Rutin prevents the ovariectomy-induced osteoporosis in rats

Q.-L. WANG¹, X.-C. HUO², J.-H. WANG³, D.-P. WANG¹, Q.-L. ZHU⁴, B. LIU⁵, L.-L. XU⁶

Oiuling Wang and Xuechen Huo contributed equally to this work

Abstract. – OBJECTIVE: Our study aimed to explore the anti-osteoporotic effect of rutin (quercetin-3-O-rutinose) on ovariectomized female rats.

MATERIALS AND METHODS: An ovariectomized (OVX) rat model of osteoporosis was employed to evaluate the anti-osteoporotic potency of rutin. One week after surgery, the rats were administered intragastrically with rutin or saline once daily respectively for 3 months. Bone mineral density (BMD) was measured by dual-energy x-ray absorptiometry (DEXA). The bone microstructure was analyzed by hematoxylin and eosin (HE) staining of the left tibia histomorphology. Estradiol, IL-6, and TNF-α were measured by ELISA kits.

RESULTS: The results showed that rutin significantly improved the bone mineral density (BMD) and increased the level of inflammatory factor of IL-6, TNF- α , and INF- γ in OVX rats. Rutin turned bone trabecula to be thickened and dense, and kept regular array. Moreover, rutin significantly improved the average thickness of trabecular bone and the average bone volume fraction.

CONCLUSIONS: Rutin possessed with significant anti-osteoporotic activity, which can be considered as an idealistic anti-osteoporotic candidate for human osteoporosis diseases.

Key Words

Rutin, Osteoporosis, Ovariectomized Rats, IL-6, TNF-α.

Introduction

Osteoporosis is a kind of metabolic disorder characterized by the imbalance between osteoblastic formation and osteoclastic resorption, resulting in skeletal fragility and susceptibility to fractures 1,2. It is a stealthy and unpredictable disease, which become symptomatic until the advanced stages³. Ovarian hormone deficiency is a major risk factor for osteoporosis in postmenopausal women. It was predicted that, by 2050, more than a half of worldwide osteoporotic hip fractures would occur in Asia4. Some pharmacological treatments have antiosteoporotic effect, such as hormone therapy (HT), bisphosphonate and selective estrogen receptor modulators (SERMs)⁵. However, these medications were exposed some side effects, including gastrointestinal tolerance problems in bisphosphonate and the potential malignancies in HT^{6,7}. Therefore, natural compounds with antiosteoporotic activity and few side effects are worth of exploring. Many studies⁸⁻¹⁴ have been shown that some vegetables and fruits such as onions, beans and tea can inhibit bone resorption. They are strong antioxidants, which can help to prevent cardiovascular disease by inhibiting oxidation of low-density lipoproteins¹⁵. Onions is rich in flavonol glycosides, and the antioxidant effect is considered the strongest in vegetables¹⁶. Several independent evidence indicated that some particular natural products containing polyphenol molecules possible may be useful for these beneficial treatments. For example, the protective action against osteopenia or osteoporosis was discovered in catechin¹⁷, rutin¹⁸, green tea

¹Department of Endocrinology, The Affiliated Yantai Yuhuangding Hospital of Qingdao University, Yantai, Shandong, China

²Department of Hepatobiliary Surgery, The Affiliated Yantai Yuhuangding Hospital of Qingdao University, Yantai, Shandong, China

³Department of Health Care for Cadres, The Affiliated Yantai Yuhuangding Hospital of Qingdao University, Yantai, Shandong, China

⁴Department of Pharmacy, The Affiliated Yantai Yuhuangding Hospital of Qingdao University, Yantai, Shandong, China

⁵Department of Vascular Surgery, The Affiliated Hospital of Qingdao University, Qingdao, Shandong, China

⁶Department of Endocrinology and Metabolism, The Affiliated Hospital of Qingdao University, Qingdao, Shandong, China

polyphenol¹⁹, olive oil polyphenol²⁰ and apple polyphenol. Flavonoids represent a large number of phenolic compounds that commonly found in daily nutrition with proven health benefits as well as antiosteoporotic properties²¹. Flavonoids have been used widely implicated in the alleviation of postmenopausal osteoporosis. Studies have been shown that isoflavonoids (such as genistein and daidze) and flavonols (such as quercetin) in the form of rutin can prevent bone loss in ovariectomized (OVX) rats. Quercetin is the primary flavonol, which mainly found as a glycoside (rutin also called quercetin-3-O-rutinose). It binds to estrogen receptors and influences the development of cell lines from several hormone-dependentcancers^{22,23}. In foods, quercetin occurs mainly as its glycoside, rutin. Like quercetin, rutin displays antioxidative properties and inhibits the growth of cancer cell lines^{24,25}. In the present study, we studied the effect of rutin on bone metabolism in the ovariectomized (OVX) rats.

Materials and Methods

Chemicals

Rutin was purchased from the National Institute for the Control of Pharmaceuticals and Biological Products (Beijing, China) and its purity was determined to be $\geq 98\%$ by HPLC measurement. Unless otherwise stated, all other reagents were of analytical grade.

Animal Groups and Experiments

Fifty female Sprague-Dawley (SD) rats aged 8 months (purchased from Slacom, Shanghai, China, 340-350 g) were housed under controlled conditions including a room temperature of $22 \pm 2^{\circ}$ C with a 12 h light/dark cycle and allowed free access to food and distilled water. One week later, the acclimatized rats were anesthetized with chloral hydrate (300 mg/kg, i.p.) (Sinopharm, Shanghai, China) under aseptic conditions. Ten rats underwent bilateral laparotomy without removing the ovaries as Sham control group, the others were underwent bilateral ovariectomy and equally randomized into three groups: intragastric administered vehicle as model group (OVX), OVX with rutin (R) of graded doses (R5, n = 10, 5 mg/kg body weight/day and R10, n = 10, 10 mg/kg body weight/day) according to preliminary experiments. The administration of rutin lasted for 3 months. The uterus wet weight and uterus index of rats were recorded weekly during the experimental period. The procedures of the animal study, including the raising, feeding, and the whole surgical process, followed the APS's "Guiding Principles on the Care and Use of Animals" and were approved by the Committee of Animal Study at Shandong University.

Enzyme-Linked Immunosorbent Assay (ELISA)

After the completion of treatments, the animals were sacrificed under deep anesthesia with chloral hydrate (300 mg/kg, i.p.) and bloodletting via arteria femoralis. Blood samples were collected and centrifuged (4°C) at 3000 r/min for 15 min. The upper serum was separated immediately and was stored at -20°C for future analysis. According to the manufacturer's instructions, ELISA assay was performed to determine serum levels of osteocalcin, estrogen (E2), 1, 25-dihydroxycholecalciferol (1, 25(OH)₂D₃) and inflammatory cytokines (IL-6, TNF-α, INF-γ) with ELISA kits (BioSource International, Camarillo, CA, USA).

Bone Histomorphometric Analysis

Sections of un-decalcified femurs fixed in 10% neutral buffered formalin and embedded in paraffin wax were taken using a microtome and stained with Goldner's trichrome. The sections were mounted and observed for histopathological changes. Histomorphology of the left femur was analyzed for bone microstructure after hematoxylin and eosin (HE) staining (Beijing Solar Bioscience Technology Co., Ltd, Beijing, China). HE staining was performed according to the previous studies of Huang et al²⁶. The histomorphometric study of the metaphysis of the proximal tibiae was performed as previously described with image analysis software (Image Pro Plus 6.1 for Windows; Media Cybernetics, Silver Spring, MD, USA)²⁷.

Bone Mineral Density

The total bone mineral density (BMD) of the right femur was measured using Lunar Prodigy Advance by dual-energy X-ray absorptiometry (GE Healthcare, Pittsburgh, PA, USA) equipped with appropriate software for bone density assessment in small laboratory animals as reported elsewhere²⁸. Distal and proximal regions corresponded to the cancellous bone and central to the cortical bone²⁹. Results are given in g/cm².

Statistical Analysis

These data were presented as means ± standard deviation (SD). Statistical comparisons among all groups were analyzed by using one-way analysis

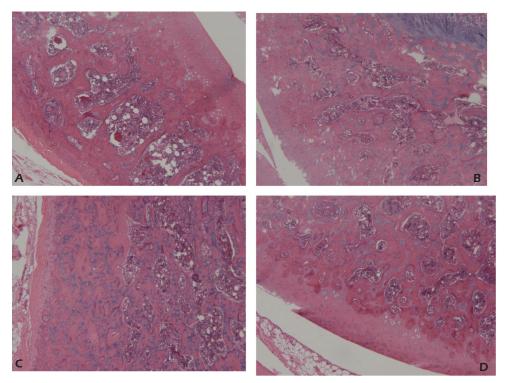


Figure 1. Body weight gain during the treatment period. The body weight of OVX group was significantly higher than other groups. The interventions of rutin did not significantly affect body weight gain throughout the period of experiment. Data were presented as means \pm SD. Sham: sham control group; OVX: the ovariectomized rats group; R5: OVX rats with rutin at a dose of 5 mg/kg body weight/day; R10: OVX rats with rutin at a dose of 10 mg/kg body weight/day.

of variance (ANOVA). When significant levels (p<0.05) were revealed, pair-wise comparisons between groups were made using the Fisher's least significant difference (LSD) method. Statistical analysis software, SPSS 11.5 version (SPSS Inc., Chicago, IL, USA) was adopted for processing the data of the present study.

Results

Body Weight

No rats died during the study period in all groups. Five groups of rats had a similar initial mean body weight. During the 3-month-experimental period, body weights of rats in all groups were weekly monitored to study whether rutin altered body weight following ovariectomy. After operation, body weight gain was typically and consistently higher in the ovariectomized rats in each group compared with the Sham control group (p<0.05) (Figure 1). The body weight of OVX group was significantly higher than Sham control group (p<0.05) on week 4 after surgery. Except for the slightly lower body weight gain of

the R10 (rutin in high dose) group, the interventions we used, R5 groups, did not significantly affect body weight gain throughout the period of the experiment.

Index of Uterus

The variation tendency of uterine index was similar to body weight. OVX caused significant atrophy of uterine tissue comparison with Sham control group (p<0.05). Rutin significantly increased the uterine weight comparison with OVX group (p<0.05), which appeared to be dose dependent (Table I).

Table I. Index of uterus during the treatment period.

Group	Weight (mg)	Index of uterus (mg/g)
Sham	326.8±16.4	2.15±0.75
OVX	81.6±6.3*	0.99±0.83*
R10	168.8±12.9*#	1.85±0.75*#
R5	159.4±13.2*#	1.69±0.94*#

Data were presented as means \pm SD. Power, power values calculated using α =0.05; *p<0.05 vs. Sham; *p<0.05 vs. OVX.

Table II. The level of cytokines IL-6, IFN- γ and TNF- α .

Group	IL-6 (pg/ml)	TNF-α (pg/ml)	INF-γ (pg/ml)	
Sham	369.72±38.82	38.92±17.95	0.948 ± 0.135	
OVX	583.82±41.73*	104.74±27.84*	2.582±0.852*	
R5	482.83±28.89**	82.84±21.37*	1.862±0.957**	
R10	427.95±38.48*#	63.95±24.53 [#]	1.264±0.729#	

Data were presented as means \pm SD. Power, power values calculated using α =0.05; *p<0.05 vs. Sham; *p<0.05 vs. OVX.

Table III. The level of E_2 , OCN, $1,25(OH)_2D_3$ in serum.

Group	E ₂ (ng/l)	OCN (ng/ml)	1,25(OH) ₂ D ₃ (ng/ml)
Sham	369.72±38.82	38.92±17.95	0.948 ± 0.135
OVX	583.82±41.73*	104.74±27.84*	2.582±0.852*
R5	482.83±28.89**	82.84±21.37*	1.862±0.957**
R10	427.95±38.48*#	63.95±24.53 [#]	1.264±0.729#

Data were presented as means \pm SD. Power, power values calculated using α =0.05; *p<0.05 vs. Sham; *p<0.05 vs. OVX.

Effect of Rutin on Inflammatory Markers

ELISA was performed to evaluate serum cytokines level in different treatment groups.

As shown in Table II, the cytokines IL-6, IFN- γ and TNF- α were significantly elevated in OVX group compared to Sham control group (p<0.05). However, intervention with rutin reversed the upregulated levels of cytokine IL-6, IFN- γ and TNF- α on OVX rats in a dose-dependent manner.

Serum Parameters

Results of serum parameters in different groups of rats were shown in Table III. The results indicated a significant reduction in the levels of serum E_2 and 1, $25(OH)_2D_3$ in OVX rats compared with Sham control group, while intervention with rutin the level of E_2 and 1, $25(OH)_2D_3$ significantly increased. In contrast, statistically significant (p<0.05) elevated levels of serum OCN were observed in OVX rats when compared with Sham control group. Interestingly, rutin treatment reversed the upregulated serum OCN levels on OVX rats in a dose-dependent manner.

Table IV. Bone mineral density of the femur.

Group	BMD (g/cm²)
Sham OVX	0.251±0.007 0.148±0.004 *
R10	0.198±0.008 *#
R5	0.176±0.005 *#

Data were presented as means \pm SD. Power, power values calculated using α =0.05; *p<0.05 vs. Sham; *p<0.05 vs. OVX.

Bone Mineral Density of the Femur

OVX significantly decreased the right femur BMD comparison with the Sham control group (p<0.05). The administration of rutin to the OVX rats significantly recovered the BMD of femur. The 3-month treatment with rutin at higher doses increased the right femur BMD compared to the OVX group (p<0.05), which appeared to be dose dependent (Table IV).

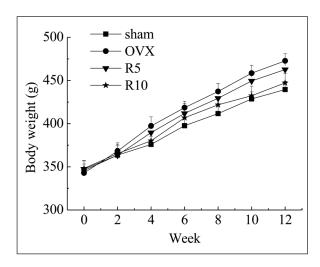


Figure 2. Histological sections of femur in rats of different groups (magnification 200×). The ovariectomized rat showed sparse, uniform thinning of the trabeculae resulting in the wideness of trabecular interspace, but the administration of rutin significantly protected bone from ovariectomy-induced osteopenia. Sham: sham control group; OVX: the ovariectomized rats group; R5: OVX rats with rutin at a dose of 5 mg/kg body weight/day; R10: OVX rats with rutin at a dose of 10 mg/kg body weight/day.

Rutin Decreased Trabecular Bone Loss

The bone architecture of the femur was analyzed with Harris hematoxylin and eosin. The rats of the Sham control group revealed normal compactness of the diaphysis and competent trabeculae (Figure 2A). The OVX rats showed sparse, uniform thinning of the trabeculae resulting in the wideness of trabecular interspace (Figure 2B). The low (R5) dosage rutin groups exhibited both restorative progress with mineralization and quite well-distributed osteocytes. Uniform trabeculae with dense matrix and shaft size were observed (Figure 2C). The group treated with high dose rutin (R10) showed nearly complete recovery with essential features of normal bone and complete formation of trabeculae, which was similar to the Sham control group (Figure 2D). These results demonstrated there were beneficial effects of rutin on the prevention of bone loss, which induced by ovariectomy.

Discussion

Osteoporosis is a kind of metabolic disorder characterized by the imbalance between osteoblastic formation and osteoclastic resorption, resulting in skeletal fragility and susceptibility to fractures. Ovarian hormone deficiency is a major risk factor for osteoporosis in postmenopausal women. It is well known that estrogen deficiencies are important risk factors in the pathogenesis of osteoporosis. Ovariectomy results in a dramatic decrease in uterine weight, bone mineral density, and biomechanical strength, and these changes are in part due to estrogen deficiency. Some studies have shown that several vegetables and fruits such as onions, beans and tea can inhibit bone resorption. Tea consumption was associated with a high bone mineral density in older women³⁰, which was assumed to be caused by catechins that counteracted the adverse bone metabolism effects of caffeine. Bone resorption inhibition occurred in rats on an onion-rich diet (the main source of quercetin in many diets)³⁰. Being the glycosylated form of quercetin, rutin was often found in onions as main flavonols³¹. Onion extracts have been shown to inhibit bone resorption in vitro and vivo³². The present study was designed to evaluate the effect of rutin on the protection against ovariectomy-induced osteopenia in rats. The data showed that OVX decreased body and uterine weight compared

with the sham control group. Administration of rutin increased both the body and uterine weight in the ovariectomized rat. In the histopathology examination, the OVX rats showed sparse, uniform thinning of the trabeculae resulting in the wideness of trabecular interspace. The administration of rutin exhibited restorative progress with mineralization along with quite well-distributed osteocytes. Histopathology examination of the femurs of rutin groups revealed ossification, mineralization, and calcified cartilaginous deposits, all of which indicate marked restorative action, thus suggesting that the protective action of rutin may be due to an increase in bone formation with a reduction in bone resorption. It is well-known that estradiol (E2), osteocalcin (OCN), calcium and phosphorus are widely accepted phenotype markers for bone formation³³. A previous study¹⁸ showed that rutin consumption was more efficient in restoring bone mineral loss in OVX rats than isofiavones, decreased a bone resorption marker (deoxypyridinoline) after urinary excretion, and increased an osteoblastic activity marker (osteocalcin). In the present study, the results indicated a significant reduction in the levels of serum E, and 1,25(OH),D, in OVX rats when compared with Sham control group, while intervention with rutin significantly increased the level of E₂ and 1, 25(OH)₂D₃. Interestingly, rutin treatment reversed the upregulated serum OCN levels on OVX rats in a dose-dependent manner. Considerable evidence has demonstrated that pro-inflammatory cytokines, such as interleukin-1 (IL-1), interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α), are involved in the regulation of bone turnover, by increasing bone resorption^{34,35}. Ralston et al³⁶ demonstrated that interleukin-1 (IL-1), tumor necrosis factor (TNF), and IL-6 mRNAs are expressed more frequently in bone cells from untreated postmenopausal women than from those on estrogen replacement therapy. Those cytokines are also involved in the release of free radicals^{37,38}. This pathway will, in turn, inhibit osteoblastic recruitment and the activity of mature cells³⁹ and increase osteoclastic resorption. In the present study, intervention with rutin reversed the upregulated levels of cytokine IL-6, IFN- γ and TNF- α on OVX rats in a dose-dependent manner. Our work demonstrated the usefulness and beneficial effects of rutin on the prevention of bone loss induced by ovariectomy. OVX significantly decreased the right femur BMD compared to the Sham control group, while the administration of rutin to the OVX rats significantly recovered the BMD of femur in a dose-dependent manner. Gaumet et al⁴⁰ discovered that an increase in fecal and urinary calcium excretions, as well as a decrease in calcium absorption efficiency, might contribute to the reduction of BMD. Rutin could counteract the ovariectomy-induced osteopenia in rats in a dose-dependent manner.

Conclusions

The present study clearly demonstrates that rutin contributes to the prevention of bone loss and deterioration of trabecular microarchitecture induced by ovariectomization in rats. Furthermore, the results indicate that rutin is a potential alternative therapeutic agent for treatment of postmenopausal osteoporosis.

Conflict of interest statement

We declare that we have no conflict of interest.

References

- 1) GARNERO P, DELMAS PD. Osteoporosis. Endocrinol Metab Clin North Am 1997; 26: 913-936.
- HANS D, FUERST T, LANG T, MAJUMDAR S, Lu Y, GENANT HK, GLÜER C. How can we measure bone quality? Baillieres Clin Rheumatol 1997; 11: 495-515.
- Kanis JA, Melton LJ, Christiansen C, Johnston CC, Khaltaev N. The diagnosis of osteoporosis. J Bone Miner Res 1994; 9: 1137-1141.
- GULLBERG B, JOHNELL O, KANIS JA. World-wide projections for hip fracture. Osteoporos Int 1997; 7: 407-413.
- GOLTZMAN D. Discoveries, drugs and skeletal disorders. Nat Rev Drug Discov 2002; 1: 784-796.
- Reid IR. Pharmacotherapy of osteoporosis in postmenopausal women: focus on safety. Expert Opin Drug Saf 2002; 1: 93-107.
- YEH IT. Postmenopausal hormone replacement therapy: endometrial and breast effects. Adv Anat Pathol 2007; 14: 17-24.
- MÜHLBAUER RC, LI F. Effects of vegetables on bone metabolism. Nature 1999; 401: 343-344.
- CHEN YM, Ho SC, Woo JL. Greater fruit and vegetable intake is associated with increased bone mass among postmenopausal Chinese women. Br J Nutr 2006; 96: 745-751.
- 10) New SA, Robins SP, Campbell MK, Martin JC, Garton MJ, Bolton-Smith C. Dietary influences on bone mass and bone metabolism: further evidence of a positive link between fruit and vegetable consumption and bone health? Am J Clin Nutr 2000; 71: 142-151.
- 11) MIYAMOTO K, HOSOI Y. Dietary patterns associated with bone mineral density in premenopausal

- Japanese farmwomen. Am J Clin Nutr 2006; 83: 1185-1192.
- 12) PRYNNE CJ, MISHRA GD, O'CONNELL MA, MUNIZ G, LASKEY MA, YAN L. Fruit and vegetable intakes and bone mineral status: a cross sectional study in 5 age and sex cohorts. Am J Clin Nutr 2006; 83: 1420-1428.
- 13) TUCKER KL, CHEN H, HANNAN MT, CUPPLES LA, WILSON PW, FELSON D. Bone mineral density and dietary patterns in older adults: the framingham osteoporosis study. Am J Clin Nutr 2002; 76: 245-252.
- 14) VATANPARAST H, BAXTER-JONES A, FAULKNER RA, BAILEY DA, WHITING SJ. Positive effects of vegetable and fruit consumption and calcium intake on bone mineral accrual in boys during growth from child-hood to adolescence: the University of Saskatchewan Pediatric Bone Mineral Accrual Study. Am J Clin Nutr 2005; 82: 700-706.
- Duthie G, Crozier A. Plant-derived phenolic antioxidants. Curr Opin Clin Nutr Metab Care 2000; 3: 447-451.
- 16) JUSTESEN U, KNUTHSEN P, LETH T. Quantitative analysis of flavonols, flavones and flavanones in fruits, vegetables and beverages by high performance liquid chromatography with photo-diode array and mass spectrometric detection. J Chromatogr A 1998; 799: 101-110.
- 17) Delaisse JM, Eeckhout Y, Vaes G. Inhibition of bone resorption in culture by (+)-catechin. Biochem Pharmacol 1986; 35: 3091-3094.
- HORCAJADA-MOLTENI MN, CRESPY V, COXAM V, DAVICCO MJ, REMESY C, BARLET JP. Rutin inhibits ovariectomy-induced osteopenia in rats. J Bone Miner Res 2000; 15: 2251-2258.
- 19) PARK YH, HAN DW, SUH H, RYU GH, HYON SH, CHO BK, PARK JC. Protective effects of green tea polyphenol against reactive oxygen species-induced oxidative stress in cultured rat calvarial osteoblast. Cell Biol Toxicol 2003; 19: 325-337.
- 20) PUEL C, QUNTIN A, AGALIAS A, MATHEY J, OBLED C, MAZUR A, DAVICCO MJ, LEBECQUE P, SKALTSOUNIS AL, COXAM V. Olive oil and its main phenolic micronutrient (oleuropein) prevent inflammation-induced bone loss in the ovarietomised rat. Br J Nutr 2004; 92: 119-127.
- 21) LÉOTOING L, DAVICCO MJ, LEBECQUE P, WITTRANT Y, COX-AM V. The flavonoid fisetin promotes osteoblasts differentiation through Runx2 transcriptional activity. Mol Nutr Food Res 2014; 58: 1156-1159.
- 22) Monje P, Zanello S, Holick M, Boland R. Differential cellular localization of estrogen receptor alpha in uterine and mammary cells. Mol Cell Endocrinol 2001; 181: 117-129.
- Ross JA, Kasum CM. Dietary flavonoids: bioavailability, metabolic effects, and safety. Annu Rev Nutr 2002; 22: 19-34.
- 24) CHEN YC, SHEN SC, LEE WR, HOU WC, YANG LL, LEE TJ. Inhibition of nitric oxide synthase inhibitors and lipopolysaccharide induced inducible NOS and cyclooxygenase-2 gene expressions by rutin, quercetin, and quercetin pentaacetate in RAW 264.7 macrophages. J Cell Biochem 2001; 82: 537-548.
- 25) Wang S, De Groff VL, CLINTON SK. Tomato and soy polyphenols reduce insulin-like growth factor-I-stimulated rat prostate cancer cell pro-

- liferation and apoptotic resistance in vitro via inhibition of intracellular signaling pathways involving tyrosine kinase. J Nutr 2003; 133: 2367-2376.
- 26) HUANG TH, YANG RS, HSIEH SS, LIU SH. Effects of caffeine and exercise on the development of bone: a densitometric and histomorphometric study in young Wistar rats. Bone 2002; 30: 293-299.
- 27) Parfitt AM, Drezner MK, Glorieux FH, Kanis JA, Malluche H, Meunier PJ. Bone histomorphometry: standardization of nomenclature, symbols, and units. Report of the ASBMR Histomorphometry Nomenclature Committee. J Bone Miner Res 1987; 2: 595-610.
- 28) ARJMANDI BH, LUCAS EA, JUMA S, SOLIMAN A, STOECKER BJ, KHALIL DA, SMITH BJ, WANG C. Dried plums prevent ovariectomy-induced bone loss in rats. JANA 2001; 4: 50-56.
- 29) PASTOUREAU P, CHOMEL A, BONNET J. Specific evaluation of localized bone mass and bone loss in the rat using dual-energy X-ray absorptiometry subregional analysis. Osteoporos Int 1995; 5: 143-149.
- SCALBERT A, MANACH C, MORAND C, REMESY C, JIMENEZ L. Dietary polyphenols and the prevention of diseases. Crit Rev Food Sci Nutr 2005; 45: 287-306.
- 31) PRICE KR, RHODES MJC. Analysis of the major flavonol glycosides present in four varieties of onion and changes in composition resulting from autolysis. J Sci Food Agric 1997; 74: 331-339.
- 32) MÜHLBAUER RC, LI F, GUENTHER HL. Common vegetables consumed by humana potently modulate bone metabolism in vitro and in vivo. Bone 1998; 23: S388.
- 33) EVANS DB, BUNNING RAD, RUSSELL RGG. The effects of recombinant human interleukin-1b on cellular proliferation and the production of prostaglandin

- E2, plasminogen activator, osteocalcin and alkaline phospha-tase by osteoblast-like cells derived from human bone. Biochem Biophys Res Commun 1990; 166: 208-216.
- 34) PFEILSCHIFTER J, CHENU C, BIRD A, MUNDY GR, ROOD-MAN GD. Interleukin-1 and tumor necrosis factor stimulate the formation of human osteoclast differentiation of human osteoclast-like cells in vitro. J Bone Miner Res 1989; 4: 113-118.
- 35) PACIFI R. Cytokines and osteoclast activity. Calcif Tissue Int 1995; 56: S27-28.
- 36) RALSTON SH. Analysis of gene expression in human bone biopsies by polymerase chain reaction: evidence for enhanced cytokine expression in postmenopausal osteoporosis. J Bone Miner Res 1994; 9: 883-890.
- 37) DAS UN. Interaction(s) between essential fatty acids, eicosanoids, cytokines, growth factors and free radicals: relevance to new therapeutic strategies in rheumatoid arthritis and other collagen vascular diseases. Prostaglandins Leukot Essent Fatty Acids 1991; 44: 201-210.
- 38) Wang XM, Zhang YG, Li AL, Long ZH, Wang D, Li XX, Xia JH, Luo SY, Shan YH. Relationship between levels of inflammatory cytokines in the peripheral blood and the severity of depression and anxiety in patients with Parkinson's disease. Eur Rev Med Pharmacol Sci 2016; 20: 3853-3856.
- 39) Mody N, Parhami F, Sarafian TA, Demer L. Oxidative stress modulates osteoblastic differenciation of vascular and bone cells. Free Rad Biol Med 2001; 31: 509-519
- 40) GAUMET N, SEIBEL MJ, COXAM V, DAVICCO MJ, LEBECQUE P, BARLET JP. Influence of ovariectomy and estradiol treatment on calcium homeostasis during aging in rats. Arch Physiol Biochem 1997; 105: 435-444.