) were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA). Cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM; Hyclone) with Log-Gro (AT, USA) supplemented with 10% fetal bovine serum (FBS; Gibco, Rockville, MD, USA) in an incubator containing 5% CO₂ at 37°C.

Cell Transfection

After OSCC cells were cultured for 24 h in 96-well plates, the cells were transfected with lentivirus targeting specifically targeting hsa_circ_0011946 (shRNA) or control (GenePharma, Shanghai, China) using Lipofectamine 3000 (Invitrogen, Carlsbad, CA, USA). GFP-positive cells were chosen for the following experiments.

RNA Extraction and Real Time-Quantitative Polymerase Chain Reaction (RT-qPCR)

TRIzol RNA isolation kit (Invitrogen, Carlsbad, CA, USA) was utilized to separate the total mRNA from tissues and cells. The synthesis of complementary deoxyribose nucleic acids (cDNA) was conducted through the reverse Transcription Kit (TaKaRa Biotechnology Co., Ltd., Beijing, China). The mRNA expression was normalized to that of glyceraldehyde 3-phosphate dehydrogenase (GAPDH). QRT-PCR reaction conditions were as follows: 94°C for 30 s, 55°C for 30 s, 72°C for 30 s, for a total of 40 cycles. Relative expression of the target gene was expressed by the $2^{-\Delta\Delta Ct}$ method. The primer sequences used for RT-PCR were as follows: Hsa_circ_0011946, forward: GCTGGT-GTTCCTTACTG-5'; Hsa_circ_0011946, reverse: 3'-CACTGTAGTAAACCAGCATTCT-5'; GAPDH primers forward: CCAAATCAGATGTCGCAATGCTGG-3', reverse: 5'-TGATGG-CTGGACTGTCTCATTCA-3'.

Cell Proliferation Assay

2×10⁴ cells were seeded in 96-well plates. The cell proliferation was assessed by Cell Proliferation Reagent Kit I (MTT; Roche, Basel, Switzerland) at 0 h, 24, 48, and 72 h following the manufacturer's protocol. Absorbance at 490 nm was assessed using an enzyme-linked immunosorbent assay (ELISA) reader system (Roche, Basel, Switzerland).

Colony Formation Assay

OSCC cells were inoculated in a 6-well plate for 10 days. Visible colonies were treated with 10% formaldehyde for 30 min and stained for 5 min with 0.5% crystal violet. The Image-Pro Plus 6.0 (Media Cybernetics, Silver Springs, MD, USA) was used for data analysis.

Wound Healing Assay

OSCC cells were seeded in 6-well plates and incubated in DMEM overnight. Cells were scratched with a plastic tip and cultured in serum-free DMEM. Each assay was repeated in triplicate independently. The relative distance of wound healing was viewed under a light microscope (Olympus, Tokyo, Japan) at 48 h.

Transwell Assay

After transfection, 1×10⁵ cells suspended in 200 μ L of serum-free DMEM were applied in the top chamber (Corning, Corning, NY, USA)

pre-coated with or without 50 μ g Matrigel (BD Biosciences, Franklin Lakes, NJ, USA). DMEM containing 10% FBS was added to the lower chamber. After overnight culture, the top surface of chambers was treated by methanol for 30 min after wiping off unpenetrated cells using a cotton swab. Then, they were stained in crystal violet for 20 min. Five fields per sample were randomly chosen and captured under a Leica DMI4000B microscope (Leica Microsystems, Heidelberg, Germany).

Western Blot Analysis

Protein was extracted from cells by the reagent radioimmunoprecipitation assay (RIPA; Yeasen, Shanghai, China). Dodecyl sulfate, sodium salt-polyacrylamide gel electrophoresis (SDS-PAGE) was utilized to extract the target proteins which were then transferred to polyvinylidene difluoride (PVDF) membranes (Millipore, Billerica, MA, USA). These membranes were incubated with antibodies rabbit anti-GAPDH and rabbit anti-PCNA (Cell Signaling Technology, Danvers, MA, USA) at 4°C overnight, and goat anti-rabbit secondary antibody (Cell Signaling Technology, Danvers, MA, USA) at room temperature for 2 h. Image J software (NIH, Bethesda, MD, USA) was applied for the assessment of the protein expression.

Statistical Analysis

Data analysis was performed using Statistical Product and Service Solutions (SPSS, SPSS Inc., Chicago, IL, USA). GraphPad 5.0 (Graph-

Pad Software, Inc., La Jolla, CA, USA) helped to present these consequences. The differences between the two groups were compared by the Student's *t*-test. The statistical significance was defined as $p < 0.05$.

Results

The Expression of Hsa_circ_0011946 in OSCC Tissues and Cells

RT-qPCR was used to detect hsa_circ_0011946 expression in 58 OSCC tissues samples and adjacent normal ones. As shown in Figure 1A, hsa_circ_0011946 expression was higher in tumor tissue samples than that in normal samples. Moreover, hsa_circ_0011946 expression was also higher in OSCC cell lines (Tca8113, TSCCA, CAL-27, SCC-9) than that in normal human oral keratinocytes (NHOK) (Figure 1B), suggesting that the abundant expression of hsa_circ_0011946 might be associated with OSCC development.

Downregulation of Hsa_circ_0011946 Inhibits OSCC Proliferation in OSCC Cells

To explore the effect of hsa_circ_0011946 on OSCC proliferation, MTT assay, colony formation assay, and cell cycle assay were conducted. CAL-27 cell line was selected for transfection of hsa_circ_0011946 lentivirus. RT-qPCR was used to measure the transfection efficiency (Figure 2A). MTT assay showed that the cell growth ability of OSCC was significantly repressed after hsa_circ_0011946 was

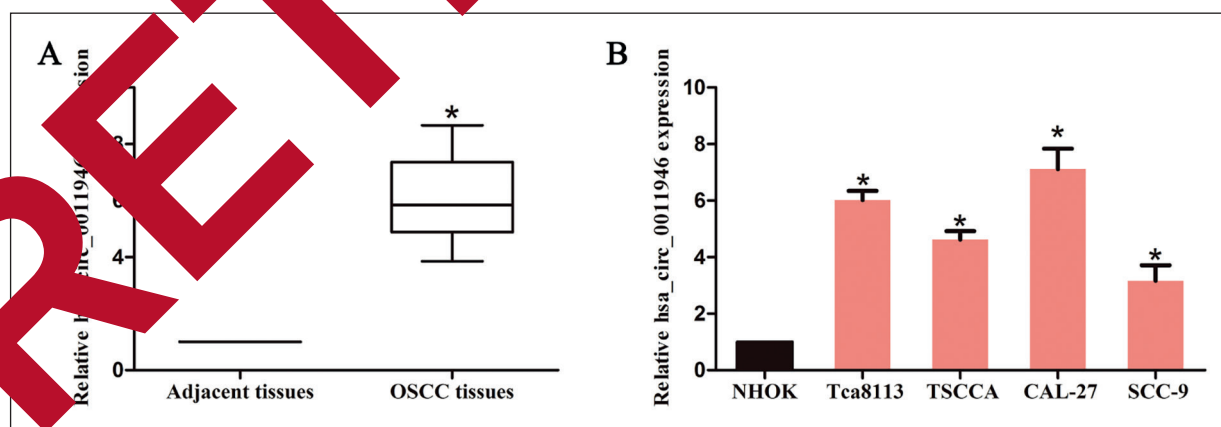


Figure 1. Expression levels of hsa_circ_0011946 were upregulated in OSCC tissues and cell lines. **A**, Hsa_circ_0011946 expression was significantly upregulated in the OSCC tissues compared with adjacent tissues. **B**, Expression level of hsa_circ_0011946 relative to GAPDH was determined in the human OSCC cell lines and normal human oral keratinocyte (NHOK) by RT-qPCR. GAPDH was used as an internal control. Data are presented as the mean $\pm$ standard error of the mean. * $p < 0.05$.

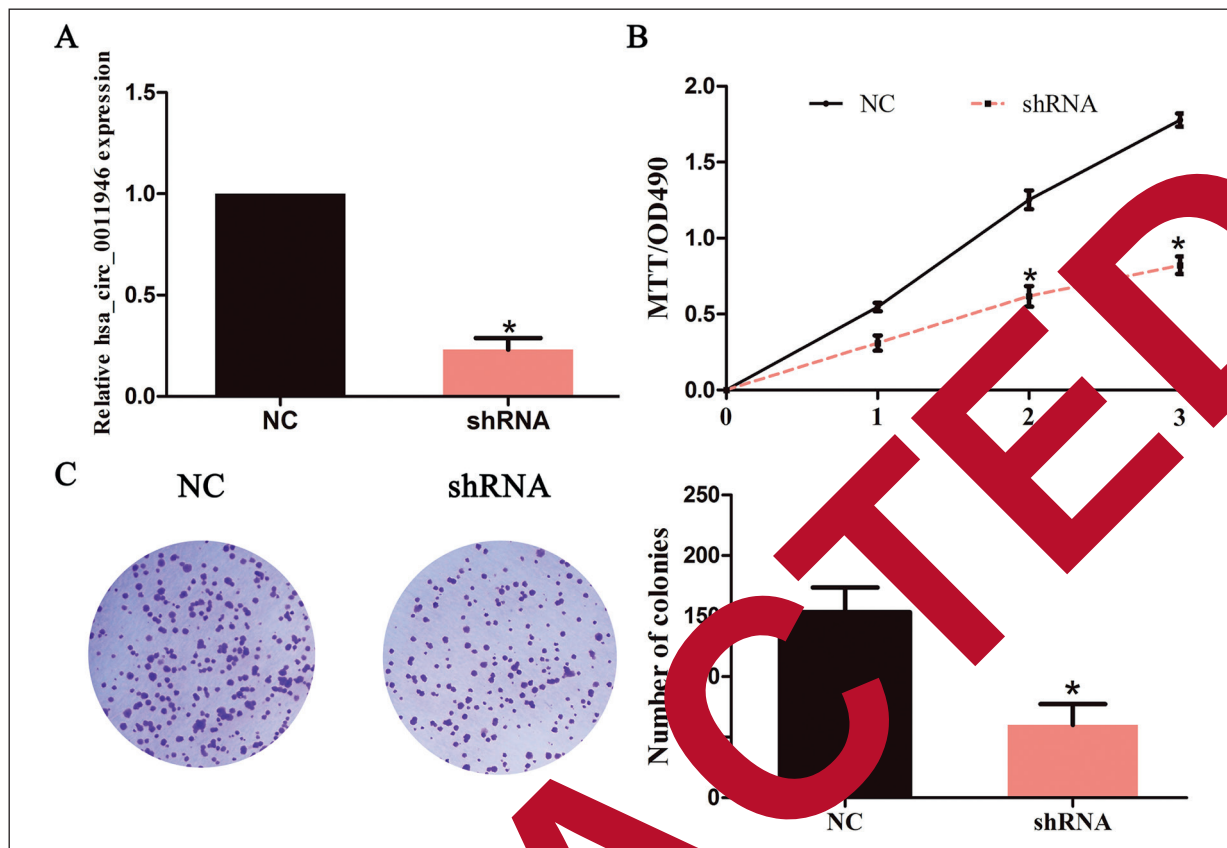


Figure 2. Downregulation of hsa_circ_0011946 inhibits OSCC cell proliferation. **A**, Hsa_circ_0011946 expression in OSCC cells transfected with hsa_circ_0011946 shRNA or NC was detected by RT-qPCR. **B**, MTT assay showed that downregulation of hsa_circ_0011946 significantly repressed the viability of OSCC cells. **C**, Colony formation assay showed that the number of colonies significantly decreased after downregulation of hsa_circ_0011946 in OSCC cells (magnification: 40 $\times$). The results represent the average of three independent experiments (mean $\pm$ standard error of the mean). * $p < 0.05$, as compared with the control cells.

downregulated (Figure 2B). Colony formation assay showed that the colony number was significantly reduced after the knockdown of hsa_circ_0011946 (Figure 2C).

Downregulation of Hsa_circ_0011946 Inhibited Cell Migration and Invasion in OSCC Cells

To further explore the regulatory effects of hsa_circ_0011946 on cell migration and invasion of OSCC cells, wound healing assay and transwell assay were performed. As shown in Figure 3A, the migrated length of OSCC cells was reduced after the knockdown of hsa_circ_0011946. As shown in Figure 3B, the number of migratory cells remarkably decreased after the silence of hsa_circ_0011946 in OSCC cells. As shown in Figure 3C, after hsa_circ_0011946 was downregulated, the number of invasive cells was remarkably reduced.

Downregulation of Hsa_circ_0011946 Repressed PCNA Expression in OSCC

Our previous study showed that hsa_circ_0011946 could promote tumor proliferation and metastasis of OSCC. The downstream target of hsa_circ_0011946 remained unknown. To explore the potential targets of hsa_circ_0011946, Starbase v2.0 was utilized for the prediction. In our work, the interaction between PCNA and hsa_circ_0011946 was firstly researched. RT-qPCR was performed to detect PCNA expression in OSCC cells transfected with hsa_circ_0011946 shRNA or negative control (NC). Results revealed that the downregulation of hsa_circ_0011946 decreased the mRNA expression of PCNA (Figure 4A). Moreover, the protein level of PCNA was measured through the Western blot assay. Downregulation of hsa_circ_0011946 overexpression reduced the protein level of PCNA as well (Figure 4B). Besides, the Pearson correlation analysis

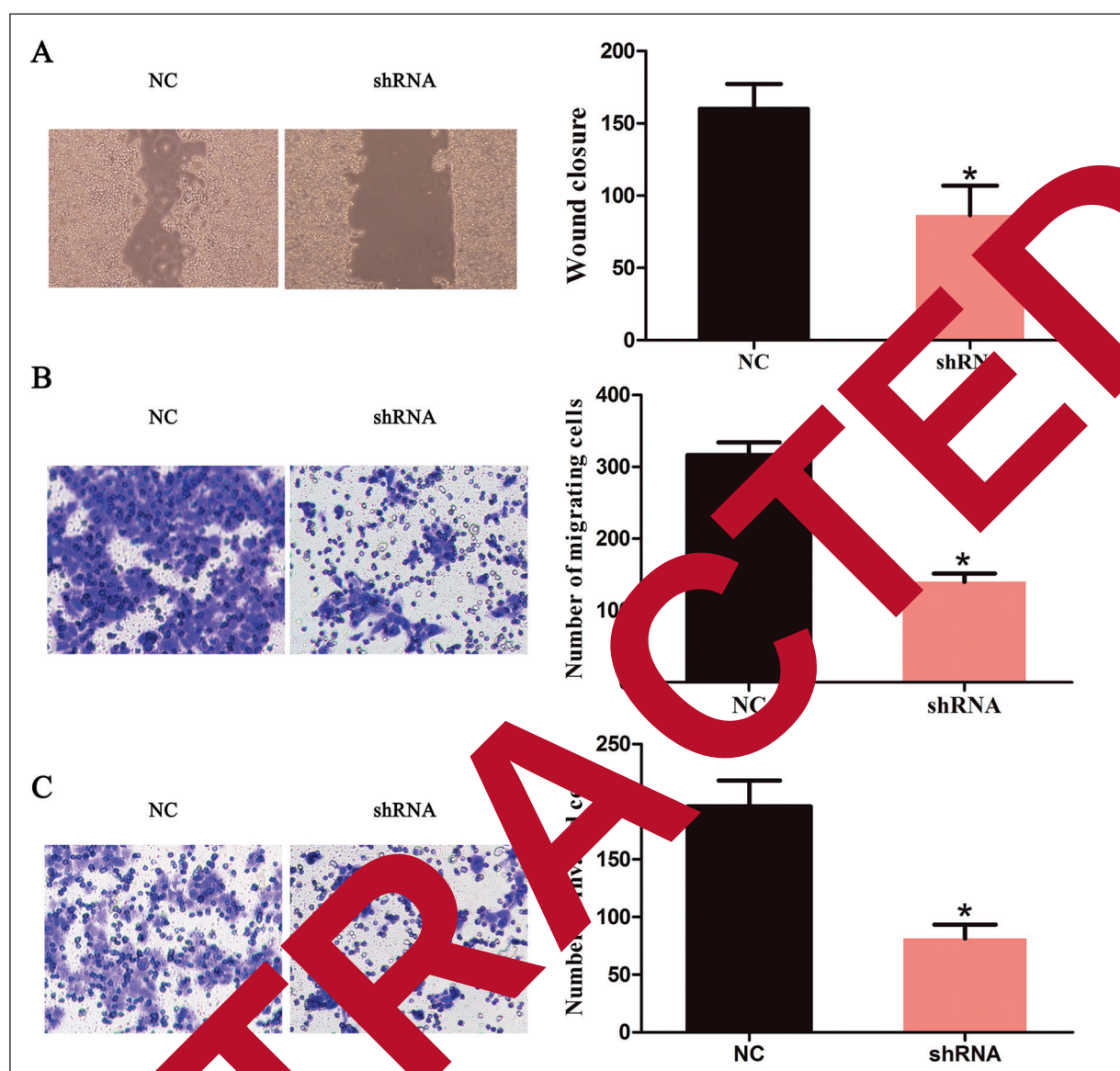


Figure 3. Downregulation of hsa_circ_0011946 inhibited OSCC cell migration and invasion. **A**, Wound healing assay showed that knockdown of hsa_circ_0011946 significantly repressed migrated length of OSCC cells (magnification: 10 $\times$). **B**, Transwell assay showed that knockdown of hsa_circ_0011946 significantly repressed cell migration in OSCC cells (magnification: 40 $\times$). **C**, Transwell assay showed that knockdown of hsa_circ_0011946 significantly repressed cell invasion in OSCC cells (magnification: 40 $\times$). The results represent the average of three independent experiments (mean $\pm$ standard error of the mean). $P < 0.05$, * compared with the control cells.

that PCNA expression level was positively correlated to hsa_circ_0011946 expression in OSCC tissues (Figure 4C).

Discussion

Increasing evidence has indicated the critical functions of circRNAs in modulating tu-

mor development. Serving as a sponge to miR-196a5p, circDOCK1 inhibits cell apoptosis in OSCC by targeting BIRC3¹². Downregulation of hsa_circ_0109291 suppresses cell proliferation and migration in OSCC, which may be a novel therapeutic target for OSCC¹³. By regulating the AKT/mTOR signaling, the upregulation of hsa_circ_0007059 restrains tumorigenesis and facilitates cell apoptosis in OSCC¹⁴.

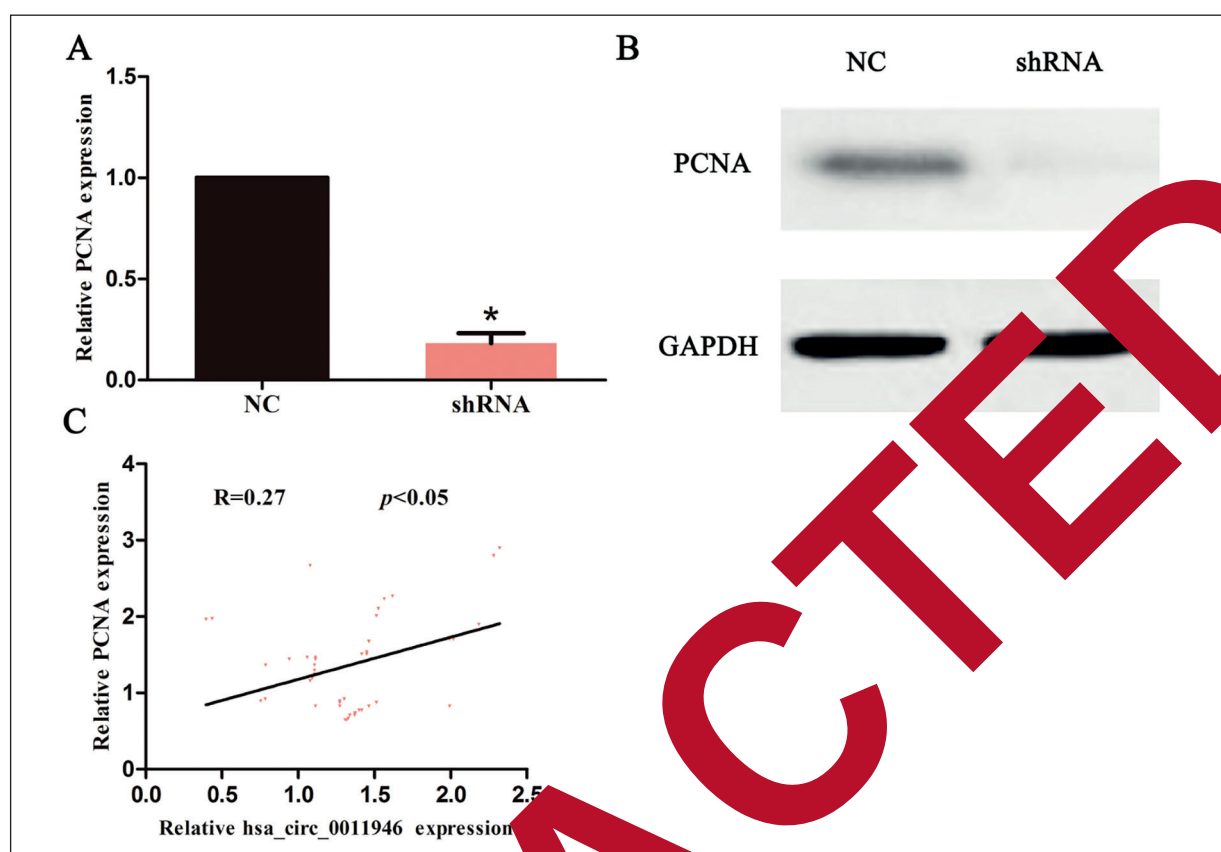


Figure 4. Downregulation of hsa_circ_0011946 inhibits PCNA expression in OSCC cells. **A**, RT-qPCR results showed that PCNA expression was downregulated in hsa_circ_0011946 shRNA group compared with NC group in OSCC cells. **B**, Western blot results showed that PCNA expression was downregulated in hsa_circ_0011946 shRNA group compared with NC group. **C**, Linear correlation between the expression level of PCNA and hsa_circ_0011946 in OSCC tissues. The results represent the average of three independent experiments. Data are presented as the mean $\pm$ standard error of the mean. * $p<0.05$.

Through the EGF pathway, downregulation of hsa_circ_000523 inhibits cell migration and proliferation in OSCC.

Hsa_circ_0011946 is a novel circRNA, which has been reported to be downregulated in many cancers. In addition, low expression of hsa_circ_0011946 inhibits cell migration and invasion in breast cancer by targeting RFC3¹⁶.

To further determine the function of hsa_circ_0011946 in OSCC, hsa_circ_0011946 shRNA was constructed for transfection in OSCC cells. Functional assays showed that downregulation of hsa_circ_0011946 repressed proliferation of OSCC cells. We further explored the effect of hsa_circ_0011946 on metastasis of OSCC cells. Results showed that the downregulation of hsa_circ_0011946 contributed to the inhibition of migration and invasion in OSCC cells. All these results indicated that hsa_circ_0011946 inhibited tumor proliferation and metastasis of OSCC.

Then, the downstream proteins of hsa_circ_0011946 were further explored. Proliferating cell nuclear antigen (PCNA), initially discovered in cancer cells and proliferating cells, was predicted as the target of hsa_circ_0011946 through bio-informative analysis. Numerous reports have proved that PCNA is directly correlated to tumor differentiation, tumor staging, and prognosis. The expression level of PCNA is significantly higher in colorectal cancer, especially in those with liver metastases, which may help to assess organ metastatic condition in patients with colorectal cancer¹⁷. The positive expression rate of PCNA was 73% in breast cancer tissues, indicating the important clinical significance of PCNA in evaluating the prognosis of breast cancer¹⁸. Besides, the upregulation of PCNA is closely related to poor prognosis of patients with osteosarcoma, which may be a potential biomarker¹⁹.

The potential interaction between PCNA and hsa_circ_0011946 was first researched in our study. Results showed that hsa_circ_0011946 knockdown decreased PCNA expression *in vitro* and was positively correlated with PCNA expression in OSCC tissues. Above results indicated that hsa_circ_0011946 might promote tumor development of OSCC by upregulating PCNA.

Conclusions

The above results provided evidence that hsa_circ_0011946 was remarkably upregulated in OSCC tissues, which promotes cell proliferation and metastasis of OSCC by upregulating PCNA. Our findings uncovered that hsa_circ_0011946 may contribute to therapy for OSCC as a prospective target.

Conflict of Interest

The Authors declare that they have no conflict of interests.

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