

The functional analysis of MicroRNAs involved in NF- κ B signaling

Y. YANG, J.-K. WANG

The State Key Laboratory of Bioelectronics, Southeast University, Nanjing, People's Republic of China

Abstract. – Nuclear factor κ B (NF- κ B) is a transcriptional factor that regulates a large number of genes that controls diverse biological functions, ranging from inflammation, cell proliferation and tumor development to learning and memory. MicroRNAs (miRNAs) are small non-coding RNA molecules involved in most aspects of physiological and pathological processes, including cancer, viral infections, inflammation and autoimmune diseases. miRNAs also play an important role in the regulation of NF- κ B signaling pathway, some being inhibitory and others activating. Here, we analyzed the convergence of miRNAs involved in NF- κ B signaling regulation and dysregulation of miRNAs and NF- κ B activation in human diseases, particularly in cancer. The function of miR-146, miR-125b, miR-21, miR-301a, miR-30b, and miR-199 and their impacts on tumorigenesis are analyzed in this work. miRNAs as one of the most abundant classes of regulatory molecules, deciphering their biological function and pathological contribution in NF- κ B dysregulation is essential to understand the complexity of immune systems and to develop therapeutics against cancer.

Key Words:

NF- κ B, MicroRNAs, Cancer, Inflammation.

Introduction

NF- κ B is a dimeric transcriptional factor first defined by Ranjan, which was latently present in cells and could be induced into its DNA-binding state, known as κ B site¹. All NF- κ B proteins share a reticuloendotheliosis (Rel) homology domain (RHD), which is essential for binding to cognate DNA sequence motifs and dimerization to the members of NF- κ B proteins as well as nuclear translocation². The Rel protein family consists of five members, including p50, p52, p65, RelB and c-Rel³. Except for RelB that can only form heterodimers, all Rel proteins can form homodimers or heterodimers⁴. Among

these proteins, p50-p65 heterodimer is the most abundant form of NF- κ B in most unstimulated cells, which we discussed in this work unless indicated otherwise. NF- κ B activation is mainly regulated by two pathways in response to extracellular stimuli⁵. The classical pathway is usually induced by microbial, viral infections and proinflammatory cytokines, such as tumor necrosis factor (TNF α), which all can activate the β -subunit of I κ B kinase (IKK β) complex through the toll-like receptor (TLR). I κ B kinases (IKKs) phosphorylate I κ Bs (inhibitors of κ B) binded to NF- κ B, resulting in ubiquitin-dependent degradation of I κ Bs and translocation of NF- κ B dimers to the nucleus⁶. The non-classical pathway is induced by certain members of the TNF cytokine family that selectively activate the α -subunit of IKK (IKK α) through the TNF receptor, BAFFR, RANK (receptor activator for nuclear factor kappaB), TNFR2, Fn14 and CD40R, along with NF- κ B inducing kinase (NIK), to phosphorylate p100. This phosphorylation leads to polyubiquitination-dependent degradation of p100 to generate p52, forming p52-RelB heterodimers, which then translocate to the nucleus and activate target genes⁷.

NF- κ B plays an important role in regulating a large number of genes as well as the regulation of innate and adaptive immunity^{8,9}, cell proliferation^{10,11}, inflammation^{12,13}, tumorigenesis and tumor progression^{14,15}. Therefore, NF- κ B is a pluripotent and vital transcription factor for physiological and pathological processes. Besides, its transcriptional regulation system is complex, especially considering the fact that different NF- κ B dimers have different affinities for different DNA-binding sequences¹⁶ and various target genes are differentially regulated by distinct NF- κ B dimers in various cell contexts¹⁷. In addition, for activation and crosstalk with other signaling pathways, NF- κ B subunits contain sites available for phosphorylations and other post-translational modifications¹⁸.

MicroRNAs (miRNAs) are a subclass of short (20-23 nucleotides in length), endogenous, non-coding, single-stranded RNAs that regulate gene expression post-transcriptionally by binding mainly to the 3' untranslated region (UTR) of target mRNAs. miRNAs are transcribed mostly by RNA polymerase II, as a long primary miRNA transcript (pri-miRNA)¹⁹ with a stem-loop structure²⁰. It is, then, recognized and cleaved in the nucleus by the microprocessor complex, Drosha-DGCR8 (DiGeorge syndrome critical region gene 8), resulting in a hairpin-structured precursor of miRNAs (pre-miRNA) ranging from 60 to 110 nucleotides in length. The pre-miRNA is exported from the nucleus to the cytoplasm by a nuclear transport receptor (exportin-5) and Ran-GTP^{21,22}. In the cytoplasm, Dicer cleaves the pre-miRNA hairpin into a ~22 bp miRNA duplex^{23,24}. The mature miRNA is incorporated with Argonaute (Ago2) proteins into the RNA-induced silencing complex (RISC)²⁵⁻²⁷, where miRNA guides the complex to partial complementary binding sites located in the 3' untranslated region (UTR)²⁸, 5' untranslated region (UTR)^{29,30}, or coding regions^{31,32} of target mRNAs to induce translational repression or degradation of targeted mRNAs. Moreover, a small part of miRNAs are derived from introns of protein-coding genes, termed as mirtrons^{20,33}; the mirtron production is Drosha-independent to generate pre-miRNAs and is spliced by Spliceosome³⁴.

miRNAs-mediated gene silencing was via RISC to induce translational repression or degradation of targeted mRNAs. Numerous investigations have confirmed the important roles of miRNAs in the regulation of human cancer, as well as in physiological function including immune responses, cellular proliferation, differentiation, and apoptosis³⁵⁻³⁷. These processes are also known to be regulated by NF- κ B¹¹ and miRNAs play an important role in modulating NF- κ B signaling pathway^{38,39}. Thus, NF- κ B and miRNAs play important roles in the gene expression regulatory network of the organism. Here we analyzed some key miRNAs involved in NF- κ B signal pathway regulation (Figure 1).

miR-146

miR-146 is a vital modulator of differentiation and biology function of cells for the innate and adaptive immunity. In addition, it plays an important role in regulating different types of diseases and cancers⁴⁰. Induction of miR-146 is NF- κ B dependent through the toll-like

receptor (TLR) and its expression is upregulated in human monocytic THP-1 cells treated by lipopolysaccharides (LPS)⁴¹. As to the NF- κ B signal pathway regulation, TRAF6 and IRAK1 were identified as direct targets of miR-146. Generally, NF- κ B activation upregulates miR-146 gene expression and, then, miR-146 down-regulates IRAK1 and TRAF6 to suppress the activity of NF- κ B. Thus, this is a negative regulatory loop. According to previous literatures, miR-146 was demonstrated as an NF- κ B negative regulator⁴² and it exhibited an important role in suppressing tumor genesis and progression by inhibiting tumor cell migration and invasion^{42,43}.

In the stressed human brain primary neural cells, the close connection between NF- κ B and miR-146a was confirmed by experiments which had tremendous potential to modulate neurotrophic support, neuroinflammation, synaptogenesis, innate immune signaling and amyloidogenesis⁴⁴. Moreover, miR-146a was highly complementary to the 3'-untranslated region of complement factor H (CFH), an important repressor of the inflammatory response of the brain. Up-regulation of miR-146a accompanied with down-regulation of CFH was discovered in Alzheimer disease (AD) brain and as well as interleukin-1 β , A β 42, and/or oxidatively stressed human neural (HN) cells in primary culture. It indicated that miR-146a-mediated modulation of CFH gene expression could affect the inflammatory response in AD brain and stressed HN cells. In summary, miRNAs could be an effective therapeutic target for treatment of AD disease and inflammation⁴⁵. Besides, the NF- κ B-miR-146a complex, as a novel regulatory mechanism, was approved to attenuate inflammation in response to respiratory toxicants through suppressing the expression of cyclooxygenase-2 (COX-2) in mouse lung fibroblasts⁴⁶. It was reported that IL-1 β could induce upregulation of miR-146a, which in turn negatively regulates the expression of proinflammatory chemokines IL-8 and CCL5 in human lung alveolar epithelial tumor A549 cells. Besides, IL-8 and CCL5 are regulated by NF- κ B activation, which provided additional evidence for the negative feedback regulation of inflammation response by miRNAs and NF- κ B⁴⁷. However, IRAK1 and TRAF6 were not involved in the pathway in A549 cell, which indicated that the regulatory effects of miR-146a may be cell type-specific⁴⁰. All in all, miR-146 is a target gene of NF- κ B and it could negatively modulate IRAK1

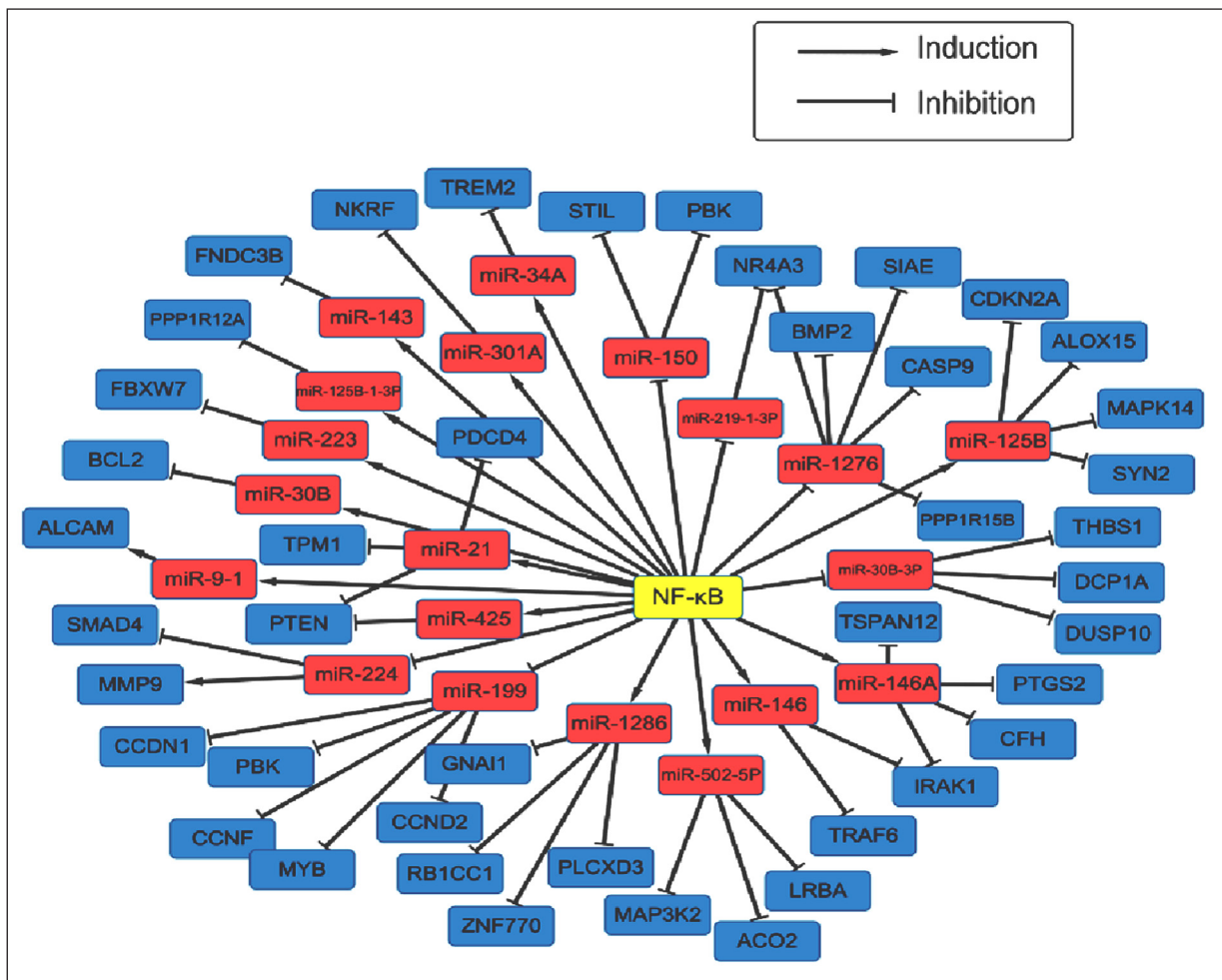


Figure 1. Panoramic view of the NF- κ B miRNA target genes and target genes of miRNAs.

and TRAF6, constituting a negative feedback loop. miR-146 is involved in the regulation of the adaptive and innate immune response and tumor progression. Further studies were needed to fully understand the relationship between miR-146 and NF- κ B.

miR-125b

Previous literature showed that up-regulation of miR-125b is associated with the suppression of the 15-lipoxygenase (ALOX15) and the synaptic vesicle-associated phosphoprotein synapsin-2 (SYN-2) in the brain. This indicated that miR-125b was involved in the regulation of innate and adaptive immune signaling, inflammation response, synaptogenesis, amyloidogenesis and neurotrophic support⁴⁴. As one of the most human brain abundant miRNAs⁴⁸, miR-125b was shown to be up-regulated by neurotoxic metal sulfates and was also up-regulated in brain

cancers to suppress the expression of cyclin-dependent kinase inhibitor 2A (CDKN2A), which is a negative regulator of cell growth. These effects of miRNA-125b were related to regulation of astrogliosis and cell cycle^{49,50}.

A previous study⁵² indicated that miR-125a and miR-125b constitutively activate NF- κ B by repressing TNF α -induced protein 3 (TNFAIP3)⁵¹. TNFAIP3 is a critical inhibitor of NF- κ B signaling. Thus, miR-25b-1 plays a vital role in the NF- κ B signal pathway through forming a positive feedback regulation loop⁵⁵. Meanwhile, up-regulation of miR-125b by ultraviolet can promote cell survival in human embryonic kidney cell line HEK293 and keratinocyte cell line HaCaT through preventing prolonged p38 α activation⁵⁶. PPP1R12A, a gene regulating cell survival, is the target gene of miR-125b-1⁵⁵. All these researches suggest that miR-125b plays a role in the regulation of cell survival.

miR-21

The overexpression of miR-21 was identified in most types of human carcinomas⁵⁷, and NF- κ B activation is also reported in all of these cancers¹¹, indicating the interplay of miR-21 and NF- κ B in cancer. In human breast cancer cells, NF- κ B-dependent miR-21 up-regulation following genotoxic treatment contributes to both therapeutic resistance and metastasis through repressing expression of PTEN and PDCD4⁵⁸. Knockdown of miR-21 using peptide nucleic acids (PNAs) inhibits proliferation and migration of MCF-7 and MDA-MB-231 cells⁵⁹. The up-regulation of miR-21 promotes growth, migration, invasion and chemo/radio-resistance of non-small cell lung cancer cells through suppressing its target's expression of PTEN, a tumor suppressor gene⁶⁰. The up-regulation of miR-21 by STAT3 under the treatment of IL-6 induces enhanced proliferation and suppressed apoptosis in human nasopharyngeal carcinoma (NPC) through PTEN-AKT pathway⁶¹. In human skin and head and neck squamous cell carcinoma (SCC), up-regulation of miR-21 also suppresses the development related transcription factor GRHL3 and PTEN, a direct GRHL3 target, through PI3K/AKT/mTOR signaling pathway, promoting tumorigenesis⁶². In human glioblastoma tissue and glioblastoma-derived cell lines, either downregulation of mir-21 or up-regulation of its target, programmed cell death 4 (Pcd4), leads to decreased proliferation, colony formation, and increased apoptosis⁶³. The up-regulation of mir-21 possesses the important significances as the indicators of prognostic and tumor stage in human hepatocellular carcinoma (HCC)⁶⁴. Furthermore, Ma et al. confirmed that knocking out the miR-21 allele in mice could promote cell apoptosis and suppress cell proliferation, which showed that miR-21 plays its oncogenic function through downregulating its target genes such as Spry1, Pten, and PDCD4⁶⁵. In all, mir-21 may act as a potential diagnostic and prognostic biomarker^{66,67} and a novel therapeutic target for cancers.

In human glioblastoma (GBM) cell lines, downregulation of miR-21 could suppress cell proliferation and induced cell apoptosis through inhibiting EGFR pathway with the manner of PTEN-independent⁶⁸. miR-21 induced by LPS attenuates pro-inflammatory effects of TLR4 signaling through suppressing NF- κ B activity⁶⁹. Mice deficient in PDCD4, a confirmed miR-21 target, exhibit the lower LPS-induced mortality rates, lower IL-6 production and increased IL-

10 protein levels compared to the WT mice. Meanwhile, reduction of PDCD4 by increased miR-21 expression can account for the increased neoplastic transformation in mice JB6 cell lines⁷⁰. As depletion of the NF- κ B subunit p65 abolished LPS-induced miR-21 expression, the authors then show that miR-21 is an NF- κ B transactivational gene⁷¹. Moreover, during earlier stages of liver regeneration, the up-regulation of miR-21 leads to down-regulation of pellino1, an activator of NF- κ B, indicating that miR-21 may act as an NF- κ B inhibitor⁷². Thus, cell-type specificity may determine the various functions of miR-21 in NF- κ B signal pathway. Generally, in epithelial cells, miR-21 acts to down-regulate PTEN, activate AKT, and induce NF- κ B activation. On the other hand, miR-21 can act as an NF- κ B inhibitor to suppress PDCD4, a proinflammatory protein that promotes activation of NF- κ B in LPS-stimulated macrophages. It needs more research to dissect the correlation of miR-21 overexpression and NF- κ B activation in cancer, as well as the role of miR-21 in NF- κ B signaling, inflammation and immune diseases.

miR-301a

miR-301a was identified as an activator of NF- κ B by negatively regulating its target gene of NF- κ B repressing factor (NKRF)⁷³; NKRF broadly suppressed the expression of NF- κ B transactivational targets. Besides, the promoter of miR-301a contained a bona fide κ B site. Thus, it forms a positive feedback loop as a mechanism for persistent NF- κ B activation in which miR-301a suppresses NKRF to activate NF- κ B activity, which in turn, promote the expression of miR-301a.

miR-301a, which is the most potent NF- κ B activator is over expressed in pancreatic adenocarcinoma and other tumor cell lines⁷³. Over expression of miR-301a promoted pancreatic cancer (PC) cell proliferation, and repressed the expression of Bim gene *in vitro* and *in vivo*. Meanwhile, Bim re-expression could suppress PC cell proliferation induced by miR-301a⁷⁴. In human pancreatic ductal adenocarcinoma (PDAC), miR-301a over expression promotes cancer growth through suppressing manganese superoxide dismutase (MnSOD) expression, a tumor suppressor gene. On the other hand, decreased miR-301a levels are associated with increased MnSOD expression and inhibition of PDAC growth⁷⁵. In human colorectal cancer, the up-regulation of miR-301a represses the

expression of suppressor of cytokine signaling 6 (SOCS6)⁷⁶ and Smad4 through TGF- β /Smad pathway⁷⁷, which in turn, promotes cell proliferation, migration and invasion and tumor growth. Meanwhile, miR-301a is an activator of both NF- κ B and Stat3, generating a pro-inflammatory microenvironment that promotes colorectal cancer as well as lung cancer tumorigenesis⁷⁸. Suppression of miR-301a can repress tumor cells proliferation, migration and invasion. It indicates that miR-301a acts as an oncogene miRNAs facilitating tumorigenesis. The significant up-regulation of miR-301a both in cells and tissues of gastric cancer can promote cell proliferation, soft agar clonogenicity, cell migration and invasion through downregulating RUNX3 expression⁷⁹, which plays a key role in the clinical progression and prognosis of gastric cancer⁸⁰. The significant up-regulation of miR-301a and downregulation of Gax in human hepatocellular carcinoma (HCC) promote cell proliferation, migration and invasion, while inhibiting miR-301a expression induces the up-regulation of Gax and repression of NF- κ B expression⁸¹. Moreover, miR-301a plays an important role in prostate cancer through regulating the miR-301a/androgen receptor (AR)/TGF- β 1/Smad/MMP9 signals pathway⁸² and in autoimmune demyelination through regulating immune response⁸³. Intriguingly, over expression of miR-301a promotes breast cancer cell migration, invasion and metastasis with the hyper-activation of Wnt/ β -catenin signaling through suppression of PTEN expression⁸⁴. Meanwhile, the up-regulation of miR-301a and down-regulation of PTEN, a target of miR-301a, reduces the effect of IL-6-induced insulin resistance and hepatic glycogenesis through the AKT/GSK pathway⁸⁵. It shows that PTEN is a vital component in miR-301a target genes. Altogether, miR-301a exerts important roles in many physiological processes and is an important therapeutic target for cancers.

miR-30b

In HER2-positive breast cancer cells, the up-regulation of miR-30b induced by trastuzumab can inhibit cell growth through repressing CCNE2⁸⁶. Up-regulation of miR-30b can promote the apoptosis of gastric cancer cells and significantly inhibit tumorigenicity of gastric cancer through negatively regulating its target of plasminogen activator inhibitor-1 (PAI-1)⁸⁷. miR-30b expression in human colorectal cancer (CRC)

is significantly lower than that in normal tissues. The literature showed that over expression of miR-30b could suppress cell proliferation in vitro and tumor growth *in vivo* through regulating its target genes of KRAS, PIK3CD and BCL2. These researches indicated that miR-30b could act as a potential prognostic marker and therapeutic target for CRC⁸⁸. In laryngeal carcinoma cells, the up-regulation of miR-30b can promote p53-mediated tumor cell apoptosis⁸⁹. However, the up-regulation of miR-30b in glioma cells impairs tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)-dependent apoptosis by inhibiting the expression of caspase-3⁹⁰. And the up-regulation of miR-30b in human melanoma promotes the metastatic behavior of melanoma cells by repressing the GalNAc transferase GALNT7, which may lead to increased synthesis of the immunosuppressive cytokine IL-10, and decreased immune cell activation and recruitment⁹¹. Thus, the opposite function of miR-30b in cancers can be explained by its cell type-specific function.

Moreover, miR-30b is a negative regulator of cell death induced by loss of attachment (anoikis) through regulating the expression of caspase 3⁹². miR-30b regulates cell death in cardiomyocytes by repressing Bcl-2⁹³. miR-30b also plays an important role in schizophrenia⁹⁴, angiogenesis⁹⁵ and phagocytosis⁹⁶.

miR-199

The down-regulation of miR-199a/b is found in non-small cell lung cancer (NSCLC), which promotes cell proliferation, migration and invasion through negatively regulating Axl expression⁹⁷. The up-regulation of miR-199a can increase survival in aggressive diffuse large B-cell lymphoma patients by modifying drug sensitivity to immunochemotherapy⁹⁸. The up-regulation of miR-199a suppresses renal cancer cell growth and expression of GSK-3 β , which indicates that miR-199a can act as a potential therapeutic target of renal cancer⁹⁹. The down-regulation of miR-199a induced by reactive oxygen species (ROS) in ovarian cancer cells can elevate the expression of ERBB2 and ERBB3, which in turn promote cancer progression¹⁰⁰. On the other hand, miR-199a-3p is significantly up-regulated in gastric cancer (GC) cell lines and tissues which promote cell proliferation, suppresses cell apoptosis through suppressing the expression of zinc fingers and homeoboxes 1 (ZHX1)¹⁰¹. The up-regulation of miR-199a-3p in colorectal cancer suppresses the expression of its target gene NLK, which in turn

to promote the lymph node metastasis, venous invasion, liver metastasis of colorectal cancer¹⁰². Thus, the members of miR-199 have diverse functions in different cancers.

In primary hepatocellular carcinomas (HCCs) and HCC cell lines, miR-199 can modulate Ad-199T virus replication, which is an oncolytic adenovirus. This indicated that miR-199 can be used as a therapeutic potential against liver cancer¹⁰³. The up-regulation of miR-199a is positively and significantly correlated to the progression of liver fibrosis, while the expression levels of fibrosis-related genes in hepatic stellate cells (HSC) are significantly increased by over-expression of miR-199a¹⁰⁴. Generally, chronic hepatitis can develop into liver cirrhosis (LC) and hepatocellular carcinoma (HCC) consequently. Thus, miR-199 plays important roles in the physiological progression of liver diseases. Interestingly, miR-199 also plays an important role in somatic cell reprogramming through the p53 signal pathway¹⁰⁵. In summary, the members of miR-199 possess the vital functions in many physiological processes and more researches are needed to uncover its full functions and mechanisms.

Other miRNAs Target Genes Related to NF- κ B

There are many other miRNAs that are related to NF- κ B signal pathway. In human biliary epithelial cells, miR-125b-1, miR-21, miR-30b and miR-23b-27b-24-1 genes involve in the immune responses following *C. parvum* infection being relevant to the regulation of epithelial anti-microbial defense¹⁰⁶. In Alzheimer's disease, up-regulation of miR-34a represses TREM2 expression and may shape innate immune and phagocytic responses that contribute to inflammatory neurodegeneration through an epigenetic mechanism¹⁰⁷. In human esophageal squamous cancer EC109 cell, up-regulation of miR-34a transcribed by NF- κ B and p53 plays an important role in tumor progression¹⁰⁸. NF- κ B and p53 also repress miR-224 expression and induce Smad4 expression to influence the proliferation of mouse ovarian granulosa cells¹⁰⁹. Celastrol can inhibit the migration and invasion of HepG2 cells by efficiently decreasing the expression of miR-224 and MMP-2 and MMP-9¹¹⁰. The up-regulation of miR-223 expression represses the tumor suppressor FBXW7 in T-cell acute lymphoblastic leukemia (T-ALL), which in turn to promote cancer progression through Notch signal

pathway¹¹¹. Up-regulation of miR-143 expression promotes HCC invasion/migration and tumor metastasis by repression of fibronectin type III domain containing 3B (FNDC3B) expression¹¹². Up-regulation of miR-425 expression increases gastric cancer cell survival by repressing PTEN expression¹¹³. In human hepatoma cell lines, HepG2, GQY-7701 and Bel-7402, up-regulation of miR-9 represses CD166 expression and promotes cells migration¹¹⁴. The up-regulation of miR-145 inhibits glucose uptake and induces insulin resistance through repressing IRS-1 expression in HepG2 cells¹¹⁵. In summary, miR-155, miR-193b, miR-34a, miR-451, miR-150 and miR-199 involve in the transformation of human B-cells and diffuse large B cell lymphoma (DLBCL) through NF- κ B pathway¹¹⁶.

Conclusions

miRNAs act as posttranscriptional regulators of gene expression and regulate many target genes including NF- κ B, I κ B, IKK and regulators in the NF- κ B signaling pathway forming positive or negative sophisticated feedback loops. miRNAs constitute an important layer of regulation of gene expression with profound impacts on biological organisms. miRNAs have the unique expression profile in cells of the innate and adaptive immune system and have vital roles in the regulation of cell development, function and epigenetic inheritance. Dysregulation of miRNAs often associates with tumor development and progression. miRNAs can function as both oncogenes or tumor suppressors in different tumors and cell types, which is cell type specific. Therefore, miRNAs can act as a therapeutic target of cancers. However, it needs more work to fully understand the role and mechanism of miRNAs in normal and pathologic conditions as well as to identify target genes of miRNAs involved in NF- κ B signaling pathway.

Acknowledgements

This work was partially supported by the grants from the National Important Science Research Program of China (2011CB933503, 2006CB933205), the National Natural Science Foundation of China (61171030) and the Technology Support Program of Jiangsu (BE2012741).

Conflict of Interests

The Authors declare that they have no conflict of interests

References

- 1) SEN R, BALTIMORE D. Multiple nuclear factors interact with the immunoglobulin enhancer sequences. *Cell* 1986; 46: 705-716.
- 2) MAY MJ, GHOSH S. Rel/NF- κ B and I κ B proteins: an overview. *Seminars in cancer biology*. 8: Elsevier, 1997. p. 63-73.
- 3) GHOSH S, MAY MJ, KOPP EB. NF- κ B and Rel proteins: evolutionarily conserved mediators of immune responses. *Annu Rev Immunol* 1998; 16: 225-260.
- 4) RYSECK R-P, NOVOTNY J, BRAVO R. Characterization of elements determining the dimerization properties of RelB and p50. *Mol Cell Biol* 1995; 15: 3100-3109.
- 5) HAYDEN MS, GHOSH S. Signaling to NF- κ B. *Genes Dev* 2004; 18: 2195-2224.
- 6) PERKINS ND, GILMORE TD. Good cop, bad cop: the different faces of NF-kappaB. *Cell Death Differ* 2006; 13: 759-772.
- 7) SUN SC. Non-canonical NF-kappaB signaling pathway. *Cell Res* 2011; 21: 71-85.
- 8) HAYDEN MS, GHOSH S. NF-kappaB in immunobiology. *Cell Res* 2011; 21: 223-244.
- 9) BONIZZI G, KARIN M. The two NF-kappaB activation pathways and their role in innate and adaptive immunity. *Trends Immunol* 2004; 25: 280-288.
- 10) ZHANG JY, TAO S, KIMMEL R, KHAVARI PA. CDK4 regulation by TNFR1 and JNK is required for NF-kappaB-mediated epidermal growth control. *J Cell Biol* 2005; 168: 561-566.
- 11) BAUD V, KARIN M. Is NF-kappaB a good target for cancer therapy? Hopes and pitfalls. *Nat Rev Drug Discov* 2009; 8: 33-40.
- 12) KARIN M. Nuclear factor-kappaB in cancer development and progression. *Nature* 2006; 441: 431-436.
- 13) INOUE J, GOHDA J, AKIYAMA T, SEMBA K. NF-kappaB activation in development and progression of cancer. *Cancer Sci* 2007; 98: 268-274.
- 14) BHARTI AC, AGGARWAL BB. Chemopreventive Agents Induce Suppression of Nuclear Factor- κ B Leading to Chemosensitization. *Ann N Y Acad Sci* 2002; 973: 392-395.
- 15) KARIN M, GRETEN FR. NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat Rev Immunol* 2005; 5: 749-759.
- 16) HUANG HY, ZHANG ZJ, CAO CB, WANG N, LIU FF, PENG JQ, REN XJ, QIAN J. The TLR4/NF-kappaB signaling pathway mediates the growth of colon cancer. *Eur Rev Med Pharmacol Sci* 2014; 18: 3834-3843.
- 17) WONG D, TEIXEIRA A, OIKONOMOPOULOS S, HUMBURG P, LONE IN, SALIBA D, SIGGERS T, BULYK M, ANGELOV D, DIMITROV S. Extensive characterization of NF- κ B binding uncovers non-canonical motifs and advances the interpretation of genetic functional traits. *Genome Biol* 2011; 12: R70.
- 18) HOESEL B, SCHMID JA. The complexity of NF- κ B signaling in inflammation and cancer. *Mol Cancer* 2013; 12: 1-15.
- 19) OECKINGHAUS A, GHOSH S. The NF- κ B family of transcription factors and its regulation. *Cold Spring Harb Perspect Biol* 2009; 1: a000034.
- 20) LEE Y, KIM M, HAN J, YEOM KH, LEE S, BAEK SH, KIM VN. MicroRNA genes are transcribed by RNA polymerase II. *EMBO J* 2004; 23: 4051-4060.
- 21) CAI X, HAGEDORN CH, CULLEN BR. Human microRNAs are processed from capped, polyadenylated transcripts that can also function as mRNAs. *RNA* 2004; 10: 1957-1966.
- 22) LUND E, GÜTTINGER S, CALADO A, DAHLBERG JE, KUTAY U. Nuclear export of microRNA precursors. *Science* 2004; 303: 95-98.
- 23) BOHNSACK MT, CZAPLINSKI K, GÖRLICH D. Exportin 5 is a RanGTP-dependent dsRNA-binding protein that mediates nuclear export of pre-miRNAs. *RNA* 2004; 10: 185-191.
- 24) CHENDRIMADA TP, GREGORY RI, KUMARASWAMY E, NORMAN J, COOCH N, NISHIKURA K, SHIEKHATTAR R. TRBP recruits the Dicer complex to Ago2 for microRNA processing and gene silencing. *Nature* 2005; 436: 740-744.
- 25) GREGORY RI, CHENDRIMADA TP, COOCH N, SHIEKHATTAR R. Human RISC couples microRNA biogenesis and posttranscriptional gene silencing. *Cell* 2005; 123: 631-640.
- 26) KHVOROVA A, REYNOLDS A, JAYASENA SD. Functional siRNAs and miRNAs exhibit strand bias. *Cell* 2003; 115: 209-216.
- 27) SCHWARZ DS, HUTVÁGNER G, DU T, XU Z, ARONIN N, ZAMORE PD. Asymmetry in the assembly of the RNAi enzyme complex. *Cell* 2003; 115: 199-208.
- 28) MATRANGA C, TOMARI Y, SHIN C, BARTEL DP, ZAMORE PD. Passenger-strand cleavage facilitates assembly of siRNA into Ago2-containing RNAi enzyme complexes. *Cell* 2005; 123: 607-620.
- 29) DOENCH JG, SHARP PA. Specificity of microRNA target selection in translational repression. *Genes Dev* 2004; 18: 504-511.
- 30) LYTLE JR, YARIO TA, STEITZ JA. Target mRNAs are repressed as efficiently by microRNA-binding sites in the 5' UTR as in the 3' UTR. *Proc Natl Acad Sci U S A* 2007; 104: 9667-9672.
- 31) ØROM UA, NIELSEN FC, LUND AH. MicroRNA-10a binds the 5'UTR of ribosomal protein mRNAs and enhances their translation. *Mol Cell* 2008; 30: 460-471.
- 32) HAFNER M, LANDTHALER M, BURGER L, KHORSHID M, HAUSSER J, BERNINGER P, ROTHBALLER A, ASCANO M, JUNGKAMP A-C, MUNSCHAUER M. Transcriptome-wide identification of RNA-binding protein and microRNA target sites by PAR-CLIP. *Cell* 2010; 141: 129-141.
- 33) RIGOUTSOS I. New tricks for animal microRNAs: targeting of amino acid coding regions at conserved and nonconserved sites. *Cancer Res* 2009; 69: 3245-3248.
- 34) KIM VN, HAN J, SIOMI MC. Biogenesis of small RNAs in animals. *Nat Rev Mol Cell Biol* 2009; 10: 126-139.

- 35) FALLER M, GUO F. MicroRNA biogenesis: there's more than one way to skin a cat. *Biochim Biophys Acta* 2008; 1779: 663-667.
- 36) AMBROS V. The functions of animal microRNAs. *Nature* 2004; 431: 350-355.
- 37) BARTEL DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004; 116: 281-297.
- 38) BARTEL DP. MicroRNAs: target recognition and regulatory functions. *Cell* 2009; 136: 215-233.
- 39) LECOLLIER C-H, DUNOYER P, ARAR K, LEHMANN-CHE J, EYQUEM S, HIMBER C, SAÏB A, VOINNET O. A cellular microRNA mediates antiviral defense in human cells. *Science* 2005; 308: 557-560.
- 40) JOPLING CL, YI M, LANCASTER AM, LEMON SM, SARNOV P. Modulation of hepatitis C virus RNA abundance by a liver-specific MicroRNA. *Science* 2005; 309: 1577-1581.
- 41) RUSCA N, MONTICELLI S. MiR-146a in immunity and disease. *Mol Biol Int* 2011; 2011: 437301.
- 42) TAGANOV KD, BOLDIN MP, CHANG K-J, BALTIMORE D. NF- κ B-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. *Proc Natl Acad Sci U S A* 2006; 103: 12481-12486.
- 43) BHAUMIK D, SCOTT G, SCHOKRUPUR S, PATIL C, CAMPISI J, BENZ C. Expression of microRNA-146 suppresses NF- κ B activity with reduction of metastatic potential in breast cancer cells. *Oncogene* 2008; 27: 5643-5647.
- 44) HURST DR, EDMONDS MD, SCOTT GK, BENZ CC, VAIDYA KS, WELCH DR. Breast cancer metastasis suppressor 1 up-regulates miR-146, which suppresses breast cancer metastasis. *Cancer Res* 2009; 69: 1279-1283.
- 45) LUKIW WJ. NF- κ B-regulated micro RNAs (miRNAs) in primary human brain cells. *Exp Neurol* 2012; 235: 484-490.
- 46) LUKIW WJ, ZHAO Y, CUI JG. An NF- κ B-sensitive micro RNA-146a-mediated inflammatory circuit in Alzheimer disease and in stressed human brain cells. *J Biol Chem* 2008; 283: 31315-31322.
- 47) ZAGO M, DE SOUZA AR, HECHT E, ROUSSEAU S, HAMID O, EIDELMAN DH, BAGLOLE CJ. The NF- κ B family member RelB regulates microRNA miR-146a to suppress cigarette smoke-induced COX-2 protein expression in lung fibroblasts. *Toxicol Lett* 2014; 226: 107-116.
- 48) PERRY MM, MOSCHOS SA, WILLIAMS AE, SHEPHERD NJ, LARNER-SVENSSON HM, LINDSAY MA. Rapid changes in microRNA-146a expression negatively regulate the IL-1 β -induced inflammatory response in human lung alveolar epithelial cells. *J Immunol* 2008; 180: 5689-5698.
- 49) SEMPERE LF, FREEMANTLE S, PITHA-ROWE I, MOSS E, DMITROVSKY E, AMBROS V. Expression profiling of mammalian microRNAs uncovers a subset of brain-expressed microRNAs with possible roles in murine and human neuronal differentiation. *Genome Biol* 2004; 5: R13.
- 50) POGUE A, CUI J, LI Y, ZHAO Y, CULICCHIA F, LUKIW W. Micro RNA-125b (miRNA-125b) function in astrogliosis and glial cell proliferation. *Neurosci Lett* 2010; 476: 18-22.
- 51) LUKIW WJ, POGUE AI. Induction of specific micro RNA (miRNA) species by ROS-generating metal sulfates in primary human brain cells. *J Inorg Biochem* 2007; 101: 1265-1269.
- 52) KIM S-W, RAMASAMY K, BOUAMAR H, LIN A-P, JIANG D, AGUIAR RC. MicroRNAs miR-125a and miR-125b constitutively activate the NF- κ B pathway by targeting the tumor necrosis factor alpha-induced protein 3 (TNFAIP3, A20). *Proc Natl Acad Sci USA* 2012; 109: 7865-7870.
- 53) VEREECKE L, BEYAERT R, VAN LOO G. The ubiquitin-editing enzyme A20 (TNFAIP3) is a central regulator of immunopathology. *Trends Immunol* 2009; 30: 383-391.
- 54) ZHOU F, WANG W, XING Y, WANG T, XU X, WANG J. NF- κ B target microRNAs and their target genes in TNF α -stimulated HeLa cells. *Biochim Biophys Acta* 2014; 1839: 344-354.
- 55) TAN G, NIU J, SHI Y, OUYANG H, WU Z-H. NF- κ B-dependent microRNA-125b up-regulation promotes cell survival by targeting p38 α upon ultraviolet radiation. *J Biol Chem* 2012; 287: 33036-33047.
- 56) LIU M, JIANG S, LU Z, YOUNG K, LI Y. Physiological and pathological functions of mammalian microRNAs. *Comprehensive Toxicol* 2010; 2: 427-446.
- 57) NIU J, SHI Y, TAN G, YANG CH, FAN M, PFEFFER LM, WU Z-H. DNA damage induces NF- κ B-dependent microRNA-21 up-regulation and promotes breast cancer cell invasion. *J Biol Chem* 2012; 287: 21783-21795.
- 58) YAN LX, WU QN, ZHANG Y, LI YY, LIAO DZ, HOU JH, FU J, ZENG MS, YUN JP, WU QL. Knockdown of miR-21 in human breast cancer cell lines inhibits proliferation, in vitro migration and in vivo tumor growth. *Breast Cancer Res* 2011; 13: R2.
- 59) LIU Z-L, WANG H, LIU J, WANG Z-X. MicroRNA-21 (miR-21) expression promotes growth, metastasis, and chemo- or radioresistance in non-small cell lung cancer cells by targeting PTEN. *Mol Cell Biochem* 2013; 372: 35-45.
- 60) OU H, LI Y, KANG M. Activation of miR-21 by STAT3 induces proliferation and suppresses apoptosis in nasopharyngeal carcinoma by targeting PTEN gene. *PLoS One* 2014; e109929.
- 61) DARIDO C, GEORGY SR, WILANOWSKI T, DWORKIN S, AUDEN A, ZHAO Q, RANK G, SRIVASTAVA S, FINLAY MJ, PAPPENFUSS AT. Targeting of the tumor suppressor GRHL3 by a miR-21-dependent proto-oncogenic network results in PTEN loss and tumorigenesis. *Cancer Cell* 2011; 20: 635-648.
- 62) GAUR AB, HOLBECK SL, COLBURN NH, ISRAEL MA. Down-regulation of Pcdcd4 by mir-21 facilitates glioblastoma proliferation in vivo. *Neuro Oncol* 2011; 13: 580-590.
- 63) KARAKATSANIS A, PAPAConstantinou I, GAZOULI M, LYBEROPOULOU A, POLYMENEAS G, VOROS D. Expression of microRNAs, miR-21, miR-31, miR-122, miR-

- 145, miR-146a, miR-200c, miR-221, miR-222, and miR-223 in patients with hepatocellular carcinoma or intrahepatic cholangiocarcinoma and its prognostic significance. *Mol Carcinog* 2013; 52: 297-303.
- 64) MA X, KUMAR M, CHOUDHURY SN, BUSCAGLIA LEB, BARKER JR, KANAKAMEDALA K, LIU M-F, LI Y. Loss of the miR-21 allele elevates the expression of its target genes and reduces tumorigenesis. *Proc Natl Acad Sci U S A* 2011; 108: 10144-10149.
- 65) LI B, ZHAO Y, GUO G, LI W, ZHU E, LUO X, MAO X, ZOU Q, YU P, ZUO Q. Plasma microRNAs, miR-223, miR-21 and miR-218, as novel potential biomarkers for gastric cancer detection. *PLoS One* 2012; 7: e41629.
- 66) TOIYAMA Y, TAKAHASHI M, HUR K, NAGASAKA T, TANAKA K, INOUE Y, KUSUNOKI M, BOLAND CR, GOEL A. Serum miR-21 as a diagnostic and prognostic biomarker in colorectal cancer. *J Natl Cancer Inst* 2013; 105: 849-859.
- 67) ZHOU X, REN Y, MOORE L, MEI M, YOU Y, XU P, WANG B, WANG G, JIA Z, PU P. Downregulation of miR-21 inhibits EGFR pathway and suppresses the growth of human glioblastoma cells independent of PTEN status. *Lab Invest* 2010; 90: 144-155.
- 68) SHEEDY FJ, PALSSON-McDERMOTT E, HENNESSY EJ, MARTIN C, O'LEARY JJ, RUAN Q, JOHNSON DS, CHEN Y, O'NEILL LA. Negative regulation of TLR4 via targeting of the proinflammatory tumor suppressor PDCD4 by the microRNA miR-21. *Nat Immunol* 2010; 11: 141-147.
- 69) LU Z, LIU M, STRIBINSKIS V, KLINGE C, RAMOS K, COLBURN N, LI Y. MicroRNA-21 promotes cell transformation by targeting the programmed cell death 4 gene. *Oncogene* 2008; 27: 4373-4379.
- 70) SHIN VY, JIN H, NG EK, CHENG AS, CHONG WW, WONG CY, LEUNG WK, SUNG JJ, CHU KM. NF-kappaB targets miR-16 and miR-21 in gastric cancer: involvement of prostaglandin E receptors. *Carcinogenesis* 2011; 32: 240-245.
- 71) MARQUEZ RT, WENDLANDT E, GALLE CS, KECK K, McCAFFREY AP. MicroRNA-21 is upregulated during the proliferative phase of liver regeneration, targets Pcllino-1, and inhibits NF-kB signaling. *Am J Physiol Gastrointest Liver Physiol* 2010; 298: G535-G541.
- 72) LU Z, LI Y, TAKWI A, LI B, ZHANG J, CONKLIN DJ, YOUNG KH, MARTIN R, LI Y. miR-301a as an NF-kB activator in pancreatic cancer cells. *EMBO J* 2011; 30: 57-67.
- 73) CHEN Z, CHEN LY, DAI HY, WANG P, GAO S, WANG K. miR-301a promotes pancreatic cancer cell proliferation by directly inhibiting bim expression. *J Cell Biochem* 2012; 113: 3229-3235.
- 74) PANDIT H, ZHANG W, LI Y, AGLE S, LI X, LI S, CUI G, MARTIN R. Manganese superoxide dismutase expression is negatively associated with microRNA-301a in human pancreatic ductal adenocarcinoma. *Cancer Gene Ther* 2015: 481-486.
- 75) FANG Y, SUN B, XIANG J, CHEN Z. MiR-301a promotes colorectal cancer cell growth and invasion by directly targeting SOCS6. *Cell Physiol Biochem* 2015; 35: 227-236.
- 76) LIU L, NIE J, CHEN L, DONG G, DU X, WU X, TANG Y, HAN W. The oncogenic role of microRNA-130a/301a/454 in human colorectal cancer via targeting Smad4 expression. *PLoS One* 2013; 8: e55532.
- 77) MA X, YAN F, DENG Q, LI F, LU Z, LIU M, WANG L, CONKLIN DJ, McCracken J, SRIVASTAVA S. Modulation of tumorigenesis by the pro-inflammatory microRNA miR-301a in mouse models of lung cancer and colorectal cancer. *Cell Discovery* 2015; 1: 1.
- 78) WANG M, LI C, YU B, SU L, LI J, JU J, YU Y, GU Q, ZHU Z, LIU B. Overexpressed miR-301a promotes cell proliferation and invasion by targeting RUNX3 in gastric cancer. *J Gastroenterol* 2013; 48: 1023-1033.
- 79) XU XD, HE XJ, TAO HQ, ZHANG W, WANG YY, YE ZY, ZHAO ZS. Abnormal expression of miR-301a in gastric cancer associated with progression and poor prognosis. *J Surg Oncol* 2013; 108: 197-202.
- 80) ZHOU P, JIANG W, WU L, CHANG R, WU K, WANG Z. miR-301a is a candidate oncogene that targets the homeobox gene Gax in human hepatocellular carcinoma. *Dig Dis Sci* 2012; 57: 1171-1180.
- 81) XIE H, LI L, ZHU G, DANG Q, MA Z, HE D, CHANG L, SONG W, CHANG H, KROLEWSKI J. Infiltrated pre-adipocytes increase prostate cancer metastasis via modulation of the miR-301a/androgen receptor (AR)/TGF- β 1/Smad/MMP9 signals. *Oncotarget* 2015; 6: 12326-12339.
- 82) MYCKO MP, CICHALEWSKA M, MACHLANSKA A, CWIKLINSKA H, MARIASIEWICZ M, SELMAJ KW. MicroRNA-301a regulation of a T-helper 17 immune response controls autoimmune demyelination. *Proc Natl Acad Sci U S A* 2012; 109: E1248-E1257.
- 83) MA F, ZHANG J, ZHONG L, WANG L, LIU Y, WANG Y, PENG L, GUO B. Upregulated microRNA-301a in breast cancer promotes tumor metastasis by targeting PTEN and activating Wnt/ β -catenin signaling. *Gene* 2014; 535: 191-197.
- 84) DOU L, WANG S, SUI X, MENG X, SHEN T, HUANG X, GUO J, FANG W, MAN Y, XI J. MiR-301a mediates the effect of IL-6 on the AKT/GSK pathway and hepatic glycogenesis by regulating PTEN expression. *Cell Physiol Biochem* 2015; 35: 1413-1424.
- 85) ICHIKAWA T, SATO F, TERASAWA K, TSUCHIYA S, TOI M, TSUJIMOTO G, SHIMIZU K. Trastuzumab produces therapeutic actions by upregulating miR-26a and miR-30b in breast cancer cells. *PLoS One* 2012; 7: e31422.
- 86) ZHU E-D, LI N, LI B-S, LI W, ZHANG W-J, MAO X-H, GUO G, ZOU Q-M, XIAO B. miR-30b, Down-regulated in gastric cancer, promotes apoptosis and suppresses tumor growth by targeting plasminogen activator inhibitor-1. *PLoS One* 2014; 9: e106049.
- 87) LIAO WT, YE YP, ZHANG NJ, LI TT, WANG SY, CUI YM, QI L, WU P, JIAO HL, XIE YJ. MicroRNA-30b functions as a tumour suppressor in human colorectal cancer by targeting KRAS, PIK3CD and BCL2. *J Pathol* 2014; 232: 415-427.

- 88) LI L, WANG B. Overexpression of microRNA-30b improves adenovirus-mediated p53 cancer gene therapy for laryngeal carcinoma. *Int J Mol Sci* 2014; 15: 19729-19740.
- 89) QUINTAVALLE C, DONNARUMMA E, IABONI M, ROSCIGNO G, GAROFALO M, ROMANO G, FIORE D, DE MARINIS P, CROCE C, CONDORELLI G. Effect of miR-21 and miR-30b/c on TRAIL-induced apoptosis in glioma cells. *Oncogene* 2013; 32: 4001-4008.
- 90) GAZIEL-SOVRAN A, SEGURA MF, DI MICCO R, COLLINS MK, HANNIFORD D, DE MIERA EV-S, RAKUS JF, DANKERT JF, SHANG S, KERBEL RS. miR-30b/30d regulation of GalNAc transferases enhances invasion and immunosuppression during metastasis. *Cancer Cell* 2011; 20: 104-118.
- 91) MORENO-MATEOS MA, BARRAGÁN V, TORRES B, RODRÍGUEZ-MATEO C, MÉNDEZ-VIDAL C, BEREZIKOV E, MUDDULURU G, ALLGAYER H, PINTOR-TORO JA. Novel small RNA expression libraries uncover hsa-miR-30b and hsa-miR-30c as important factors in anoikis resistance. *RNA* 2013; 19: 1711-1725.
- 92) WEI C, LI L, GUPTA S. NF- κ B-mediated miR-30b regulation in cardiomyocytes cell death by targeting Bcl-2. *Mol Cell Biochem* 2014; 387: 135-141.
- 93) MELLIOS N, GALDZICKA M, GINNS E, BAKER SP, ROGAEV E, XU J, AKBARIAN S. GENDER-SPECIFIC REDUCTION OF ESTROGEN-SENSITIVE small RNA, miR-30b, in subjects with schizophrenia. *Schizophr Bull* 2012; 38: 433-443.
- 94) BRIDGE G, MONTEIRO R, HENDERSON S, EMUSS V, LAGOS D, GEORGIOPOULOU D, PATIENT R, BOSHOFF C. The microRNA-30 family targets DLL4 to modulate endothelial cell behavior during angiogenesis. *Blood* 2012; 120: 5063-5072.
- 95) NAOVI AR, FORDHAM JB, NARES S. miR-24, miR-30b, and miR-142-3p regulate phagocytosis in myeloid inflammatory cells. *J Immunol* 2015; 194: 1916-1927.
- 96) MUDDULURU G, CEPPI P, KUMARSWAMY R, SCAGLIOTTI G, PAPOTTI M, ALLGAYER H. Regulation of Axl receptor tyrosine kinase expression by miR-34a and miR-199a/b in solid cancer. *Oncogene* 2011; 30: 2888-2899.
- 97) TROPAN K, WENZL K, PICHLER M, PURSCHE B, SCHWARZENBACHER D, FEICHTINGER J, THALLINGER GG, BEHAM-SCHMID C, NEUMEISTER P, DEUTSCH A. miR-199a and miR-497 are associated with better overall survival due to increased chemosensitivity in diffuse large B-cell lymphoma patients. *Int J Mol Sci* 2015; 16: 18077-18095.
- 98) TSUKIGI M, BILIM V, YUUKI K, UGOLKOV A, NAITO S, NAGAOKA A, KATO T, MOTOYAMA T, TOMITA Y. Re-expression of miR-199a suppresses renal cancer cell proliferation and survival by targeting GSK-3 β . *Cancer Lett* 2012; 315: 189-197.
- 99) HE J, XU Q, JING Y, AGANI F, QIAN X, CARPENTER R, LI Q, WANG XR, PEIPER SS, LU Z. Reactive oxygen species regulate ERBB2 and ERBB3 expression via miR-199a/125b and DNA methylation. *EMBO reports* 2012; 13: 1116-1122.
- 100) WANG Z, MA X, CAI Q, WANG X, YU B, CAI Q, ZHU Z, LI C. MiR-199a-3p promotes gastric cancer progression by targeting ZHX1. *FEBS Lett* 2014; 588: 4504-4512.
- 101) HAN Y, KUANG Y, XUE X, GUO X, LI P, WANG X, GUO X, YUAN B, ZHI Q, ZHAO H. NLK, a novel target of miR-199a-3p, functions as a tumor suppressor in colorectal cancer. *Biomed Pharmacother* 2014; 68: 497-505.
- 102) CALLEGARI E, ELAMIN BK, D'ABUNDO L, FALZONI S, DONVITO G, MOSHIRI F, MILAZZO M, ALTAVILLA G, GIACOMELLI L, FORNARI F. Anti-tumor activity of a miR-199-dependent oncolytic adenovirus. *PLoS One* 2013; 8: e73964.
- 103) MURAKAMI Y, TOYODA H, TANAKA M, KURODA M, HARADA Y, MATSUDA F, TAJIMA A, KOSAKA N, OCHIYA T, SHIMOTOHNO K. The progression of liver fibrosis is related with overexpression of the miR-199 and 200 families. *PLoS One* 2011; 6: e16081.
- 104) WANG J, HE Q, HAN C, GU H, JIN L, LI Q, MEI Y, WU M. p53-FACILITATED miR-199A-3P REGULATES SOMATIC CELL reprogramming. *Stem Cells* 2012; 30: 1405-1413.
- 105) ZHOU R, HU G, LIU J, GONG AY, DRESCHER KM, CHEN X-M. NF-KAPPA B p65-DEPENDENT TRANSACTIVATION OF miRNA genes following *Cryptosporidium parvum* infection stimulates epithelial cell immune responses. *PLoS Pathog* 2009; 5: e1000681.
- 106) ZHAO Y, BHATTACHARJEE S, JONES BM, DUA P, ALEXANDROV PN, HILL JM, LUKIW WJ. REGULATION OF TREM2 EXPRESSION BY AN NF- κ B-sensitive miRNA-34a. *Neuroreport* 2013; 24: 318.
- 107) LI J, WANG K, CHEN X, MENG H, SONG M, WANG Y, XU X, BAI Y. TRANSCRIPTIONAL ACTIVATION OF MICRORNA-34A BY NF-kappa B in human esophageal cancer cells. *BMC Mol Biol* 2012; 13: 4.
- 108) LIANG M, YAO G, YIN M, LÜ M, TIAN H, LIU L, LIAN J, HUANG X, SUN F. Transcriptional cooperation between p53 and NF- κ B p65 regulates microRNA-224 transcription in mouse ovarian granulosa cells. *Mol Cell Endocrinol* 2013; 370: 119-129.
- 109) LI H, LI Y, LIU D, SUN H, LIU J. miR-224 is critical for celastrol-induced inhibition of migration and invasion of hepatocellular carcinoma cells. *Cell Physiol Biochem* 2013; 32: 448-458.
- 110) KUMAR V, PALERMO R, TALORA C, CAMPESE A, CHECQUOLO S, BELLAVIA D, TOTTONE L, TESTA G, MIELE E, INDRACOLO S. NOTCH AND NF- κ B SIGNALING PATHWAYS REGULATE miR-223/FBXW7 axis in T-cell acute lymphoblastic leukemia. *Leukemia* 2014: 2324-2335.
- 111) ZHANG X, LIU S, HU T, LIU S, HE Y, SUN S. Up-regulated microRNA-143 transcribed by nuclear factor kappa B enhances hepatocarcinoma metastasis by repressing fibronectin expression. *Hepatology* 2009; 50: 490-499.
- 112) MA J, LIU J, WANG Z, GU X, FAN Y, ZHANG W, XU L, ZHANG J, CAI D. NF-kappaB-dependent microRNA-425 upregulation promotes gastric cancer cell growth by targeting PTEN upon IL-1 β induction. *Mol Cancer* 2014; 13: 40.

- 113) WANG J, GU Z, NI P, QIAO Y, CHEN C, LIU X, LIN J, CHEN N, FAN Q. NF-kappaB P50/P65 hetero-dimer mediates differential regulation of CD166/ALCAM expression via interaction with microRNA-9 after serum deprivation, providing evidence for a novel negative auto-regulatory loop. *Nucleic Acids Res* 2011; gkr302.
- 114) WEN F, YANG Y, JIN D, SUN J, YU X, YANG Z. MiR-NA-145 is involved in the development of resistin-induced insulin resistance in HepG2 cells. *Biochem Biophys Res Commun* 2014; 445: 517-523.
- 115) VENTO-TORMO R, RODRÍGUEZ-UBREVA J, DI LISIO L, ISLAM AB, URQUIZA JM, HERNANDO H, LÓPEZ-BIGAS N, SHANNON-LOWE C, MARTÍNEZ N, MONTES-MORENO S. NF-κB directly mediates epigenetic deregulation of common microRNAs in Epstein-Barr virus-mediated transformation of B-cells and in lymphomas. *Nucleic Acids Res* 2014; gku826.
- 116) BAO JL, LIN L. MiR-155 and miR-148a reduce cardiac injury by inhibiting NF-kappaB pathway during acute viral myocarditis. *Eur Rev Med Pharmacol Sci* 2014; 18: 2349-2356.