Clinical tools to assess nutritional risk and malnutrition in hospitalizaed children and adolescents

E. RINNINELLA¹, A. RUGGIERO², P. MAURIZI², S. TRIARICO², M. CINTONI¹, M.C. MELE¹

¹Area Gastroenterologia, Nutrition Team, Fondazione Policlinico Universitario "A. Gemelli", Catholic University of the Sacred Heart, School of Medicine, Rome, Italy

Abstract. – Malnutrition in children and adolescents may be underestimated during hospital stay. In western countries, children were often hospitalized for acute or chronic diseases that are not necessarily related to malnutrition. However, acute or chronic injuries may hamper nutritional status, prolonging recovery after admission and consequently length of hospital stay.

Several methods and techniques are known to investigate malnutrition in children, even if their use is not widespread in clinical practice. Many of these are simple and easy to perform and could be useful to a better management of every kind of illness.

In this review, we will focus on clinical tools necessary to reveal a nutritional risk at admission and to assess nutritional status in hospitalized children and adolescents.

Key Words

Nutritional risk screening tools, Malnutrition assessment, Hospitalized children.

Introduction

An adequate nutritional state plays a crucial role in normal growth, treatment response, comorbidities, quality of life, cost of care and long-term survival among pediatric hospitalized patients with clinical conditions¹. In 2013, the Academy of Nutrition and Dietetics and American Society of Parenteral and Enteral Nutrition (ASPEN) defined pediatric malnutrition as "an imbalance between nutrient requirements and intake, resulting in cumulative deficits of energy, protein or micronutrients that may negatively affect growth, development and other relevant outcomes"².

In pediatric patients, the illness-related malnutrition is a dynamic and multifactorial process sustained by several factors such as inflammation, nutrient losses, increased energy expenditure, decreased nutrient intake or utilization. These conditions may be related to acute (trauma, burns, infections) or chronic diseases (cancer, chronic kidney diseases, cystic fibrosis, heart failure, inflammatory bowel diseases, neurological and neuromuscular diseases, etc.)³⁻⁸.

Pediatric illness-related malnutrition is yet an undervalued issue, even if abnormalities of nutritional state may produce significant morbidity and mortality among pediatric patients and several studies have reported a prevalence of 6%-51% of this condition among hospitalized children^{9,10}.

Consequently, an accurate screening of nutritional risk and an appropriate assessment of the nutritional state may be crucial for the clinical management of these patients. In hospitalized children, a prompt nutritional intervention on body composition is useful to reverse linear growth arrest, promote tolerance to therapeutic regimens, improve the quality of life and reduce the length of hospital stay (LOS)^{11,12}.

The European Society for Clinical Nutrition and Metabolism (ESPEN) and the European Society for Pediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) recommend nutritional risk screening for hospitalized children during admission, to facilitate the detection of children nutritionally at risk and to allow the physician to make an appropriate nutritional support plan¹³. Even if several pediatric nutritional risk scores are reported in literature, there is not consensus on the "ideal" screening tool and, often, nutritional screening is not yet widely performed¹⁴.

²Area Salute del Bambino, Pediatric Oncology, Fondazione Policlinico Universitario "A. Gemelli", Catholic University of the Sacred Heart, School of Medicine, Rome, Italy

A complete nutritional assessment requires at least five steps^{2,15}:

- Medical and dietary history;
- Detailed physical examination;
- Biochemical parameters;
- Accurate anthropometric measurements (weight, height, weight for height [WFH], head circumference, body mass index [BMI], mid-upper-arm circumference [MUAC], triceps skinfold [TSF] thickness);
- Body composition measurements.

Besides, body composition analysis requires the evaluation of fat mass (FM), fat-free mass (FFM) and body cell mass (BCM)¹⁶. Several methods are available for this purpose¹⁷⁻¹⁹.

In this review, we describe nutritional tools reported in scientific literature for the screening and the diagnostic assessment of nutritional status among hospitalized children and adolescents.

Nutritional Risk Screening Tools

According to ESPEN statements, nutritional risk screening tools have been designed to detect protein and energy undernutrition and/or predict if undernutrition may develop or worsen²⁰. An appropriate nutritional screening tool may inform about patient's nutritional status and its relation to patient's illness. By definition, a nutritional screening tool should be not time-consuming, simple, easily comprehensible, sensitive and specific, applicable and reliable for a wide disease group and in the daily practice²¹.

To date, in our knowledge, there are seven main nutritional risk screening tools available for children^{22,23} as listed in Table I.

Firstly, in 1995 Reilly et al²⁴ proposed the *Nutritional Risk Score (NRS)*, based on four items: Body Mass Index (BMI), weight loss in the last 3 months, dietary intake in the last week, severity of the disease. NRS is simple to use and applicable to all medical and surgical patient categories and ages, for assessing risk of undernutrition at admission to the hospital and for identifying need of nutritional intervention

The *Pediatric Nutrition Risk Score (PNRS)*, proposed in 2000 by Sermet-Geydelus et al²⁵ analyses three items: patient's medical condition (valuated from 1 to 3 points according to the presence of mild, moderate or severe disease), presence of pain (1 point if pain is present), reduction of food intake (1 point if food intake is <50%). A score ≥ 3 indicates that the patient is at

high risk of malnutrition and must be referred to a nutrition team. This method is rather quick, but it does not identify the nutritional status of the patient^{23,25}.

Secker and Jeejeebhoy²⁶ proposed the Subjective Global Nutritional Assessment (SGNA) in surgical patients between 1 month and 18 years of age within 30 days after a surgical intervention. Anthropometric measurements (length or height, weight, percentage of ideal body weight for height, body mass index-for-age, MUAC, TSF, mid-arm muscle area, handgrip strength), biochemical investigations (concentrations of serum albumin, transferrin, and hemoglobin and total lymphocyte count), parental height, dietary intake, GI symptoms, functional status of the patient and physical exam were assessed. Patients are divided into three groups: well nourished, moderately malnourished, severely malnourished. So classified, malnourished children had higher rates of infectious complications and a longer post-operative LOS than well-nourished children^{23,26}. This method allows a nutritional evaluation of hospitalized children who may be at risk of malnutrition, although it is lengthy and time-consuming²⁷.

McCarthy et al²⁸ validated the *Screening Tool* for the Assessment of Malnutrition in Pediatrics (STAMP) in a study performed in the United Kingdom among medical and surgical patients between 2 and 17 years of age. The score evaluates patient's clinical diagnosis, nutritional intake during hospitalization and anthropometric measurements, developing a care plan based on the child's overall malnutrition risk (low, medium or high). STAMP is reported to have a high specificity (90%) and sensitivity (72%) in identifying malnutrition risk^{23,27}.

Gerasimides et al²⁹ adopted the Pediatric Yorkhill Malnutrition Score (PYMS) in the United Kingdom among medical and surgical patients between 1 and 16 years of age. The PYMS assesses four items: BMI, history of recent weight loss, changes in nutritional intake and the expected effect of current medical condition on patient's nutritional status. The total score reflects the degree of the patient's nutritional risk. PYMS reported a moderate sensitivity (59%), a high specificity (92%) and fewer false-positive cases than STAMP score^{23,27}.

In a multicenter study conducted in Netherland among medical and surgical patients between 1 month and 18 years, Hulst et al³⁰ proposed the *Screening Tool for Impaired Nutritional Status and Growth (STRONGkids)*.

Table I. Main nutritional risk screening tools for hospitalized children.

				4 Main pri	nciples item	4 Main principles items according to ESPEN ²⁰	to ESPEN ²⁰	Other items	items
Tools	Authors	Population	Age	Current nutritional status	Weight	Reduced	Disease	Anthropometric measurements	Signs and symptoms
NRS ²⁴	Reilly et al, 1995	Medical	0-17 years	×	×	×	×	×	
PNRS ²⁵	Sermet-Geydelus Medical and et al, 2000 surgical	Medical and surgical	>1 month- 18 years	×		×	×		Pain
SGNA ²⁶	Secker et al, 2007 Surgical	Surgical	>1 month- 18 years	×	×	×	×	×	GI symptoms, functional capacity, parental height
$STAMP^{28}$	McCarthyet al, 2012	Medical and surgical	2-18 years	×		×	×	×	
$ m PYMS^{29}$	Gerasimidis et al, 2010	Medical and surgical, 1-16 years except cardiologic, renal, orthopedic conditions	1-16 years	×	×	×	×	×	
STRONG kids ³⁰	Hulst et al, 2010	Medical and surgical	>1 month-x 18 years	×	×	×			
PNST ³⁵	White et al, 2016	White et al, 2016 Medical and surgical 0-16 years	0-16 years	×	×	×			

NRS: Nutritional Risk Score; PNRS: Pediatric Nutritional Risk Score; SGNA: Surgical Global Nutritional Assessment; STAMP: Subjective Global Nutritional Assessment; PYMS: Pediatric Nutrition Score; STRONGkids: Screening Tool for Risk of Impaired Nutritional Status and Growth; PNST: Pediatric Nutrition Score; Screening Tool.

It consists of four items: subjective clinical assessment, high-risk diseases, nutritional intake and losses, weight loss or poor weight gain. Patients classified at high nutritional risk have a longer hospitalization and a negative standard deviation score (SDS) for weight-for-height (WFH), which indicated a state of acute malnutrition. This tool appeared rapid and easy-to-use (it needs a mean of three minutes); additionally it may predict LOS and identifies a need for nutritional interventions during the hospitalization. Huysentruyt et al³¹ in a large Belgian population of hospitalized children, demonstrated the reproducibility of STRONGkids: a good correlation between STRONGkids score and the patient's current nutritional status (defined by WFH and SDS) was found, thus identifying patients needing a nutritional intervention during hospitalization.

Joosten et al²¹ analyzed the six nutritional screening tools mentioned above, concluding that *STRONGkids* may be considered the quickest, reliable and practical to use, compared to others. In fact, it can be performed at the admission by every health care professional; it is based on a subjective clinical assessment without anthropometric measurements or additional items.

Conversely, in a multicenter study performed in 12 Italian hospitals, Spagnuolo et al³² pointed out that notwithstanding its feasibility and sensitivity, STRONGkids is not specific. Consequently, it may be used as a very preliminary screening tool to be integrated with other clinical data.

Moeeni et al³³ compared the use of *STAMP*, *PYMS*, and *STRONGkids* for assessing nutritional risk among 150 Iranian hospitalized children. They demonstrated that *STRONGkids* correlates better than the others with the anthropometric measurements and with LOS. The same group showed similar results in a pediatric population of New Zealand³⁴. On the other hand, Wonoputri et al²⁷ recommended *PYMS* as the most reliable screening tool in hospitalized children in Indonesia.

Recently, White et al³⁵ proposed the *Pediatric Nutrition Screening Tool (PNST)*, based on 4 simple nutrition screening questions: involuntary weight loss in recent days, poor weight gain in the last few months, reduction in food intake in the last few weeks and presence of obesity.

Nutritional risk is assessed by the presence of almost two positive answers to the above-mentioned questions. *PNST* may provide a sensitive, valid, and simpler alternative to existing pediatric nutrition screening tools such as *STAMP*, *STRON-Gkids*, and *PYMS*.

Even if several studies have been conducted for evaluating these various nutritional screening tools, at present, there is not yet a consensus on any tool as in the adults.

In 2015, Huysentruyt et al³⁶ performed a meta-analysis, including 11 studies comprehending at least one score among *PNRS*, *STAMP*, *PYMS*, and *STRONGkids*. Authors concluded that it is not advisable to prefer one single nutritional screening tool because each screening category should be linked to a specific setting. For example, *STRONGkids* may be the best option as a quick tool for testing risk in all age groups, whereas *PYMS* or STAMP may be preferred if anthropometric measures are needed at hospital admission and during the screening process.

Nutritional Global Assessment

After the nutritional risk assessment, necessary for all hospitalized patients, a percentage of patients may result at risk of malnutrition. Hence, a specific nutritional assessment is mandatory, as explained below.

Medical and Dietary History

The checklist in the Table II allows to collect an accurate medical and dietary history, necessary for a global nutritional assessment.

The examination of medical history should include the growth history, the eventual onset of puberty and the psychomotor development with feeding abilities. Previous acute and chronic illness, hospitalization and surgical procedures should be investigated, with emphasis on nutrition-related illness¹⁵.

It is important to establish the duration of the current disease, documenting oral motor skills and swallow ability, GI symptoms, weight changes. The use of certain drugs, which may cause nutritional deficiency, may be documented, as well as the use of any vitamin, mineral or herbal supplement, for their possible interactions with drugs³⁷.

The detection of dietary history provides information about the child's dietary patterns, the number of meals, food allergies and intolerances, self-imposed and prescribed diets. The dietary food records can be retrospective (usually a 24-hours diet recall) or prospective (usually for three to seven days)³⁸.

Moreover, food frequency questionnaires give information on the amount and frequency of spe-

Table II. Checklist of medical and dietary history.

Patient's personal and medical history

- Growth history
- Onset of puberty
- Psycomotory development
- Acute or chronic illness
- Hospitalization, surgical procedures

About current disease

- Duration of the current disease
- Oral motor skills, swallow safety
- GI symptoms
- Weight changes
- Use of drugs, vitamin, mineral or herbal supplements

About dietary patterns

- Dietary patterns
- Number of meals
- Food allergies and intolerances
- Self-imposed and prescribed diets
- Food frequency questionnaire (FFQ)

cific dietary patterns, providing an insight into the relation between diet and disease. In their Eating Assessment in Toddlers study, Mills et al³⁹ demonstrated the validity and the high reproducibility of the Food Frequency Questionnaire (FFQ), for identifying dietary patterns in children.

Physical Examination

The physical examination assesses patient's general conditions and investigates on the presence of signs of specific nutritional deficiencies¹⁵.

In particular, abnormal findings in the examination of hair, eyes, lips and mouth, tongue, teeth, skin, nails usually are related to specific micronutrient deficiency (for example zinc, iron, essential fatty acid, selenium, magnesium, vitamin A, C, B12, folate)⁴⁰.

A visual inspection may reveal protein-energy malnutrition in the presence of extremity edema, distended abdomen, muscle wasting. The adequacy of fat stores may be assessed by the inspection and palpation of orbital, triceps, bony, iliac crest prominence and depressions between ribs. Furthermore, the palpation of muscles overlying the clavicle, scapular area, shoulders, quadriceps and calves may allow information about the adequacy of muscle stores: in a well-nourished patient, the muscles appear rounded and well-developed^{41,42}.

Biochemical Parameters

Laboratory data play a complementary role in the assessment of nutritional status, even if no one lab test can give a comprehensive assessment of nutritional status.

Illness-related malnutrition is often associated with an inflammatory status that promotes a

catabolic effect on free fat body mass and muscle protein. The presence of inflammation should be established, because it may decrease the effectiveness of nutritional intervention. Acute phase proteins (C-reactive protein, fibrinogen, haptoglobin, ceruloplasmin, ferritin, and alpha-1-antitripsin) levels are high during an acute inflammation or a catabolic state; conversely, albumin, prealbumin, retinol binding protein (RBP) and transferrin are decreased in these cases⁴³. However, the magnitude of a positive acute phase response may be attenuated in "protein energy wasting" (PEW), a process characterized by a first phase of impaired nutrient intake and absorption, followed by a second phase of depletion of body stores, with alteration of biochemical and physiologic functions^{44,45}.

Anthropometric Measurements

Weight is a measure of overall nutritional status, but it may be influenced by many variables, such as age, sex, daily intakes and hydration status. It is important to remove all excess clothing, measuring weight in light or no clothing and without diaper for infants. Furthermore, the scales should be calibrated monthly, using objects with known weight. Patients older than 2 years of age and able to stand should be weighted on a platform scale with movable weights or with digital scales. Weights are recorded in kilograms and rounded to the nearest 100 grams. For patients unable to stand, bed scales or wheelchair scales may be used as alternative measures. Children under 2 years of age should be weighted placed supine in a pan scale, making sure the weight is scattered equally on each side of the center of the scale. Weights are recorded in kilograms and rounded to the nearest 10 grams⁴⁶.

Stature (length, height or alternative height measures) is a very important measure for observing long-term nutritional status. For children under 2 years of age recumbent length is obtained using an infantometer (a solid length board) in the supine position. This measurement requires two individuals: one who holds the child's head straight on the board and a second that extends the child's legs and feet flattened and moves a perpendicular moveable plate against the child's feet. For children older than 2 years of age height is found using a vertical stadiometer, if possible fixed on the wall, with a perpendicular arm moved down to the crown of the head, removing shoes and putting head, shoulders, hells, buttock against the flat surface⁴⁶.

For patients with limitations (such as contractures, hip dysplasia, hypertonicity, and inability to

stand) alternative height measures may be found: for details, we suggest *ad hoc* references⁴⁷⁻⁵⁰.

Head Circumference (HC) may obtained in children until they reach 36 months of age, using a flexible measuring tape placed around the head across the frontal bones, above the eyebrows and the right and left ears, over the occipital prominence at the back of the head. HC should be considered an index of brain development and nutritional status, correlated with undernutrition⁵¹.

Weight for length may be evaluated in patients under 2 years of age to detect a state of overweight or underweight. This calculation corresponds to BMI, used in patients older than 2 years. BMI, also known as "Quetelet's index", is calculated with weight in kilograms divided by height in meters squared [BMI=Weight (kg)/Height² (m²). Given the variability in sex and age, different specific BMI values on growth charts are available in children¹⁵.

BMI has been used to assess obesity since the 1960s in adult⁵²; more recently the International BMI cut offs in pediatric overweight and obesity have been developed, based respectively on the adult cut offs of 25 and 30^{53,54} kg/m².

Despite World Health Organization (WHO) expert committee validated the use of BMI also for assessing thinness in adolescence⁵⁵, BMI is not recommended to assess indistinctly underweight or wasting in adolescents or children: in children in fact, underweight is expressed by "low weight for age", whereas wasting is indicated by "low weight for height"⁵⁶. Furthermore, BMI should not be used as the only indicator of nutritional status in children with clinical conditions, because BMI does not consider differences in the composition of the body.

Mid-Upper-Arm Circumference (MUAC), also known as Mid-Arm Circumference (MAC) is a simple measure taken by a flexible tape placed perpendicular to the long axis of the arm, which is flexed at 90° angle. The midpoint of the upper arm halfway between the acromion and the olecranon is measured and marked. Then, with the patient's arm relaxed at the side, the tape is placed around the previously marked midpoint⁴⁷. The MUAC is a better indicator of body composition than BMI in those patients with edema or fluid shifts, because it is not influenced by hydration status. In a study among children at high risk of malnutrition in rural Bangladesh, Roy et al⁵⁷ suggested that MUAC may be a potential anthropometric indicator of nutrition in children aged between 6 and 60 months.

Triceps skinfold thickness (TSF) is measured using a skinfold caliper on the right arm at the point marked previously for the MUAC on the back of the arm. The examiner grasps the skin and subcutaneous fat tissue between thumb and forefinger above the point previously marked. After the skin, where the skinfold caliper is placed at the midpoint marked, maintaining a grasp of the skinfold. TSF is commonly adopted for research setting, but it can be also useful for identifying patient's body fat stores.

Once MUAC and TSF are obtained, it is possible to calculate *Mid-Arm-Muscle Circumference (MAMC)*, *Arm Muscle Area (AMA)* and *Arm Fat Area (AFA)*, which are useful to distinguish muscle from fat stores. *MAMC* may be calculated from MUAC and TSF using the formula:

MAMC (cm) = MUAC (cm) – (TSF (cm)
$$\times \pi$$
)

AMA derives from the formula:

AMA (cm²) = [MUAC (cm)
$$-\pi \times TSF$$
 (cm)]² / 4π .

For calculating AFA, it is necessary to obtain *Total Arm Area (TAA)* with the formula:

TAA (cm²) = MUAC (cm)² /
$$4\pi$$

Finally, AFA derives from the formula⁴⁵:

$$AFA (cm)^2 = TAA (cm)^2 - AMA (cm)^2$$

Handgrip test is a strength measure performed using a handheld dynamometer, which is a non-invasive and low-cost instrument for measuring muscle functional status. Using the dynamometer, the patient performs a sequence of movements that reproduce the maximum strength of the hand and forearm muscles. In adult cohorts it has been evaluated as a sensitive marker of energy intake⁵⁸ and bone mineral density⁵⁹, while a low handgrip strength is associated with a poor prognosis in cardiovascular and cancer diseases^{60,61}. Nutritional changes affect earlier the muscle function than the muscle mass, consequently handgrip strength may help to prompt detect the presence of malnutrition in children. There are some efforts in parameterizing this test^{62,63}, but appropriate age and gender-specific reference ranges must be used⁶⁴.

Percentiles for age and sex traditionally express the position of a child's measurement (weight, length or height, weight for length or BMI) on a bell-shaped standard reference curve, derived from population data.

A percentile indicate the percentage of population that stay above or below that measured in the child, helping to compare the child's position to a population of other children similar for age and sex. However, according to WHO statements, percentiles do not indicate precisely the actual degree of patient's deviation from population standards; conversely, the use of Z scores would be better for expressing anthropometric measures⁶⁵. Z scores are more sensitive than percentiles, because they express in standard deviation (SD) how far from the mean the child is, comparing the individual anthropometric measurement with data from reference age groups. Z scores are available in chart form for several anthropometric measurements (such as weight, height, BMI, head circumference, MUAC, TSFT). Online calculators such as "Peditools.org" are actually available for automatically calculating Z scores.

In their study Green Corkins et al⁴¹ derived the degree of malnutrition from the Z score of weight for height, BMI for age, MUAC: -1 to -1.9 Z scores assess mild malnutrition, -2 to -2.9 Z scores assess moderate malnutrition, below -3 Z scores assess severe malnutrition (Table III).

Growth charts collect the patient's anthropometric measurements, allowing an assessment of the growth over time and facilitating clinicians in an early identification of a faltering growth⁶⁶. It is recommended to use the 2006 WHO charts as normative standards for term infants and children up to 2 years of age and the 2000 Center for Disease Control and Prevention (CDC) charts for children and adolescents from 2 to 20 years of age. These charts are available for both female and male, allowing the assessment of percentiles and Z score for several anthropometric measurements (weight for age, height or length for age, head circumference for age, BMI for age)^{67,68}.

Furthermore, several growth charts are currently available for different disease and syndromes (such as prematurity, Down syndrome, Turner syndrome, Prader-Willi syndrome, Noonan syndrome, achondroplasia, cerebral palsy, Duchenne muscular dystrophy)⁶⁹.

Indicators of the Body Composition

Body composition measurements can predict clinical outcomes and nutritional status in children and adolescent. Body composition measurements are really demanding in children and adolescents, because of the growth-related changes in height, weight, fat-free mass (FFM), total body water (TBW) and total extracellular tissue.

Numerous techniques exist for routine determination of body composition, including total body potassium counting (TBK), dual-energy X-ray absorptiometry (DXA), single and multifrequency bioelectrical impedance analysis (BIA). Although these reference methods are used routinely, each one has intrinsic practical limitations⁷⁰.

The detection of nutritional status in children with clinical conditions requires the measurement of both fat mass (FM) and fat free mass (FFM). Body cell mass (BCM) is the metabolically active component of FFM and expresses the functional cellular component of the body. BCM may be an ideal indicator of nutritional status in children with clinical condition, because it is independent of hydration changes that occur with disease. BCM is calculated from total body potassium counting (TBK), using the formula of Wang et al⁷¹:

$$BCM$$
 (kg): (TBK (g) \times 9.18) / 39.1

Murphy et al^{72,73} demonstrated that BCM measurement by TBK might be a valid indicator of nutritional status in children with cancer, because it is independent of extracellular fluid changes produced by the disease. However, TBK measurements may not be widely available, so alternative simple methods such as DXA and BIA can provide measures on FM and FFM⁷⁴.

Dual-energy X-ray Absorptiometry (DXA) is a noninvasive method that can be applied at all ages for the measure of regional body composition. DXA may allow the determination of three main body components (bone mineral, bone-free FFM, and fat mass) with low-radiation exposure, short scanning time and low cost⁷⁵. DXA is accepted for the analysis of body composition and for the measure of adipose tissue mass in pediatric population⁷⁶.

Table III. Z scores ranges in nutritional assessment.

	Mild malnutrition	Moderate malnutrition	Severe Malnutrition
Weight for height Z score	-1 to -1.9	-2 to -2.9	-3 or greater
BMI for age Z score	-1 to -1.9	-2 to -2.9	-3 or greater
MUAC Z score	-1 to -1.9	-2 to -2.9	-3 or greater

§adapted from Green Corkins K.et al41

BMI: Body Mass Index; MUAC: Mid-Upper Arm Circumference

Bioelectrical impedance analysis (BIA) is another safe, non-invasive and manageable method generally used for the indirect determination of body composition⁷⁷. BIA method is based on the principle that the conduction of an alternate electric current in a body may find a resistance to the passage (impedance) inversely proportional to the contents of water and electrolytes. In this concept, legs and arms are theoretically comparable to cylindrical conductors in which FFM (made of well hydrated cells) offers a relative slow impedance, while FM (poor in water and electrolytes) opposes an high impedance. The bones, air (in the lungs) and parenchymal organs are not considered good conductors and are not taken into account. The impedance (Z) at the passage of current trough the body consists of two components: resistance (R) and reactance (Xc). R depends essentially on the extracellular water (ECW) and FM. Xc is an indirect measure of body cell mass: it is the quality of healthy cell membranes of taking an electric load and liberate it in a second moment, after a brief delay. This is a capacitance-like property, similar to that of vessels or condensers in electrical circuits. There are two kind of measure in BIA methods: the single frequence (SF-BIA) and the multi-frequencies (MF-BIA). At single frequency (often 50 kHz), Z is given principally by R, since the only resistance is offered by ECW. In MF-BIA, current could pass at higher frequencies (100-200 kHz), recruiting during the passage many functioning cells, whose contents (water and electrolytes) enhance its conduction: the results is a lower resistance and a higher reactance (Figure 1).

SF-BIA is commonly used to estimate total body water (TBW) and fat free mass (FFM), con-

versely MF-BIA allows the advantage of a differentiation between intracellular and extracellular TBW. Nevertheless, SF-BIA is more validated in children, because only recently MF-BIA devices have been marketable⁷⁸.

Tyrrel at al⁷⁹ demonstrated that BIA performs better than anthropometric indices in the estimation of fat-free mass (FFM), fat mass (FM) and percentage body fat (PBF) in children.

BIA measures total water content of body (TBW) not directly, but through prediction equations for calculating total body water (TBW) and FFM as a function of impedance, weight, height, sex and age. BIA equations to estimate TBW, FFM or FM are based on adult proportions and they may be less accurate in children, because the hydration fraction changes during childhood and adolescence. Consequently, pre and post pubertal age, gender, ethnic differences and changes in hydration must be take into consideration, when validating separate BIA equations for children^{80,81}.

The use of Phase angle (PhA) is remarkable in a clinical setting, it reflects body cell mass (BCM), and it is also one of the best indicators of cell membrane function⁸². PhA is a derived measure of BIA method, calculated from R and Xc with the following formula:

PhA = arctan (Xc/R)
$$\times$$
 (180/ π)

PA is an indirect measure but it is proportional to body cell mass and its value depends on tissue health and age. In healthy adults the mean value is around 5.6, with lower values in females and older subjects ⁸³⁻⁸⁵. It has been recognized as a measure of

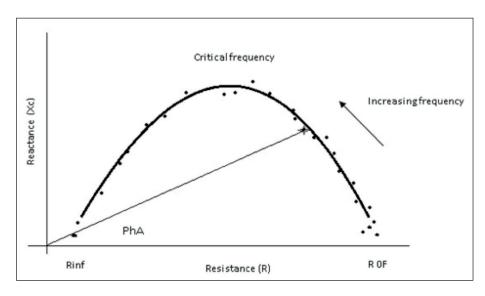


Figure 1. Relationship between Resistance (R) and Reactance (Xc): at increasing frequencies, Xc increases, R decreases. Rinf: Resistance at infinite frequency; R 0F: Resistance at zero frequency; PhA: Phase Angle. Modified from ESPEN textbook: Basics in clinical nutrition, fourth edition⁷⁸.

nutritional status both in adults and in hospitalized children^{58,86} and a prognostic factor of survival in adults affected by cancer⁸⁷⁻⁸⁹. Farias et al⁹⁰ showed that PhA may be a prognostic and nutritional status indicator for children and adolescents undergoing hematopoietic stem cell transplantation (HSCT). On the same way, Pileggi et al⁹¹ demonstrated that PhA may be a good and sensitive method for identifying nutritional risk at hospital admission and monitoring nutritional status of children during hospitalization. In this cross-sectional study, PhA in children was around 5 in healthy control subjects (with some difference due to age and sex) with significantly lower values in hospitalized patient. Actually, PhA is strongly recommended by ESPEN as a prognostic nutritional measure⁹².

Edefonti et al ^{93,94} assessed the prevalence of malnutrition in children on chronic peritoneal dialysis, using the anthropometry-biompedance analysis nutrition (ABN) score. This score uses six parameters based on anthropometry BIA values. The sum of each score gives a result ranging from 10.33 to 15.00 in healthy children, and below 10.33 in malnourished children. This method appears to be non-invasive, reliable, and easy to measure both in ill and healthy children. Moreover, it was used in a special cohort of patient, affected by protein energy malnutrition. In the future, several studies may be necessary to validate this method in other similar clinical context.

Conclusions

Hospitalized children should be firstly assessed for nutritional risk. In this setting no a defined tool is suitable for every situation, even if STRONGkids score appear to be the most quick, reliable and practical to use since from the admission in the hospital. When a high risk of malnutrition is found in a hospitalized child or adolescent, a nutritional global assessment must be performed by a pediatrician or a clinical nutritionist. For this purpose, medical and dietary history, physical examination and anthropometric measurements are well accepted and validated methods. Among body composition analysis procedures, BIA represents a non-invasive, safe and easily performed tool with an increasing number of supporting studies. Given the availability of so many resources in modern clinical settings, every effort should be carried out to early identify and promptly correct malnutrition among hospitalized children.

Conflict of interest

The Authors declare that they have no conflict of interests.

References

- Murphy AJ, Hill RJ, Buntain H, White M, Brookes D, Davies PS. Nutritional status of children with clinical conditions. Clin Nutr 2017; 36: 788-792.
- MEHTA NM, CORKINS MR, LYMAN B, MALO.NE A, GODAY PS, CARNEY LN, MONCZKA JL, PLOGSTED SW, SCHWENK WF. Defining pediatric malnutrition: a paradigm shift toward etiology-related definitions. JPEN J Parenter Enteral Nutr 2013; 37: 460-481.
- LAI HJ. Classification of nutritional status in cystic fibrosis. Curr Opin Pulm Med 2006; 12: 422-427.
- KIM S, KOH H. Nutritional aspect of pediatric inflammatory bowel disease: its clinical importance. Korean J Pediatr 2015; 58: 363-368.
- QUITADAMO P, THAPAR N, STAIANO A, BORRELLI O. Gastrointestinal and nutritional problems in neurologically impaired children. Eur J Paediatr Neurol 2016; 20: 810-815.
- Verger J. Nutrition in the pediatric population in the intensive care unit. Crit Care Nurs Clin North Am 2014; 26: 199-215.
- FURTH SL. Growth and nutrition in children with chronic kidney disease. Adv Chronic Kidney Dis 2005; 12: 366-371.
- 8) AYOUB D, LOPETUSO LR, CHAMSEDDINE F, DAJANI A, LAHIRI K, MAHMOUD H, MIODADY MS, ZIRIZZOTTI G, SULTAN MA, FRANCESCHI F, GASBARRINI A. Epidemiological evaluation of acute gastroenteritis and therapeutic approaches in Middle East Countries. Eur Rev Med Pharmacol Sci 2016; 20: 3891-3901.
- BEER SS, JUAREZ MD, VEGA MW, CANADA NL. Pediatric malnutrition: putting the new definition and standards into practice. Nutr Clin Pract 2015; 30: 609-624.
- JOOSTEN KF, HULST JM. Prevalence of malnutrition in pediatric hospital patients. Curr Opin Pediatr 2008; 20: 590-596.
- HARTMAN C, SHAMIR R, HECHT C, KOLETZKO B. Malnutrition screening tools for hospitalized children. Curr Opin Clin Nutr Metab Care 2012; 15: 303-309.
- MOTIL KJ. Sensitive measures of nutritional status in children in hospital and in the field. Int J Cancer Suppl 1998; 11: 2-9.
- 13) AGOSTONI C, AXELSON I, COLOMB V, GOULET O, KOLETZKO B, MICHAELSEN KF, PUNTIS JW, RIGO J, SHAMIR R, SZAJEW-SKA H, TURCK D; ESPGHAN Committee on Nutrition; European Society for Paediatric Gastroenterology. The need for nutrition support teams in pediatric units: a commentary by the ESPGHAN committee on nutrition. J Pediatr Gastroenterol Nutr 2005; 41: 8-11.
- 14) Teixeira AF, Viana KD. Nutritional screening in hospitalized pediatric patients: a systematic review. J Pediatr (Rio J) 2016; 92: 343-352.
- 15) Green Corkins K, Teague EE. Pediatric Nutrition Assessment. Nutr Clin Pract 2017; 32: 40-51.

- SALA A, PENCHARZ P, BARR RD. Children, cancer, and nutrition--A dynamic triangle in review. Cancer 2004; 100: 677-687.
- MURPHY AJ, WHITE M, DAVIES PS. Body composition of children with cancer. Am J Clin Nutr 2010; 92: 55-60
- EDEFONTI A, PICCA M, PAGLIALONGA F, LOI S, GRASSI MR, ARDISSINO G, MARRA G, GHIO L, FOSSALI E. A novel objective nutritional score for children on chronic peritoneal dialysis. Perit Dial Int 2002: 22: 602-607.
- EDEFONTI A, MASTRANGELO A, PAGLIALONGA F. Assessment and monitoring of nutrition status in pediatric peritoneal dialysis patients. Perit Dial Int 2009; 29 Suppl 2: S176-179.
- Kondrup J, Allison SP, Elia M, Vellas B, Plauth M; Educational and Clinical Practice Committee, European Society of Parenteral and Enteral Nutrition (ESPEN). ESPEN guidelines for nutrition screening 2002. Clin Nutr 2003; 22: 415-421.
- JOOSTEN KF, HULST JM. Nutritional screening tools for hospitalized children: methodological considerations. Clin Nutr 2014; 33: 1-5.
- HARTMAN C, SHAMIR R, HECHT C, KOLETZKO B. Malnutrition screening tools for hospitalized children. Curr Opin Clin Nutr Metab Care 2012; 15: 303-309.
- ERKAN T. Methods to evaluate the nutrition risk in hospitalized patients. Turk Pediatri Ars 2014; 49: 276-281.
- 24) Reilly HM, Martineau JK, Moran A, Kennedy H. Nutritional screening--evaluation and implementation of a simple Nutrition Risk Score. Clin Nutr 1995; 14: 269-273.
- 25) SERMET-GAUDELUS I, POISSON-SALOMON AS, COLOMB V, BRUSSET MC, MOSSER F, BERRIER F, RICOUR C. Simple pediatric nutritional risk score to identify children at risk of malnutrition. Am J Clin Nutr 2000; 72: 64-70.
- Secker DJ, Jeejeebhoy KN. Subjective global nutritional assessment for children. Am J Clin Nutr 2007; 85: 1083-1089.
- WONOPUTRI N, DJAIS JT, ROSALINA I. Validity of nutritional screening tools for hospitalized children. Acta Paediatr 2013; 102: e419-423.
- 28) McCarthy H, Dixon M, Crabtree I, Eaton-Evans MJ, McNulty H. The development and evaluation of the Screening Tool for the Assessment of Malnutrition in Paediatrics (STAMP©) for use by healthcare staff. J Hum Nutr Diet 2012; 25: 311-318.
- 29) GERASIMIDIS K, KEANE O, MACLEOD I, FLYNN DM, WRIGHT CM. A four-stage evaluation of the Paediatric Yorkhill Malnutrition Score in a tertiary paediatric hospital and a district general hospital. Br J Nutr 2010; 104: 751-756.
- HULST JM, ZWART H, HOP WC, JOOSTEN KF. Dutch national survey to test the STRONGkids nutritional risk screening tool in hospitalized children. Clin Nutr 2010; 29: 106-111.
- 31) HUYSENTRUYT K, ALLIET P, MUYSHONT L, ROSSIGNOL R, DEVREKER T, BONTEMS P, DEJONCKHEERE J, VANDENPLAS Y, DE SCHEPPER J. The STRONG(kids) nutritional screening tool in hospitalized children: a validation study. Nutrition 2013; 29: 1356-1361.
- 32) SPAGNUOLO MI, LIGUORO I, CHIATTO F, MAMBRETTI D, GUARINO A. Application of a score system to evaluate the risk of malnutrition in a multiple hospital setting. Ital J Pediatr 2013; 39: 81.

- 33) Moeeni V, Walls T, Day AS. Assessment of nutritional status and nutritional risk in hospitalized Iranian children. Acta Paediatr 2012; 101: e446-451.
- 34) MOEENI V, WALLS T, DAY AS. Nutritional status and nutrition risk screening in hospitalized children in New Zealand. Acta Paediatr 2013; 102: e419-423.
- 35) WHITE M, LAWSON K, RAMSEY R, DENNIS N, HUTCHINSON Z, SOH XY, MATSUYAMA M, DOOLAN A, TODD A8, ELLIOTT A, BELL K, LITTLEWOOD R. Simple nutrition screening tool for pediatric inpatients. JPEN J Parenter Enteral Nutr 2016; 40: 392-398.
- 36) HUYSENTRUYT K, DEVREKER T, DEJONCKHEERE J, DE SCHEP-PER J, VANDENPLAS Y, COOLS F. Accuracy of nutritional screening tools in assessing the risk of undernutrition in hospitalized children. J Pediatr Gastroenterol Nutr 2015; 61: 159-166.
- 37) MAKA DA, MURPHY LK. Drug-nutrient interactions: a review. AACN Clin Issues 2000; 11: 580-589.
- Persson LA, Carlgren G. Measuring children's diets: evaluation of dietary assessment techniques in infancy and childhood. Int J Epidemiol 1984; 13: 506-517.
- 39) MILLS VC, SKIDMORE PM, WATSON EO, TAYLOR RW, FLEM-ING EA, HEATH AL. Relative validity and reproducibility of a food frequency questionnaire for identifying the dietary patterns of toddlers in New Zealand. J Acad Nutr Diet 2015; 115: 551-558.
- Pogatshnik C, Hamilton C. Nutrition-focused physical examination: skin, nails, hair, eyes, and oral cavity. Support Line 2011; 33: 7-13.
- Green Corkins K. Nutrition-focused physical examination in pediatric patients. Nutr Clin Pract 2015; 30: 203-209.
- COLLINS N, HARRIS C. The physical assessment revisited: inclusion of the nutrition-focused physical exam. Ostomy Wound Manage, 2010.
- 43) JENSEN GL. Inflammation as the key interface of the medical and nutrition universes: a provocative examination of the future of clinical nutrition and medicine. JPEN J Parenter Enteral Nutr 2006; 30: 453-463.
- 44) ABRAHAM AG, MAK RH, MITSNEFES M, WHITE C, MOX-EY-MIMS M, WARADY B, FURTH SL. Protein energy wasting in children with chronic kidney disease. Pediatr Nephrol 2014; 29: 1231-1238.
- 45) FRISANCHO AR. Anthropometric Standards: An Interactive Nutritional Reference of Body Size and Body Composition for Children and Adults. 2nd ed. Ann Arbor: University of Michigan Press, 2008.
- 46) CENTERS FOR DISEASE CONTROL AND PREVENTION. National Health and Nutrition Examination Survey (NHANES): Anthropometry Procedures Manual. Atlanta, GA: Centers for Disease Control and Prevention, 2007.
- 47) Hogan SE. Knee height as a predictor of recumbent length for individuals with mobility-impaired cerebral palsy. J Am Coll Nutr 1999; 18: 201-205.
- 48) CHUMLEA WMC, GUO S, STEINBAUGH ML. Prediction of stature from knee height for black and white adults and children with application to mobility-impaired or handicapped persons. J Am Diet Assoc 1994; 94: 1385-1388.
- 49) FROEHLICH-GROBE K, NARY DE, VAN SCIVER A, LEE J, LITTLE TD. Measuring height without a stadiometer: empirical investigation of four height estimates among wheelchair users. Am J Phys Med Rehabil 2011; 90: 658-666.
- STEVENSON RD. Use of segmental measures to estimate stature in children with cerebral palsy. Arch Pediatr Adolesc Med 1995; 149: 658-662.

- 51) IVANOVIC DM, LEIVA BP, PÉREZ HT, OLIVARES MG, DÍAZ NS, URRUTIA MS, ALMAGIÀ AF, TORO TD, MILLER PT, BOSCH EO, LARRAÍN CG. Head size and intelligence, learning, nutritional status and brain development. Neuropsychologia 2004; 42: 1118-1131
- GARROW JS, Webster J. Quetelet's index (W/H2) as a measure of fatness. Int J Obesity 1985; 9: 147-153.
- DIETZ WH, ROBINSON TN. Use of the body mass index (BMI) as a measure of overweight in children and adolescents. J Pediatr 1998; 132: 191-193.
- 54) COLE TJ, BELLIZZI MC, FLEGAL KM, DIETZ WH. Establishing a standard definition for child overweight and obesity: international survey. BMJ 2000; 320: 1240-1243.
- WHO. Physical status: the use and interpretation of anthropometry. Geneva: WHO, 1995.
- Cole TJ, Flegal KM, Nicholls D, Jackson AA. Body mass index cut offs to define thinness in children and adolescents: international survey. BMJ 2007; 335: 194.
- 57) Roy NC. Use of mid-upper arm circumference for evaluation of nutritional status of children and for identification of high-risk groups for malnutrition in rural Bangladesh. J Health Popul Nutr 2000; 18: 171-180.
- 58) CACCIALANZA R, CEREDA E, KLERSY C, BONARDI C, CAPPELLO S, QUARLERI L, TURRI A, MONTAGNA E, IACONA I, VALENTINO F, PEDRAZZOLI P. Phase angle and handgrip strength are sensitive early markers of energy intake in hypophagic, non-surgical patients at nutritional risk, with contraindications to enteral nutrition. Nutrients 2015; 7: 1828-1840.
- 59 KÄRKKÄINEN M, RIKKONEN T, KRÖGER H, SIROLA J, TUPPU-RAINEN M, SALOVAARA K, AROKOSKI J, JURVELIN J, HONK-ANEN R, ALHAVA E. Physical tests for patient selection for bone mineral density measurements in postmenopausal women. Bone 2009; 44: 660-665.
- GALE CR, MARTYN CN, COOPER C, SAYER AA. Grip strength, body composition, and mortality. Int J Epidemiol 2007; 36: 228-235.
- 61) RANTANEN T, VOLPATO S, FERRUCI L, HEIKKINEN E, FRIED LP, GURALNIK JM. Handgrip strength and cause-specific and total mortality in older disabled women: exploring the mechanism. J Am Geriatr Soc 2003; 51: 636-641.
- 62) SAINT-MAURICE PF, LAURSON KR, KARSAI I, KAJ M, CSÁNYI T. Establishing Normative Reference Values for Handgrip Among Hungarian Youth. Res Q Exerc Sport 2015; 86 Suppl 1: S29-36.
- 63) SANCHEZ-DELGADO G, CADENAS-SANCHEZ C, MORA-GONZALEZ J, MARTINEZ-TELLEZ B, CHILLÓN P, LÖF M, ORTEGA FB, Ruiz JR. Assessment of handgrip strength in preschool children aged 3 to 5 years. J Hand Surg Eur Vol 2015; 40: 966-972.
- 64) HÉBERT LJ, MALTAIS DB, LEPAGE C, SAULNIER J, CRÊTE M, PERRON M. Isometric muscle strength in youth assessed by hand-held dynamometry: a feasibility, reliability, and validity study. Pediatr Phys Ther 2011; 23: 289-299.
- 56) Mehta NM, Corkins MR, Lyman B, Malone A, Goday PS, Carney LN, Monczka JL, Plogsted SW, Schwenk WF; American Society for Parenteral and Enteral Nutrition Board of Directors. Defining pediatric malnutrition: a paradigm shift toward etiology-related definitions. JPEN J Parenter Enteral Nutr 2013; 37: 460-481.

- 66) WATERLOW JC, BUZINA R, KELLER W, LANE JM, NICHAMAN MZ, TANNER JM. The presentation and use of height and weight data for comparing the nutritional status of groups of children under the age of 10 years. Bull World Health Organ 1977; 55: 489-498.
- 67) CENTERS FOR DISEASE CONTROL AND PREVENTION. National Center for Health Statistics: growth charts. www.cdc.gov/growthcarts, 2016.
- 68) WHO MULTICENTRE GROWTH REFERENCE STUDY GROUP. WHO Child Growth Standards based on length/ height, weight and age. Acta Paediatr Suppl 2006; 450: 76-85.
- JOOSTEN KF, HULST JM. Malnutrition in pediatric hospital patients: current issues. Nutrition 2011; 27: 133-137.
- 70) Lazzer S, Bedogni G, Agosti F, De Col A, Mornati D, Sartorio A. Comparison of dual-energy X-ray absorptiometry, air displacement plethysmography and bioelectrical impedance analysis for the assessment of body composition in severely obese Caucasian children and adolescents. Br J Nutr 2008; 100: 918-924.
- 71) WANG Z, ST-ONGE MP, LECUMBERRI B, PI-SUNYER FX, HES-HKA S, WANG J, KOTLER DP, GALLAGHER D, WIELOPOLSKI L, PIERSON RN JR, HEYMSFIELD SB. Body cell mass: model development and validation at the cellular level of body composition. Am J Physiol Endocrinol Metab 2004; 286: E123-128.
- MURPHY AJ, WHITE M, DAVIES PS. The validity of simple methods to detect poor nutritional status in paediatric oncology patients. Br J Nutr 2009; 101: 1388-1392.
- 73) MURPHY AJ, WHITE M, ELLIOTT SA, LOCKWOOD L, HALLA-HAN A, DAVIES PS. Body composition of children with cancer during treatment and in survivorship. Am J Clin Nutr 2015; 102: 891-896.
- 74) CROOK TA, ARMBYA N, CLEVES MA, BADGER TM, ANDRES A. Air displacement plethysmography, dual-energy X-ray absorptiometry, and total body water to evaluate body composition in preschool-age children. J Acad Nutr Diet 2012; 112: 1993-1998.
- LEEAB SY AND GALLAGHERC D. Assessment methods in human body composition. Curr Opin Clin Nutr Metab Care 2008; 11: 566-572.
- 76) LOHMAN TG, GOING SB. Body composition assessment for development of an international growth standard for preadolescent and adolescent children. Food Nutr Bull 2006; 27: S314-325.
- 77) Lai S, Molfino A, Coppola B, De Leo S, Tommasi V, Galani A, Migliaccio S, Greco EA, Gnerre Musto T, Muscaritoli M. Effect of personalized dietary intervention on nutritional, metabolic and vascular indices in patients with chronic kidney disease. Eur Rev Med Pharmacol Sci 2015; 19: 3351-3359.
- 78) ESPEN TEXTBOOK: Basics in clinical nutrition, fourth edition. Galen Edition, 2011
- 79) TYRRELL VJ, RICHARDS G, HOFMAN P, GILLIES GF, ROBINSON E, CUTFIELD WS. Foot-to-foot bioelectrical impedance analysis: a valuable tool for the measurement of body composition in children. Int J Obes Relat Metab Disord 2001; 25: 273-278.
- 80) KYLE UG, EARTHMAN CP, PICHARD C, COSS-BU JA. Body composition during growth in children: limitations and perspectives of bioelectrical impedance analysis. Eur J Clin Nutr 2015; 69: 1298-1305.

- 81) TALMA H, CHINAPAW MJ, BAKKER B, HIRASING RA, TERWEE CB, ALTENBURG TM. Bioelectrical impedance analysis to estimate body composition in children and adolescents: a systematic review and evidence appraisal of validity, responsiveness, reliability and measurement error. Obes Rev 2013; 14: 895-905.
- Kabiri LS, Hernandez DC, Mitchell K. Reliability, validity, and diagnostic value of a pediatric bioelectrical impedance analysis scale. Child Obes 2015; 11: 650-655.
- 83) BARBOSA-SILVA MC, BARROS AJ, WANG J, HEYMSFIELD SB, PIERSON RN JR. Bioelectrical impedance analysis: population reference values for phase angle by age and sex. Am J Clin Nutr 2005; 82: 49-52.
- 84) Bosy-Westphal A, Danielzik S, Dörhöfer RP, Later W, Wiese S, Müller MJ. Phase angle from bioelectrical impedance analysis: population reference values by age, sex, and body mass index. JPEN J Parenter Enteral Nutr 2006; 30: 309-316.
- 85) SIDDIOUI NI, KHAN SA, SHOEB M, BOSE S. J Clin Anthropometric Predictors of Bio-Impedance Analysis (BIA) Phase Angle in Healthy Adults. Diagn Res 2016; 10: CC01-4.
- 86) KYLE UG, GENTON L, PICHARD C. Low phase angle determined by bioelectrical impedance analysis is associated with malnutrition and nutritional risk at hospital admission. Clin Nutr 2013; 32: 294-299.
- 87) GUPTA D, LAMMERSFELD CA, BURROWS JL, DAHLK SL, VASHI PG, GRUTSCH JF, HOFFMAN S, LIS CG. Bioelectrical impedance phase angle in clinical practice: implications for prognosis in advanced colorectal cancer. Am J Clin Nutr 2004; 80: 1634-1638.
- 88) Gupta D, Lis CG, Dahlk SL, Vashi PG, Grutsch JF, Lam-Mersfeld CA. Bioelectrical impedance phase angle as a prognostic indicator in advanced pancreatic cancer. Br J Nutr 2004; 92: 957-962.

- 89) SCHÜTTE K, TIPPELT B, SCHULZ C, RÖHL FW, FENEBERG A, SEIDENSTICKER R, AREND J, MALFERTHEINER P. Malnutrition is a prognostic factor in patients with hepatocellular carcinoma (HCC). Clin Nutr 2015; 34: 1122-1127.
- FARIAS CL, CAMPOS DJ, BONFIN CM, VILELA RM. Phase angle from BIA as a prognostic and nutritional status tool for children and adolescents undergoing hematopoietic stem cell transplantation. Clin Nutr 2013; 32: 420-425.
- 91) PILEGGI VN, MONTEIRO JP, MARGUTTI AV, CAMELO JS JR. Prevalence of child malnutrition at a university hospital using the World Health Organization criteria and bioelectrical impedance data. Braz J Med Biol Res 2016; 49. pii: S0100-879X2016000300705.
- 92) CEDERHOLM T, BARAZZONI R, AUSTIN P, BALLMER P, BIOLO G, BISCHOFF SC, COMPHER C, CORREIA I, HIGASHIGUCHI T, HOLST M, JENSEN GL, MALONE A, MUSCARITOLI M, NYULASI I, PIRLICH M, ROTHENBERG E, SCHINDLER K, SCHNEIDER SM, DE VAN DER SCHUEREN MA, SIEBER C, VALENTINI L, YU JC, VAN GOSSUM A, SINGER P. ESPEN guidelines on definitions and terminology of clinical nutrition. Clin Nutr 2017; 36: 49-64.
- 93) EDEFONTI A, PICCA M, DAMIANI B, GARAVAGLIA R, LOI S, ARDISSINO G, MARRA G, GHIO L. Prevalence of malnutrition assessed by bioimpedance analysis and anthropometry in children on peritoneal dialysis. Perit Dial Int 2001; 21: 172-179.
- 94) EDEFONTI A, PAGLIALONGA F, PICCA M, PERFUMO F, VERRINA E, LAVORATTI G, RINALDI S, RIZZONI G, ZACCHELLO G, CIOFANI A, SORINO P, LOI S, GRASSI MR. A prospective multicentre study of the nutritional status in children on chronic peritoneal dialysis. Nephrol Dial Transplant 2006; 21: 1946-1951.