

Immune related factors in pathogenesis of autism spectrum disorders

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Abstract. – OBJECTIVE: Etiology of Autism Spectrum Disorders (ASD) is insufficiently known. It is suggested that genes play a crucial role in ASD but additional environmental factors to exacerbate the syndrome are needed. Recently, the inflammatory factors in ASD that may predispose to the disorder attract a great attention. Therefore, the aim of this article was to review the literature on the possible association of the immune system malfunctions with the risk of developing ASD.

MATERIALS AND METHODS: Available articles from PubMed and Google Scholar were analyzed using time descriptors: 1996-2015 and key words: autism spectrum disorder, cytokines and immune system.

RESULTS: Individuals with ASD demonstrate aberrant immune response in central nervous system, peripheral blood and gastrointestinal tract.

CONCLUSIONS: Immune malfunctions may play a role in developing ASD.

Key Words:

Autism, Autism spectrum disorders, Immune system, Cytokines, Neuroinflammation,

Individuals with ASD demonstrate an aberrant immune response in central nervous system (CNS)⁴⁻⁷, peripheral blood^{8,9} and the gastrointestinal tract¹⁰⁻¹³. Overactivation within microglia and astrocytes was proved in *post-mortem* brain biopsies and biomarker studies^{14,15}. Increased autoimmunity mainly due to maternal anti-fetal brain antibodies crossing the placenta during pregnancy was found¹⁶. Inflammatory cytokines concentration was found to be altered in serum, placenta and cerebral spinal fluid in children with autism and their relatives^{4,15,17}. Moreover, skewed production of immunoglobulins or B- and T-cell dysfunction confirmed that humoral and cellular response is altered in individuals with ASD¹⁸. Additionally, the presence of gastritis, lymphoid nodular hyperplasia, colonic lymphoid nodular hyperplasia, eosinophilic infiltration and others¹² confirmed that these inflammatory alterations affect the digestive system as well.

The immune and CNS communicate extensively. While lymphoid organs are hardwired by autonomic nervous system and neuroendocrine hormones serve as a regulator of cytokines balance, the immune homeostasis is critical for proper neurodevelopment and thereby behavior¹⁹⁻²¹. These brain-to-immune interactions may be responsible for developing immune-mediated diseases.

Cytokine proteins impact directly on neurons activity. They are involved in neurodevelopment, as well in prenatal as in postnatal period²². There is also a body of evidence linking cytokines to higher order neurological functions, including cognition and memory^{23,24}. Therefore, disruption within cytokine homeostasis may be responsible for a variety of neurological consequences relevant to ASD.

Introduction

Autism spectrum disorder (ASD) is a disabling neurodevelopmental disorder with social and communication deficits and stereotypic behaviours¹. The etiology of ASD is unclear but evidence suggests multifactorial background. Onore et al² suggest that genes play a crucial role in ASD, while additional environmental factors are needed to exacerbate the syndrome. Of note, it was shown that defects in immunity play a role in brain development and synaptic functions thus in the pathogenesis of ASD³.

Interleukin 1 β (IL-1B) is present in the nervous system during neurogenesis, migration, differentiation, synapse formation, plasticity, and responses to injury²⁵. It was proved that permanent expression of IL-1B in hippocampus impairs spatial memory²⁶. The protein was found to be involved in neural progenitor cell proliferation in particular CNS regions which contribute to region-specific growth in autistic brains^{3,27}. IL-1B is responsible for the formation of excitatory synapses²⁸. Moreover, this protein has been associated with altering sleep patterns and together with other cytokines responsible for so called sickness behaviour characterized by lethargy, depression, anxiety, and difficulties in ability to focus and express social skills²⁹.

Interleukin 6 (IL-6) is involved in neuronal precursors self-renewing, neuronal migrating and promoting cell survival³⁰. The cytokine was found to be responsible for regulating neurite outgrowth, as well³¹. The protein and its receptors are expressed in brain of healthy and disease states, however a low expression is needed for proper neurodevelopment³². It was proved that permanent IL-6 overexpression results in reducing expression of glutamate receptors and L-type calcium channels³³. IL-6 also promotes the predominance of excitatory to inhibitory synapses³⁴ and the development of proper recognition memory³⁵.

Interleukin 4 (IL-4) being expressed in the brain is responsible for promoting oligodendrogenesis of neuronal progenitor cells³⁶. The cytokine was also found to be involved in regulating progenitor cell proliferation and differentiation³⁷ and synapse formation, particularly of GABAergic type³⁸. IL-4 also plays a neuroprotective role and promotes the development of higher order cognitive processes and the absence of the protein is associated with loss of T-cell function in CNS²⁴.

Interferon gamma (IFN- γ) was found to be responsible for neuronal differentiation of neural progenitor cells³⁹. The protein serves as a regulator of dendritic morphology and synapses formation and activity⁴⁰. It was proved that overexpressing IFN- γ leads to elevated concentration of major histocompatibility complex I (MHC I) proteins in the brain⁴¹ and these proteins have been lately found to be expressed in CNS to regulate synaptic scaling⁴².

Tumor necrosis factor- α (TNF- α) may induce cell death on neurons and prune synapses⁴³. Transforming growth factor β (TGF- β 1) acts as a regulator of neuronal migration, neural cells survival and synapse formation. The lack of the expression

of the protein results in altered CNS development as extracellular matrix is disintegrated and deficits in glutamatergic and GABAergic synapses function occur^{44,45} leading to seizures, motor incoordination and severe behavioral abnormalities^{46,47}.

In the light of these facts, the aim of this report was to review the literature on the links between the immune-related factors and developing ASD.

Materials and Methods

Available articles from PubMed and Google Scholar were analyzed using time descriptors: 1996-2015 and key words: autism spectrum disorder, cytokines, immune system, immunity.

In order to present the analyzed issues in the clearest and most comprehensive way, the article is divided into the following subsections:

- Altered immune function in ASD patients in prenatal period
- Altered immune function in postnatal period
 - Neuroinflammation
 - Cytokine and chemokine profile
 - Immunoglobulin levels
 - Cellular response
 - Gastrointestinal malfunctions
 - Sensitivity toward environmental toxicants

Results

Altered Immune Function in Asd Patients in Prenatal Period

Growing evidence indicates that maternal autoantibodies influence harmfully to fetus brain development^{48,49}. These are IgG class immunoglobulins providing passive immunity to the fetus as they are able to transplacental transmission, nevertheless pathogenic autoantibodies are devoid of those advantageous properties¹⁷. In animal studies it was shown that they alter socially the behavior of the offspring. Moreover, it was proved that specific neural antibody are induced during gestation and the offspring demonstrates histological abnormalities in the brain and cognitive declines in later life⁴⁹⁻⁵¹.

The direct properties of these molecules, either in terms of pathogenicity or targets of antibodies, have not been fully identified. Nevertheless in 2008, Braunschweig et al characterized those molecules and showed significantly higher autoreactivity toward a protein at 37 and 73kDa in mothers of ASD children^{16,52}. The researchers

have now identified 7 specific target antigens: lactate dehydrogenase A and B (LDH), cypin, stress-induced phosphoprotein 1 (STIP1), collapsin response mediator proteins 1 and 2 (CRMP1, CRMP2), and Y-box-binding protein and have coined the term “maternal autoantibody-related,” or MAR, autism for these cases. Exclusive reactivity to specific antigen combinations was noted in 23% of mothers of children with ASD and in only 1% of mothers of normally developing children⁵³. The putative antigens are thought to be present on GABAergic interneurons in the brain^{54,55}.

While vitamin D acts as a neuroactive molecule it participates in neuron differentiation and synapses action⁵⁶. Link between vitamin D deficiency in pregnant women and their child developing ASD exists. Autistic characteristics seem to disappear after administering the vitamin⁵⁷, and conception during low UVB penetration favors ASD children deliveries⁵⁸. Pioggia et al⁵⁹ in their systematic review concluded that vitamin D₃ deficiency is associated with ASD especially in dark skinned children and the effect of the lack of vitamin D₃ in pregnant mothers is transmitted to the offspring. Few animal model experiments proved the impact of proper vitamin D level during gestation on brain and ventricles sizes, often enlarged in autistic subjects⁶⁰. In general, there is still no conclusive evidence for an association between the mother having vitamin D deficiency and the offspring developing ASD as few studies lacking such evidence exist^{61,62}.

Exposure to various infections (viral and bacterial) in prenatal life increases the risk for developing ASD. Particularly rubella, herpes simplex, cytomegalovirus, measles or viral meningitis in the first pregnancy trimester or bacterial infection during the second trimester are associated with ASD behaviour in their offspring⁶³. Maternal infection alters specific cytokine levels as well in maternal body as in placenta and fetal brain. It was proved that cord blood concentration of IL1-B facilitates neurological outcome in infants exposed to neonatal hypoxia^{64,65}. Other study showed no association between mild common infections or febrile episodes and ASD in offspring, but reported increased risk for ASD child delivery after maternal influenza infection or prolonged fever and antibiotics administration⁶⁶. In fact when influenza virus was intranasally infused shortly after fertilization in mice, the offspring's social behaviour was altered⁶⁷. Intraperitoneal administration of poly (I:C) mimicking viral infection, at different times of gestational days resulted in various behavioral phenotypes. High anxiety and decreased social

interaction was observed when infection factor was administered at gestational day (GD) 12.5 while perseverant behaviour was noted in case of GD17 and additionally unobserved when inflammogen was infected at GD9^{68,69}. It proves that a critical period for the development of certain pro-social behaviors exists and that susceptibility to the development of ASD increases during GD17⁷⁰. Furthermore rodent experiments suggested that maternal immune activation can result in spatially restricted deficit in Purkinje cells⁷¹.

Additionally, maternal history of any autoimmune disease shows increased risk for ASD⁷². The association was confirmed in epidemiological studies in 40% of studied patients^{72,73}. The significant association was reported for autoimmune thyroiditis or hypothyroidism, rheumatic fever, rheumatoid arthritis, celiac disease, ulcerative colitis, psoriasis and Type 1 diabetes⁷⁴. Also, epigenetic regulation of transcription is believed to be associated with autoimmune diseases, therefore immunogenetical components of ASD may play a crucial role in developing ASD⁷⁵. Variations in MET proto-oncogene tyrosine kinase pathway or serine and threonine kinase C genes can be inherited by ASD children and should be considered as they are involved in innate and adaptive immunity^{76,77}. Other genes that should take attention are those related to NK cells, macrophage inhibitory factor, reelin or mitochondrial respiratory chain disease⁷⁴.

Altered Immune Function in Asd Patients in Postnatal Period

Neuroinflammation

Ongoing neuroinflammation in *post-mortem* brain biopsies and in biomarker studies is a proof of dysregulated immunity in ASD^{14,15}. Increased cell packing and small neuronal size in limbic system and paucity of Purkinje and granular cells in cerebellum can partially explain the inflammation, either systemic or local in ASD⁷⁸, however the inflammatory response within CNS has been linked to mast cells and microglia/astrocyte activation⁷⁹.

Microglia cells, present particularly in diencephalon, are mononuclear cells possessing phagocytic activity within CNS⁸⁰. They are responsible for cytokine and reactive oxygen species production within CNS. In addition they regulate synaptogenesis and neurogenesis^{81,82}. Multiple studies reported that microglia cells were prominently activated in ASD brains thereby acted neurodestructively. It was suggested that the activation disrupts brain blood barrier (BBB) in

ASD via secretion of vascular endothelial growth factor⁸³⁻⁸⁵. Moreover, cytokines and chemokines production were highly increased in brain specimens and cerebral spinal fluid (CSF). These were: IFN- γ , IL-1B, IL-6, IL-12p40, TNF- α and chemokine CCL-2^{4,14,15}. The inflammation was discovered particularly in areas with white matter overgrowth shortly after the collapse of the development^{14,15,86}. It was shown that neuron-specific reaction is associated with microglial activation in dorsolateral cortex and the neuronal pattern organization may be disrupted in later ASD life¹⁵. In other *post-mortem* study it was reported that microglial activation was widespread in the fronto-insular and visual cortices in subjects with ASD. Because of the anatomical distinct of these parts it was discussed whether density of microglia in autistic brains is higher throughout the cerebral cortex⁸⁷.

The probable reason for microglia activation may be linked to perturbations in complement production in ASD individuals. The complement component C1q seems to be a key factor which influences neurodevelopment in humans. The protein C1q is the largest component of the complement system to activate the classical complement pathway, by binding of C1 to initiate the given activator for intense antigen-antibody immune complex. C1q has been found to play a key role in microglia-mediated synaptic pruning in typical developing human brain where it can activate the complement cascade and produce C3/CR3 signaling⁸⁸. This signaling in turn further activates and promotes microglia phagocytosis. Inflammatory insults which result in chronic elevation in serum C1q inhibiting factor leading to relative C1q dysfunction^{88,89}.

Astrocytes play role in repairing brain tissue after injury, lining the BBB, balancing extracellular ion concentration and transporting nutrients to neurons⁹⁰. They are involved in synaptogenesis during development, as well. When permanently activated gliosis occurs brain damage develops. Astrocyte markers concentration, for instance, glial fibrillary acidic protein, are elevated as well in the CSF as in *post-mortem* ASD brain tissues⁹¹⁻⁹³. Other astrocyte markers concentration, such as aquaporin 4 and connexin 43 have also been found to be increased in autistic brains⁹⁴.

Cytokine and Chemokine Profile

The cytokine profile, as well pro- as anti-inflammatory, in ASD patients is altered. Few studies reported decreased concentration of TNF- β in serum samples from subjects with ASD and

its linkage with autism severity was proved. Insufficient amount of TGF- β is critical for inflammation control as the molecule is related to cell migration, apoptosis and regulation within immune system and CNS. Lower concentration of TGF- β were found to be associated with lower adaptive behaviours^{95,96}. Increased TNF- α concentration in ASD brain, cerebrospinal fluid or blood cells is able to block synaptic communication⁹⁷. Plasma level of leptin functionally mimicking IL-6 and IL-12 is also increased and able to pass the BBB⁹⁸. The observation was critically made in children with early onset autism⁹⁶. Elevated levels of macrophage inhibitory factor (MIF) was additionally observed in individuals with ASD. As the molecule is responsible for maintaining neural and endocrine systems, the highest concentration of MIF was marked in children with the most severe autism^{99,100}. The findings of recent meta-analysis identified significantly altered concentrations of cytokines as IL-1B, IL-6, IL-8, IFN- γ , eotaxin and monocyte chemotactic protein-1 in ASD, strengthening evidence of an abnormal cytokine profile in ASD where inflammatory signals dominate¹⁰¹. Cytokines are overexpressed upon toll like receptor 2 (TLR2) and TLR4 but not TLR9 stimulation. The overproduction of cytokines has been associated with altered behaviour and nonverbal communication¹⁰². Increased plasma concentrations of platelet derived growth factor (PDGF) have been reported in autistic patients with more impaired communicative and behavioral skills as well as restricted stereotypic activities^{9,103}.

On the other hand, it should be noted that peripheral blood mononuclear cells from ASD patients were found to secrete more IL1RN, sTNFRI and sTNFRII, limiting the inflammatory response, in company with higher production of anti-inflammatory IL-10^{104,105}. It was shown that during a fever, when pro-inflammatory proteins concentration is heightened and T lymphocyte subsets activated, hyperactivity and stereotypia or speech impairment are reduced¹⁰⁶. Taken together, the data indicates that the aforementioned cytokines milieu can influence behaviour and increase risk of developing ASD.

Chemokines serve chemotactic properties within the immune system. Studies confirmed the elevated levels of MCP-1, RANTES, as well as CCL-2 and CCL-5 in autistic patients. Chemokine imbalance was found to be associated with more impaired developmental and adaptive fun-

ction, thereby the exact role of these chemokines in ASD pathogenesis need to be elucidated^{9,74}. The conclusions of the Early Markers for Autism (EMA) study carried out by Zerbo et al¹⁰⁷ are promising. The authors suggested that measurement of immune system function, especially chemokines: MCP-1, RANTES, MIP-1 α , in the first few days of life may aid in the early identification of abnormal neurodevelopment.

Immunoglobulin Levels

The role of antibodies in ASD pathogenesis has been extensively studied. Lower levels of IgM and IgG classes of immunoglobulin were marked in ASD children and linkage with impaired behaviour was proved¹⁸. The later studies revealed increased level of neutralizing IgG4 antibodies among IgG subclass¹⁰⁸. Additionally, in ASD patients alterations in BBB result in inducing expression of specific autoantibodies of IgM, IgG and IgA classes as a consequence of exposure to neuron-derived antigens. The single specific target have yet to be identified nevertheless single studies indicate antibodies against serotonin receptors, myelin basic protein, heat shock proteins, various brain tissue proteins. The lack of target specificity may be a consequence of cellular damage and the emergence/revealing of sequestered or new epitopes due to antibody generation². Such processes promote neuroinflammatory conditions in ASD patients but studies are still continuing to confirm this hypothesis¹⁰⁹.

The presence of circulating antibodies directed toward brain or nuclear proteins⁹⁸ in ASD patients have been mentioned in a few reports. Wills et al⁵⁵ examined plasma samples of children with ASD for antibodies directed against human cerebellar proteins. Protein analyses revealed that 21% of subjects with ASD produce antibodies reactive toward a cerebellar 52kDa protein. Intense immunoreactivity was determined morphologically to be the Golgi cell of the cerebellum⁵⁵. A significantly higher percent seropositivity of anti-nuclear antibodies in ASD patients with/without a family history of autoimmunity in comparison to healthy children was found. Furthermore, ASD individuals have marked serum anti-myelin-associated glycoprotein antibodies¹¹⁰ and elevated serum levels of anti-ganglioside M1 antibodies than healthy children¹¹¹.

Additionally, the presence of anti-nuclear antibodies has been shown to be positively correlated with disease severity, mental retardation and electroencephalogram abnormalities¹¹².

Cellular Response

Several alterations comprising B and T cells, natural killer (NK) cells and monocytes function in company with autoantibodies production have been observed in ASD^{113,114}. There is evidence indicating increased response of Th1 cells instead of Th2¹¹⁵. HLA-DR high level in CD3+ T cells was stated previously as the allele is associated with late cellular response^{9,98}. Moreover, CD26 (dipeptidyl peptidase IV) expression was increased on CD8+ T cells. CD26 being marker associated with effector cell phenotype in human CNS diseases makes the discovery significant⁹. *In vitro* stimulation of co-stimulatory and activation markers resulted in higher levels of CD137 and decreased levels of CD134, CD25 on T cells of children suffering from ASD⁹. Increased T lymphocytes activation may be associated with maintaining activated cells due to decreased apoptosis, as seen in Crohn's disease¹¹⁶. Additionally, adhesion molecules are known to control the passage of T cells across endothelium. In high functioning autistic children, the concentration of such adhesion factors like sPECAM-1, sP-Selectin and sL-Selectin were decreased^{117,118} and the low selectin level has been found to be associated with altered social skills¹¹⁸.

NK cells serving viral response, tumor cytotoxicity and autoimmunity roles have been shown to be dysregulated in ASD^{119,120}. The expression of few NK cells receptors and effectors were altered and significantly associated with NK cells function. These were perforin, granzyme B, and IFN γ marked under resting conditions in children with ASD. Precise stimulation of NK cells obtained from ASD patients showed decreased cytotoxicity in compare to control cells. It was proposed that low glutathione, IL-2 and IL-15 may be responsible¹²¹. In general, the mechanism behind this low NK cell activity need to be clarified.

Monocytes identify pathogens *via* TLRs and subsequently direct immune response. Dormant, peripheral blood monocyte life is short but upon inflammation they tend to escape apoptosis and *via* CCL-2 and CCL-7 chemokines are transported to the inflammation area where they differentiate into macrophages^{122,123}. When recruited into CNS they develop into microglial cells and play crucial role in CNS inflammation¹¹⁴. Significant differences in pro-inflammatory cytokine production by monocytes from ASD patients were observed under different TLRs stimulation. While TLR-2 and TLR-4 favoured the condition, TLR-9 stimulation decreased cytokine production. This altered immunity response in ASD subjects can have a wide range impact on neural function in autistic people¹⁰³.

Gastrointestinal Malfunctions

Immune alterations within gastrointestinal tract have been associated with ASD phenotype as well as its contribution to the neurodevelopment or autism severity¹²⁴. Symptoms of digestive tract diseases was observed in 9-84.1% autistic individuals depending on study type (retrospective vs prospective) and inclusion criteria¹²⁵⁻¹²⁸. Symptoms such as excessive production of intestinal gases, flatulence, abdominal pain, diarrhea, belching, symptoms of gastroesophageal reflux or constipation have been attributed to changes in gut microflora as well as elevated intestinal permeability and intestinal inflammation^{129,130}. Interviewing the parents of ASD patients indicated coexisting severe behavioral symptoms with the ongoing gastrointestinal problems.

Intestinal microbiota is an important factor contributing to the organism homeostasis. Intestinal dysbiosis, being a qualitative-quantitative disorder among particular groups of microorganisms, is considered to be the etiological agent of many types of diseases, *inter alia* inflammatory bowel diseases, obesity and atopic diseases¹³¹. Gastrointestinal tract microbiome and enteric neurons are directly connected, forming part of the axis of microbes-small intestine-brain. Therefore, regulatory activity of microbiota on the CNS occurs via neuronal, endocrine, metabolic and immune pathways¹³². Shaw et al¹³³ attempted to understand the correlation of intestinal microbiota with the occurrence of clinical symptoms of autism in a case study. There were two brothers with ASD in the study who showed the presence of Krebs cycle metabolite analogues and high concentrations of arabinose in the urine. The compounds were not present in the urine of healthy subjects. The described phenomenon is considered as the result of colonization of the gastrointestinal tract by bacteria and/or yeasts, whose metabolites are inhibitors of mitochondrial Krebs cycle. Moreover, potential impact of the microbiota in the initiation/ deterioration of ASD symptoms comes from a significant improvement in behaviour of children overtreated with vancomycin^{134,135}. The frequent use of antibiotics, mainly because of ear infections, was thought to interfere strongly intestinal ecosystem, promote the multiplication of pathogenic microorganisms, including toxigenic strains of the *Clostridium sp.* Autistic neurological symptoms may develop following the direct action of tetanus neurotoxin-tetanospasmin. Accumulation of bacterial toxins in conjunction with excessive intestinal permeability observed in children with ASD, le-

ads to increased blood concentration of toxicants resulting in systemic symptoms. Further studies of the intestinal ecosystem in children with ASD found a significant increase in the number of different species of *Clostridium* species, in comparison to the control group^{136,137}. These observations were confirmed in Parracho et al¹³⁸ analysis, who described the population of *Clostridium histolyticum* to be overgrown in ASD patients. Finegold et al¹³⁷, discovered the overgrowth of *Clostridium bolteae* in the feces of persons with ASD. In addition to the best documented *Clostridium spp* overgrowth in the gastrointestinal tract of ASD individuals, there are also studies suggesting abnormal commensal bacteria milieu. It was demonstrated that in children with severe ASD *Bacteroidetes* family predominates, while *Firmicutes* were more often found in healthy children¹³⁷.

Gastrointestinal disruptions in autistic patients are functional as well as organic disorders. Immune tissue in gastrointestinal tract is the largest and most complex immune system in humans. Epithelial cells provide both innate and adaptive immune responses¹³⁹. Innocuous milieu within intestine makes proinflammatory Th2-dependent, Th1-dependent delayed-type hypersensitivity, IgG antibodies and Th17-dependent granulocytic responses to be suppressed. Histopathological findings such as gastritis, lymphoid nodular hyperplasia, colonic lymphoid nodular hyperplasia, eosinophilic infiltration and others¹² was significantly more common in children with ASD. Additionally, many inflammatory transcripts detected within gastrointestinal tract are common for ASD and Crohn's disease or ulcerative colitis¹³. Few observed changes (eosinophilic infiltration) can be partially resolved in children with autism as a result of dietary intervention as a result of treating non allergy food hypersensitivity. This is especially evident due to increased permeability of the ASD individuals small intestine mucous membrane which is observed even in patients with nonspecific inflammatory bowel disease, celiac disease or cystic fibrosis^{12,140}. Homeostasis of intestinal barrier provides selective and preferential absorption of nutrients from the intestinal lumen, thereby preventing bloodstream migration of pathogens and toxicants¹⁴¹. Selective loss of intestinal barrier function is thought to be associated with IgE-independent food hypersensitivity with delayed mechanism of reaction. Penetration of undigested food particles through the gut barrier activates the immune system, leads to the specific IgG antibodies production, immune complex formation and

thereby inflammation development¹⁴². Taking together the data, the most common practice is restrictive dietary elimination especially of gluten and casein from the diet of ASD patients¹⁴³. This is also associated with dipeptyl peptidase IV deficiency described in ASD patients. The enzyme is a brush border serine peptidase involved in the digestion of partial decomposition products of casein and gliadin named gliadomorphins and casomorphin, respectively. The described molecules have the properties of opioids therefore penetrating CNS may alter the behaviour of autistic patients¹⁴⁴. So far conducted analyses have shown a significantly increased levels of IgG antibodies specific to both gluten and casein in children with ASD compared with typically developing ones¹³⁰. In the light of these facts, it seems appropriate to assess all food components penetrating through the intestinal barrier and implement targeted elimination diet. Nevertheless, there are difficulties in determining the actual impact of an elimination diet on the functioning of patients with autism. It is known that only a certain group of patients clings to profit from this type of nutrition, while in the rest of the group diet has no effect on both the ailments of the gastrointestinal tract, as well as the level of functioning of the child^{143,145}.

Sensitivity Toward Environmental Toxicants

Environmental toxicants can disrupt proper neurodevelopment and immunity. Autistic children susceptibility to halogenated aromatic hydrocarbons (HAH) is specific as ryanodine receptor expression in autistics brain is altered⁶. All in all HAH generally disrupt immunity by diminishing cellular response and causing lymphoid organs atrophied⁶. Among few HAH toxicants polybrominated diphenyl ethers (PBDEs) have been studied. When peripheral mononuclear blood cells from ASD patients were treated with PBDEs followed by bacterial derivative lipopolysaccharide stimulation, cytokines and chemokines production in cell culture was critically increased. Opposite findings were discovered in control cell cultures originated in typically developing children¹⁴⁶. The toxicant has been, therefore, considered as an immune suppressor in neurotypical people. Many other toxicants such as solvents, phthalates, pesticides or heavy metals have been studied. Mercury and lead may lead to auto-antibody production¹⁴⁷⁻¹⁴⁹ or skewed production of cytokines³. Organochlorine pesticides disrupt calcium and sodium channels as well as GABA receptors thereby induce neuro and immuno-

toxicity^{150,151}. Organophosphates upregulate Th1 and Th2 cytokines which evolve them into malfunction of adaptive response³. Pyrethroids were found to suppress IFN- γ and IL-4¹⁵² production and upregulate production of IL-12 and TNF α ¹⁵³.

Conclusions

The immune response in individuals with ASD is altered. These malfunctions may be responsible for ASD development.

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Conflicts of interest

The authors declare no conflicts of interest.

References

- 1) LORD C, BISHOP SL. Recent Advances in Autism Research as Reflected in DSM-5 Criteria for Autism Spectrum Disorder. *Annu Rev Clin Psychol* 2015; 11: 53-70.
- 2) ONORE C, CAREAGA M, ASHWOOD P. The role of immune dysfunction in the pathophysiology of autism. *Brain Behav Immun* 2013; 26: 383-392.
- 3) GOINES PE, ASHWOOD P. Cytokine dysregulation in autism spectrum disorders (ASD): possible role of the environment. *Neurotoxicol Teratol* 2013; 36: 67-81.
- 4) VARGAS DL, NASCIBENE C, KRISHNAN C, ZIMMERMAN AW, PARDO CA. Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann Neurol* 2005; 57: 67-81.
- 5) GRIGORENKO EL, HAN SS, YRIGOLLEN CM, LENG L, MIZUE Y, ANDERSON GM, MULDER EJ, DE BILDT A, MINDERAA RB, VOLKMAR FR, CHANG JT, BUCALA R. Macrophage migration inhibitory factor and autism spectrum disorders. *Pediatrics* 2008; 122: e438-445.
- 6) VOINEAGU I, WANG X, JOHNSTON P, LOWE JK, TIAN Y, HORVATH S, MILL J, CANTOR RM, BLENCOWE BJ, GESCHWIND DH. Transcriptomic analysis of autistic brain reveals convergent molecular pathology. *Nature* 2011; 474: 380-384.
- 7) ZIATS MN, RENNERT OM. 2011. Expression profiling of autism candidate genes during human brain development implicates central immune signaling pathways. *PLoS One* 2011; 6: e24691.
- 8) MOLLOY CA, MORROW AL, MEINZEN-DERR J, SCHLEIFER K, DIENGER K, MANNING-COURTNEY P, ALTAYE M, WILLS-KARP

- M. Elevated cytokine levels in children with autism spectrum disorder. *J Neuroimmunol* 2006; 172: 198-205.
- 9) ASHWOOD P, KRAKOWIAK P, HERTZ-PICCIOTTO I, HANSEN R, PESSAH I, VAN DE WATER J. Elevated plasma cytokines in autism spectrum disorders provide evidence of immune dysfunction and are associated with impaired behavioral outcome. *Brain Behav Immun* 2011; 25: 40-45.
- 10) DeFELICE ML, RUCHELLI ED, MARKOWITZ JE, STROGATZ M, REDDY KP, KADIVAR K, MULBERG AE, BROWN KA. Intestinal cytokines in children with pervasive developmental disorders. *Am J Gastroenterol* 2003; 98: 1777-1782.
- 11) ASHWOOD P, ANTHONY A, TORRENTE F, WAKEFIELD AJ. Spontaneous mucosal lymphocyte cytokine profiles in children with autism and gastrointestinal symptoms: mucosal immune activation and reduced counter regulatory interleukin-10. *J Clin Immunol* 2004; 24: 664-673.
- 12) WASILEWSKA J, JAROCKA-CYRTA E, KACZMARSKI M. Patogeneza zaburzeń przewodu pokarmowego u dzieci z autyzmem. *Pol Merk Lek* 2009; 27: 40-43.
- 13) WALKER SJ, FORTUNATO J, GONZALEZ LG, KRIGSMAN A. Identification of unique gene expression profile in children with regressive autism spectrum disorder (ASD) and ileocolitis. *PLoS One* 2013; 8: e58058.
- 14) LI X, CHAUHAN A, SHEIKH AM. Elevated immune response in the brain of autistic patients. *J Neuroimmunol* 2009; 207: 111-116.
- 15) MORGAN JT, CHANA G, PARDO CA. Microglial activation and increased microglial density observed in the dorsolateral prefrontal cortex in autism. *Biol Psychiatry* 2010; 68: 368-376.
- 16) BRAUNSCHWEIG D, GOLUB MS, KOENIG CM, QI L, PESSAH IN, VAN DE WATER J. Maternal autism-associated IgG antibodies delay development and produce anxiety in a mouse gestational transfer model. *J Neuroimmunol* 2012; 252: 56-65.
- 17) ZIMMERMAN AW, CONNORS SL, MATTESON KJ, LEE LC, SINGER HS, CASTANEDA JA. Maternal anti-brain antibodies in autism. *Brain Behav Immun* 2007; 21: 351-357.
- 18) HEUER L, ASHWOOD P, SCHAUER J, GOINES P, KRAKOWIAK P, HERTZ-PICCIOTTO I. Reduced levels of immunoglobulin in children with autism correlates with behavioral symptoms. *Autism Res* 2008; 1: 275-283.
- 19) PATTERSON PH. Maternal infection: window on neuroimmune interactions in fetal brain development and mental illness. *Curr Opin Neurobiol* 2002; 12: 115-118.
- 20) BESEDOVSKY HO, REY AD. Physiology of psychoneuroimmunology: a personal view. *Brain Behav Immun* 2007; 21: 34-44.
- 21) ZIEMSEN T, KERN S. Psychoneuroimmunology--cross-talk between the immune and nervous systems. *J Neurol* 2007; 254 Suppl 2: II8-11.
- 22) DEVERMAN BE, PATTERSON PH. Cytokines and CNS development. *Neuron* 2009; 64: 61-78.
- 23) McAFOOSE J, BAUNE BT. Evidence for a cytokine model of cognitive function. *Neurosci Biobehav Rev* 2009; 33: 355-366.
- 24) DERECKI NC, CARDANI AN, YANG CH, QUINNIES KM, CRIHFELD A, LYNCH KR, KIPNIS J. Regulation of learning and memory by meningeal immunity: a key role for IL-4. *J Exp Med* 2010; 207: 1067-1080.
- 25) GIULIAN D, YOUNG DG, WOODWARD J, BROWN DC, LACHMAN LB. Interleukin-1 is an astroglial growth factor in the developing brain. *J Neurosci* 1988; 8: 709-714.
- 26) MOORE AH, WU M, SHAFTEL SS, GRAHAM KA, O'BANION MK. Sustained expression of interleukin-1beta in mouse hippocampus impairs spatial memory. *Neuroscience* 2009; 164: 1484-1495.
- 27) DE LA MANO A, GATO A, ALONSO M, CARNICERO E, MARTÍN C, MORO JA. Role of interleukin-1beta in the control of neuroepithelial proliferation and differentiation of the spinal cord during development. *Cytokine* 2007; 37: 128.
- 28) YOSHIDA T, YASUMURA M, UEMURA T, LEE SJ, RA M, TAGUCHI R, IWAKURA Y, MISHINA M. IL-1 receptor accessory protein-like 1 associated with mental retardation and autism mediates synapse formation by trans-synaptic interaction with protein tyrosine phosphatase δ . *J Neurosci* 2011; 31: 13485-13499.
- 29) PALIN K, BLUTHÉ RM, MCCUSKER RH, LEVADE T, MOOS F, DANTZER R, KELLEY KW. The type 1 TNF receptor and its associated adapter protein, FAN, are required for TNF α -induced sickness behavior. *Psychopharmacology (Berl)* 2009; 201: 549-556.
- 30) BAUER S, KERR BJ, PATTERSON PH. The neuropoietic cytokine family in development, plasticity, disease and injury. *Nat Rev Neurosci* 2007; 8: 221-232.
- 31) IHARA S, NAKAJIMA K, FUKADA T, HIBI M, NAGATA S, HIRANO T, FUKUI Y. Dual control of neurite outgrowth by STAT3 and MAP kinase in PC12 cells stimulated with interleukin-6. *EMBO J* 1997; 16: 5345-5352.
- 32) GADIENT RA, OTTEN U. Expression of interleukin-6 (IL-6) and interleukin-6 receptor (IL-6R) mRNAs in rat brain during postnatal development. *Brain Res* 1994; 637: 10-14.
- 33) VEREYKEN EJ, BAJOVA H, CHOW S, DE GRAAN PN, GRUOL DL. Chronic interleukin-6 alters the level of synaptic proteins in hippocampus in culture and in vivo. *Eur J Neurosci* 2007; 25: 3605-3616.
- 34) WEI H, ZOU H, SHEIKH AM, MALIK M, DOBKIN C, BROWN WT, LI X. IL-6 is increased in the cerebellum of autistic brain and alters neural cell adhesion, migration and synaptic formation. *J Neuroinflammation* 2011; 8: 52.
- 35) BAIER PC, MAY U, SCHELLER J, ROSE-JOHN S, SCHIFFELHOLZ T. Impaired hippocampus-dependent and -independent learning in IL-6 deficient mice. *Behav Brain Res* 2009; 200: 192-196.
- 36) BUTOVSKY O, ZIV Y, SCHWARTZ A, LANDA G, TALPALAR AE, PLUCHINO S, MARTINO G, SCHWARTZ M. Microglia acti-

- vated by IL-4 or IFN-gamma differentially induce neurogenesis and oligodendrogenesis from adult stem/progenitor cells. *Mol Cell Neurosci* 2006; 31: 149-160.
- 37) DA SILVA AG, CAMPELLO-COSTA P, LINDEN R, SHOLL-FRANCO A. Interleukin-4 blocks proliferation of retinal progenitor cells and increases rod photoreceptor differentiation through distinct signaling pathways. *J Neuroimmunol* 2008; 196: 82-93.
- 38) SHOLL-FRANCO A, MARQUES PM, FERREIRA CM, DE ARAUJO EG. IL-4 increases GABAergic phenotype in rat retinal cell cultures: involvement of muscarinic receptors and protein kinase C. *J Neuroimmunol* 2002; 133: 20-29.
- 39) LEIPZIG ND, XU C, ZAHIR T, SHOICHET MS. Functional immobilization of interferon-gamma induces neuronal differentiation of neural stem cells. *J Biomed Mater Res A* 2010; 93: 625-633.
- 40) KIM JJ, BECK HN, LEIN PJ, HIGGINS D. Interferon gamma induces retrograde dendritic retraction and inhibits synapse formation. *J Neurosci* 2002; 22: 4530-4539.
- 41) CORBIN JG, KELLY D, RATH EM, BAERWALD KD, SUZUKI K, POPKO B. Targeted CNS expression of interferon-gamma in transgenic mice leads to hypomyelination, reactive gliosis, and abnormal cerebellar development. *Mol Cell Neurosci* 1996; 7: 354-370.
- 42) SHATZ CJ. MHC class I: an unexpected role in neuronal plasticity. *Neuron* 2009; 64: 40-45.
- 43) CACCI E, CLAASEN JH, KOKAIA Z. Microglia-derived tumor necrosis factor-alpha exaggerates death of newborn hippocampal progenitor cells in vitro. *J Neurosci Res* 2005; 80: 789-797.
- 44) BRIONNE TC, TESSEUR I, MASLIAH E, WYSS-CORAY T. Loss of TGF-beta 1 leads to increased neuronal cell death and microgliosis in mouse brain. *Neuron* 2003; 40: 1133-1145.
- 45) HEUPEL K, SARGSYAN V, PLOMP JJ, RICKMANN M, VAROQUEAUX F, ZHANG W, KRIEGLSTEIN K. Loss of transforming growth factor-beta 2 leads to impairment of central synapse function. *Neural Dev* 2008; 3: 25.
- 46) WYSS-CORAY T, FENG L, MASLIAH E, RUPPE MD, LEE HS, TOGGAS SM, ROCKENSTEIN EM, MUCKE L. Increased central nervous system production of extracellular matrix components and development of hydrocephalus in transgenic mice overexpressing transforming growth factor-beta 1. *Am J Pathol* 1995; 147: 53-67.
- 47) DEPINO AM, LUCCHINA L, PITOSI F. Early and adult hippocampal TGF-β1 overexpression have opposite effects on behavior. *Brain Behav Immun* 2011; 25: 1582-1591.
- 48) CROEN LA, BRAUNSCHEWIG D, HAAPANEN L, YOSHIDA CK, FIREMAN B, GREYER JK, KHARRAZI M, HANSEN RL, ASHWOOD P, VAN DE WATER J. Maternal mid-pregnancy autoantibodies to fetal brain protein: the early markers for autism study. *Biol Psychiatry* 2008; 64: 583-588.
- 49) LEE JY, HUERTA PT, ZHANG J, KOWAL C, BERTINI E, VOLPE BT. Neurotoxic autoantibodies mediate congenital cortical impairment of offspring in maternal lupus. *Nat Med* 2009; 15: 91-96.
- 50) DALTON P, DEACON R, BLAMIRE A, PIKE M, MCKINLAY I, STEIN J. Maternal neuronal antibodies associated with autism and a language disorder. *Ann Neurol* 2003; 53: 533-537.
- 51) MARTIN LA, ASHWOOD P, BRAUNSCHEWIG D, CABANLIT M, VAN DE WATER J, AMARAL DG. Stereotypies and hyperactivity in rhesus monkeys exposed to IgG from mothers of children with autism. *Brain Behav Immun* 2008; 22: 806-816.
- 52) BRAUNSCHEWIG D, ASHWOOD P, KRAKOWIAK P, HERTZ-PICCIOTTO I, HANSEN R, CROEN LA. Autism: maternally derived antibodies specific for fetal brain proteins. *Neurotoxicology* 2008; 29: 226-231.
- 53) BRAUNSCHEWIG D, KRAKOWIAK P, DUNCANSON P, BOYCE R, HANSEN RL, ASHWOOD P, HERTZ-PICCIOTTO I, PESSAH N, VAN DE WATER J. Autism-specific maternal autoantibodies recognize critical proteins in developing brain. *Transl Psychiatry* 2013; 3: e277.
- 54) ENSTROM AM, VAN DE WATER JA, ASHWOOD P. Autoimmunity in autism. *Curr Opin Investig Drugs* 2009; 10: 463-473.
- 55) WILLS S, CABANLIT M, BENNETT J, ASHWOOD P, AMARAL DG, VAN DE WATER J. Detection of autoantibodies to neural cells of the cerebellum in the plasma of subjects with autism spectrum disorders. *Brain Behav Immun* 2009; 23: 64-74.
- 56) EYLES DW, FERON F, CUI X, KESBY JP, HARMS LH, KO P, MCGRATH JJ, BURNE TH. Developmental vitamin D deficiency causes abnormal brain development. *Psychoneuroendocrinology* 2009; 34 Suppl 1: S247-S257.
- 57) STEWART C, LATIF A. Symptomatic nutritional rickets in a teenager with autistic spectrum disorder. *Child Care Health Dev* 2008; 34: 276-278.
- 58) STEVENS MC, FEIN DH, WATERHOUSE LH. Season of birth effects in autism. *J Clin Exp Neuropsychol* 2000; 22: 399-407.
- 59) PIOGGIA G, TONACCI A, TARTARISCO G, BILLECI L, MURATORI F, RUTA L, GANGEMI S. Autism and lack of D3 vitamin: A systematic review. *Res Autism Spect Dis* 2014; 8: 1685-1698.
- 60) EYLES D, BROWN J, MACKAY-SIM A, MCGRATH J, FERON F. Vitamin D3 and brain development. *Neuroscience* 2003; 118: 641-653.
- 61) KOCOVSÁ E, FERNELL E, BILLSTEDT E, MINNIS H, GILLBERG C. Vitamin D and autism: clinical review. *Res Dev Disabil* 2012; 33: 1541-1550.
- 62) WHITEHOUSE AJ, HOLT BJ, SERRALHA M, HOLT PG, KUSEL MM, HART PH. Maternal serum vitamin D levels during pregnancy and offspring neurocognitive development. *Pediatrics* 2012; 129: 485-493.
- 63) ATLADOTTIR HO, THORSEN P, OSTERGAARD L, SCHENDEL DE, LEMCKE S, ABDALLAH M, PARNER ET. Maternal infection requiring hospitalization during pregnancy and autism spectrum disorders. *J Autism Dev Disord* 2010; 40: 1423-1430.

- 64) ALY H, KHASHABA MT, EL-AYOUTY M, EL-SAYED O, HASANEIN BM. IL-1beta, IL-6 and TNF-alpha and outcomes of neonatal hypoxic ischemic encephalopathy. *Brain Dev* 2006; 28: 178-182.
- 65) LIU J, FENG ZC. Increased umbilical cord plasma interleukin-1 beta levels was correlated with adverse outcomes of neonatal hypoxic-ischemic encephalopathy. *J Trop Pediatr* 2010; 56: 178-182.
- 66) ATLADÓTTIR HÓ, HENRIKSEN TB, SCHENDEL DE, PARNER ET.: Autism after infection, febrile episodes, and antibiotic use during pregnancy: an exploratory study. *Pediatrics* 2012; 130: e1447-1454.
- 67) SHI L, FATEMI SH, SIDWELL RW, PATTERSON PH. Maternal influenza infection causes marked behavioral and pharmacological changes in the offspring. *J Neurosci* 2003; 23: 297-302.
- 68) SMITH SE, LI J, GARBETT K, MIRNICS K, PATTERSON PH. Maternal immune activation alters fetal brain development through interleukin-6. *J Neurosci* 2007; 27: 10695-10702.
- 69) MEYER U, FELDON J, SCHEDLOWSKI M, YEE BK. Immunological stress at the maternal-foetal interface: a link between neurodevelopment and adult psychopathology. *Brain Behav Immun* 2006; 20: 378-388.
- 70) DEPINO AM. Peripheral and central inflammation in autism spectrum disorders. *Mol Cell Neurosci* 2013; 53: 69-76.
- 71) AMARAL DG, SCHUMANN CM, NORDAHL CW. Neuroanatomy of autism. *Trends Neurosci* 2008; 31: 137-145.
- 72) ATLADÓTTIR HO, PEDERSEN MG, THORSEN P, MORTENSEN PB, DELEURAN B, EATON WW, PARNER ET. Association of family history of autoimmune diseases and autism spectrum disorders. *Pediatrics* 2009; 124: 687-694.
- 73) SWEETEN TL, BOWYER SL, POSEY DJ. Increased prevalence of familial autoimmunity in probands with pervasive developmental disorders. *Pediatrics* 2003; 112: e420.
- 74) GESUNDHEIT B, ROSENZWEIG JP, NAOR D, LERER B, ZACHOR DA, PROCHÁZKA V, MELAMED M, KRISTT DA, STEINBERG A, SHULMAN C, HWANG P, KOREN G, WALFISCH A, PASSWEG JR, SNOWDEN JA, TAMOUZA R, LEBOYER M, FARGE-BANCEL D, ASHWOOD P. Immunological and autoimmune considerations of autism spectrum disorders. *J Autoimmun* 2013; 44: 1-7.
- 75) GRAFODATSKAYA D, CHUNG B, SZATMARI P, WEKSBERG R. Autism spectrum disorders and epigenetics. *J Am Acad Child Adolesc Psychiatry* 2010; 49: 794-809.
- 76) LINTAS C, PERSICO AM. Autistic phenotypes and genetic testing: state-of-the-art for the clinical geneticist. *J Med Genet* 2009; 46: 1-8.
- 77) CAMPBELL DB, WARREN D, SUTCLIFFE JS, LEE EB, LEVITT P. Association of MET with social and communication phenotypes in individuals with autism spectrum disorder. *Am J Med Genet B Neuropsychiatr Genet* 2010; 153B: 438-446.
- 78) PALMEN SJ, VAN ENGELAND H, HOF PR, SCHMITZ C. Neuropathological findings in autism. *Brain* 2004; 127: 2572-2583.
- 79) WANG QM, LUO AZ, KONG X. Neuroinflammation and autism. *N A J Med Sci* 2014; 7: 118-122.
- 80) PANG X, LETOURNEAU R, ROZNIECKI JJ, WANG L, THEOHARIDES TC. Definitive characterization of rat hypothalamic mast cells. *Neuroscience* 1996; 73: 889-902.
- 81) PAOLICELLI RC, BOLASCO G, PAGANI F, MAGGI L, SCIANNI M, PANZANELLI P, GIUSTETTO M, FERREIRA TA, GUIDUCCI E, DUMAS L, RAGOZZINO D, GROSS CT. Synaptic pruning by microglia is necessary for normal brain development. *Science* 2011; 333: 1456-1458.
- 82) SIERRA A, ENCINAS JM, DEUDERO JJ. Microglia shape adult hippocampal neurogenesis through apoptosis-coupled phagocytosis. *Cell Stem Cell* 2010; 7: 483-495.
- 83) GALLI SJ. New concepts about the mast cell. *N Engl J Med* 1993; 328: 257-265.
- 84) THEOHARIDES TC, DOYLE R. Autism, gut-blood-brain barrier, and mast cells. *J Clin Psychopharmacol* 2008; 28: 479-483.
- 85) VASIADI M, THERIANOU A, ALYSANDRATOS KD, KATSAROU-KATSARI A, PETRAKOPOULOU T, THEOHARIDES A, PAPADAVID E, STAVRIANAS N, ANTONIOU C, KALOGEROMITROS D, THEOHARIDES TC. Serum neurotensin (NT) is increased in psoriasis and NT induces vascular endothelial growth factor release from human mast cells. *Br J Dermatol* 2012; 166: 1349-1352.
- 86) COURCHESNE E, CARPER R, AKSHOOMOFF N. Evidence of brain overgrowth in the first year of life in autism. *JAMA* 2003; 290: 337-344.
- 87) TETREAULT NA, HAKEEM AY, JIANG S. Microglia in the cerebral cortex in autism. *J Autism Dev Disord* 2012; 42: 2569-2584.
- 88) FANG C, GARBUZOVA-DAVIS S, TAN J, OBREGON D. C1q as a regulator of brain development: implications for autism spectrum disorders. *Brain Disord Ther* 2015; 3: 152.
- 89) GASQUE P, NEAL JW, SINGHRAO SK, MCGREAL EP, DEAN YD, VAN BJ, MORGAN BP. Roles of the complement system in human neurodegenerative disorders: pro-inflammatory and tissue remodeling activities. *Mol Neurobiol* 2002; 25: 1-17.
- 90) SOFRONIEW MV, VINTERS HV. Astrocytes: biology and pathology. *Acta Neuropathol* 2010; 119: 7-35.
- 91) AHLSEN G, ROSENGREN L, BELFRAGE M, PALM A, HAGLID K, HAMBERGER A, HAMBERGER A. Glial fibrillary acidic protein in the cerebrospinal fluid of children with autism and other neuropsychiatric disorders. *Biol Psychiatry* 1993; 33: 734-743.
- 92) ROSENGREN LE, AHLSEN G, BELFRAGE M, GILLBERG C, HAGLID KG, HAMBERGER A. A sensitive ELISA for glial fibrillary acidic protein: application in CSF of children. *J Neurosci Methods* 1992; 44: 113-119.
- 93) LAURENCE JA, FATEMI SH. Glial fibrillary acidic protein is elevated in superior frontal, parietal and cerebellar cortices of autistic subjects. *Cerebellum* 2005; 4: 206-210.

- 94) FATEMI SH, FOLSOM TD, REUTIMAN TJ, LEE S. Expression of astrocytic markers aquaporin 4 and connexin 43 is altered in brains of subjects with autism. *Synapse* 2008; 62: 501-507.
- 95) OKADA K, HASHIMOTO K, IWATA Y, NAKAMURA K, TSUJII M, TSUCHIYA KJ, SEKINE Y, SUDA S, SUZUKI K, SUGIHARA G, MATSUZAKI H, SUGIYAMA T, KAWAI M, MINABE Y, TAKEI N, MORI N. Decreased serum levels of transforming growth factor-beta1 in patients with autism. *Prog Neuropsychopharmacol Biol Psychiatry* 2007; 31: 187-190.
- 96) ASHWOOD P, KWONG C, HANSEN R, HERTZ-PICCIOTTO I, CROEN L, KRAKOWIAK P, WALKER W, PESSAH IN, VAN DE WATER J. Brief report: plasma leptin levels are elevated in autism: association with early onset phenotype? *J Autism Dev Disord* 2008; 38: 169-175.
- 97) ZHANG H, DOUGHERTY PM. Acute inhibition of signaling phenotype of spinal GABAergic neurons by tumor necrosis factor-alpha. *J Physiol (Lond.)* 2011; 589: 4511-4526.
- 98) GOINES P, VAN DE WATER J. The immune system's role in the biology of autism. *Curr Opin Neurol* 2010; 23: 111-117.
- 99) FINGERLE-ROWSON GR, BUCALA R. Neuroendocrine properties of macrophage migration inhibitory factor (MIF). *Immunol Cell Biol* 2001; 79: 368-375.
- 100) GRIGORENKO EL, HAN SS, YRIGOLLEN CM, LENG L, MIZUE Y, ANDERSON GM, MULDER EJ, DE BILDT A, MINDERAA RB, VOLKMAR FR, CHANG JT, BUCALA R. Macrophage migration inhibitory factor and autism spectrum disorders. *Pediatrics* 2008; 122: e438-445.
- 101) MASI A, QUINTANA DS, GLOZIER N, LLOYD AR, HICKIE IB, GUASTELLA AJ. Cytokine aberrations in autism spectrum disorder: a systematic review and meta-analysis. *Mol Psychiatry* 2015; 20: 440-446.
- 102) ENSTROM A, ONORE C, VAN DE WATER J, ASHWOOD P. Differential monocyte responses to TLR ligands in children with autism spectrum disorders. *Brain Behav Immun* 2010; 24: 64-71.
- 103) KAJIZUKA M, MIYACHI T, MATSUZAKI H, IWATA K, SHINMURA C, SUZUKI K, SUDA S, TSUCHIYA KJ, MATSUMOTO K, IWATA Y, NAKAMURA K, TSUJII M, SUGIYAMA T, TAKEI N, MORI N. Serum levels of platelet-derived growth factor BB homodimers are increased in male children with autism. *Prog Neuropsychopharmacol Biol Psychiatry* 2010; 34: 154-158.
- 104) JYONOUCHI H, SUN SN, LE H. Proinflammatory and regulatory cytokine production associated with innate and adaptive immune responses in children with autism spectrum disorders and developmental regression. *J Neuroimmunol* 2001; 120: 170-179.
- 105) CROONENBERGHS J, BOSMANS E, DEBOUTTE D, KENIS G, MAES M. Activation of the inflammatory response system in autism. *Neuropsychobiology* 2002; 45: 1-6.
- 106) CURRAN LK, NEWSCHAFER CJ, LEE LC, CRAWFORD SO, JOHNSTON MV, ZIMMERMAN AW. Behaviors associated with fever in children with autism spectrum disorders. *Pediatrics* 2007; v120: ve1386-139.
- 107) ZERBO O, YOSHIDA C, GREYER JK, VAN DE WATER J, ASHWOOD P, DELORENZE GN, CROEN LA. Neonatal cytokines and chemokines and risk of Autism Spectrum Disorder: the Early Markers for Autism (EMA) study: a case-control study. *J Neuroinflammation*, 2014; 11: 113.
- 108) ENSTROM A, KRAKOWIAK P, ONORE C, PESSAH IN, HERTZ-PICCIOTTO I, HANSEN RL, VAN DE WATER JA, ASHWOOD P. Increased IgG4 levels in children with autism disorder. *Brain Behav Immun* 2009; 23: 389-395.
- 109) SINISCALCO D. Current findings and research prospective in autism spectrum disorders. *Autism* 2013; S2: e001.
- 110) MOSTAFA GA, EL-SAYED ZA, ABD EL AZIZ MM, EL-SAYED MF. Serum anti-myelin-associated glycoprotein antibodies in Egyptian autistic children. *J Child Neurol* 2008; 23: 1413-1418.
- 111) MOSTAFA GA, AL-AYADHI LY. Increased serum levels of anti-ganglioside M1 auto-antibodies in autistic children: relation to the disease severity. *J Neuroinflammation* 2011; 8: 39.
- 112) MOSTAFA GA, KITCHENER N. Serum anti-nuclear antibodies as a marker of autoimmunity in Egyptian autistic children. *Pediatr Neurol* 2009; 40: 107-112.
- 113) VOJDANI A, CAMPBELL AW, ANYANWU E, KASHANIAN A, BOCK K, VOJDANI E. Antibodies to neuron-specific antigens in children with autism: possible cross-reaction with encephalitogenic proteins from milk, *Chlamydia pneumoniae* and *Streptococcus* group A. *J Neuroimmunol* 2002; 129: 168-177.
- 114) JYONOUCHI H, GENG L, STRECK DL, TORUNER GA. Immunological characterization and transcription profiling of peripheral blood (PB) monocytes in children with autism spectrum disorders (ASD) and specific polysaccharide antibody deficiency (SPAD): case study. *J Neuroinflammation* 2012; 9: 4.
- 115) NORIEGA DB, SAVELKOUL HF. Immune dysregulation in autism spectrum disorder. *Eur J Pediatr* 2014; 173: 33-43.
- 116) MONTELEONE G, CARUSO R, FINA D, PELUSO I, GIOIA V, STOLFI C, FANTINI MC, CAPRIOLI F, TERSIGNI R, ALESSANDRONI L, MACDONALD TT, PALLONE F. Control of matrix metalloproteinase production in human intestinal fibroblasts by interleukin 21. *Gut* 2006; 55: 1774-1780.
- 117) TSUCHIYA KJ, HASHIMOTO K, IWATA Y, TSUJII M, SEKINE Y, SUGIHARA G, MATSUZAKI H, SUDA S, KAWAI M, NAKAMURA K, MINABE Y, YAGI A, IYO M, TAKEI N, MORI N. Decreased serum levels of platelet-endothelial adhesion molecule (PECAM-1) in subjects with high-functioning autism: a negative correlation with head circumference at birth. *Biol Psychiatry* 2007; 62: 1056-1058.
- 118) IWATA Y, TSUCHIYA KJ, MIKAWA S, MIKAWA S, NAKAMURA K, TAKAI Y, SUDA S, SEKINE Y, SUZUKI K, KAWAI M, SUGIHARA G, MATSUZAKI H, HASHIMOTO K, TSUJII M, SUGIYAMA T, TAKEI N, MORI N. Serum levels of P-selectin in

- men with high-functioning autism. *Br J Psychiatry* 2008; 193: 338-339.
- 119) GREGG JP, LIT L, BARON CA, HERTZ-PICCIOTTO I, WALKER W. Gene expression changes in children with autism. *Genomics* 2008; 91: 22-29.
- 120) PERRICONE R, PERRICONE C, DE CAROLIS C, SHOENFELD Y. NK cells in autoimmunity: a two-edged weapon of the immune system. *Autoimmun Rev* 2008; 7: 384-390.
- 121) VOJDANI A, MUMPER E, GRANPEESHEH D, MIELKE L, TRAVER D, BOCK K, HIRANI K, NEUBRANDER J, WOELLER KN, O'HARA N, USMAN A, SCHNEIDER C, HEBRONI F, BEROOKHIM J, MCCANDLESS J. Low natural killer cell cytotoxic activity in autism: the role of glutathione, IL-2 and IL-15. *J Neuroimmunol* 2008; 205:148-154.
- 122) SERBINA NV, PAMER EG. Coordinating innate immune cells to optimize microbial killing. *Immunity* 2008; 29: 672-674
- 123) PARIHAR MS, PARIHAR A, FUJITA M, HASHIMOTO M, GHAFOURIFAR P. α -Synuclein overexpression and aggregation exacerbates impairment of mitochondrial functions by augmenting oxidative stress in human neuroblastoma cells. *Int J Biochem Cell Biol* 2009; 41: 2015-2024.
- 124) ADAMS JB, JOHANSEN LJ, POWELL LD, QUIG D, RUBIN RA. Gastrointestinal flora and gastrointestinal status in children with autism--comparisons to typical children and correlation with autism severity. *BMC Gastroenterol* 2011; 11: 22.
- 125) BLACK C, KAYE JA, HERSHEL J. Relation of childhood gastrointestinal disorders to autism: Nested case-control study using data from the UK General Practice Research Database. *Br Med J* 2002; 325: 419-421.
- 126) HORVATH K, PAPADIMITRIOU JC, RABSTYAN A. Gastrointestinal abnormalities in children with autistic disorder. *Disabil Rehabil* 1999; 135: 559-563.
- 127) KUDDO T, NELSON KB. How common are gastrointestinal disorders in children with autism? *Curr Opin Pediatr* 2003; 15: 339-343.
- 128) LEVY SE, HYMAN SL. Complementary and alternative medicine treatments for children with autism spectrum disorders. *Child Adolesc Psychiatr Clin N Am* 2008; 17: 803-820.
- 129) ASHWOOD P, ANTHONY A, PELLICER AA, TORRENTE F, WALKER-SMITH JA, WAKEFIELD AJ. Intestinal lymphocyte populations in children with regressive autism: evidence for extensive mucosal immunopathology. *J Clin Immunol* 2003; 23: 504-517.
- 130) DE MAGISTRIS, PICARDI A, SINISCALCO D, RICCIO MP, SAPONE A, CARIELLO R, BRAVACCIO C. Antibodies against food antigens in patients with autistic spectrum disorders. *Biomed Res Int* 2013; 2013: 729349.
- 131) CANDELA M, BIAGI E, MACCAFERRI S, TURRONI S, BRIGIDI P. Intestinal microbiota is a plastic factor responding to environmental changes. *Trends Microbiol* 2012; 20: 385-391.
- 132) WANG Y, KASPER LH. The role of microbiome in central nervous system disorders *Brain Behav Immun* 2014; 38: 1-12.
- 133) SHAW W, KASSEN E, CHAVES E. Increased excretion of analogs of Krebs cycle metabolites and arabinose in two brothers with autistic features. *Clin Chem* 1995; 41: 1094-1104.
- 134) BOLTE, ER. Autism and Clostridium tetani. *Med Hypotheses* 1988; 51: 133-144.
- 135) SANDLER RH, FINEGOLD SM, BOLTE ER, BUCHANAN CP, MAXWELL AP, VÄISÄNEN ML, NELSON MN, WEXLER HM. Short-term benefit from oral vancomycin treatment of regressive-onset autism. *J Child Neurol* 2000; 15: 429-435.
- 136) FINEGOLD, SM, MOLITORIS D, SONG Y, LIU C, VAISANEN ML, BOLTE E, McTEAGUE M, SANDLER R, WEXLER H, MARLOWE EM, COLLINS MD, LAWSON PA, SUMMANEN P, BAYSALLAR M, TOMZYNSKI TJ, READ E, JOHNSON E, ROLFE R, NASIR P, SHAH H, HAAKE DA, MANNING P, KAUL A. Gastrointestinal microflora studies in late-onset autism. *Clin Infect Dis* 2002; 35: S6-S16.
- 137) FINEGOLD SM, DOWD SE, GONTCHAROVA V. Pyrosequencing study of fecal microflora of autistic and control children. *Anaerobe* 2010; 16: 444-453.
- 138) PARRACHO HM, BINGHAM MO, GIBSON GR, MCCARTNEY AL. Differences between the gut microflora of children with autistic spectrum disorders and that of healthy children. *J Med Microbiol* 2005; 54: 987-991.
- 139) KAGNOFF MF. Microbial-epithelial cell crosstalk during inflammation: the host response. *Ann N Y Acad Sci* 2006; 1072: 313-320.
- 140) SALLES TEIXEIRA TF, BORONI MOREIRA AP, SILVA SOUZA NC, FRIAS R, GOUVEIA PELUZIO MDO C. Intestinal permeability measurements: general aspects and possible pitfalls. *Nutr Hosp* 2014; 29: 269-281.
- 141) VIGGIANO D, IANIRO G, VANELLA G, BIBBÒ S, BRUNO G, SIMEONE G, MELE G. Gut barrier in health and disease: focus on childhood. *Eur Rev Med Pharmacol Sci* 2015; 19: 1077-1085.
- 142) ZAWISZA E. Non IgE mediated food reactions. *Alergia* 2010. Available at: http://alergia.org.pl/lek/index.php?option=com_content&task=view&id=243
- 143) PENNESI CM, KLEIN LC. Effectiveness of the gluten-free, casein-free diet for children diagnosed with autism spectrum disorder: based on parental report. *Nutr Neurosci* 2012; 15: 85-91.
- 144) KRAL TV, ERIKSEN WT, SOUDERS MC, PINTO-MARTIN JA. Eating behaviors, diet quality, and gastrointestinal symptoms in children with autism spectrum disorders: a brief review. *J Pediatr Nurs* 2013; 28: 548-556.
- 145) WHITELEY, HARACOPOS D, KNIVSBERG AM, REICHELTL KL, PARLAR S, JACOBSEN J, SHATTOCK P. The ScanBrit randomised, controlled, single-blind study of a gluten-and casein-free dietary intervention for children with autism spectrum disorders. *Nutr Neurosci* 2010; 13: 87-100.
- 146) ASHWOOD P, SCHAUER J, PESSAH IN, VAN DE WATER J. Preliminary evidence of the in vitro effects of BDE-47 on innate immune responses in children with autism spectrum disorders. *J Neuroimmunol* 2009; 208: 130-135.

- 147) WATERMAN SJ, EL-FAWWAL HA, SNYDER CA. Lead alters the immunogenicity of two neural proteins: a potential mechanism for the progression of lead-induced neurotoxicity. *Environ Health Perspect* 1994; 102: 1052-1056.
- 148) BAGENSTOSE LM, SALGAME P, MONESTIER M. Murine mercury-induced autoimmunity: a model of chemically related autoimmunity in humans. *Immunol Res* 1999; 20: 67-67.
- 149) ROWLEY B, MONESTIER M. Mechanisms of heavy metal-induced autoimmunity. *Mol Immunol* 2005; 42: 833-838.
- 150) CASIDA JE. Pest toxicology: the primary mechanisms of pesticide action. *Chem Res Toxicol* 2009; 22: 609-619.
- 151) HEUSINKVELD HJ, WESTERINK RH. Organochlorine insecticides lindane and dieldrin and their binary mixture disturb calcium homeostasis in dopaminergic PC12 cells. *Environ Sci Technol* 2012; 46: 1842-1848.
- 152) DIEL F, HÖRR B, BORCK H, IRMAN-FLORJANC T. Pyrethroid insecticides influence the signal transduction in T helper lymphocytes from atopic and nonatopic subjects. *Inflamm Res* 2003; 52: 154-163.
- 153) ZHANG Y, ZHAO M, JIN M, XU C, WANG C, LIU W. Immunotoxicity of pyrethroid metabolites in an in vitro model. *Environ Toxicol Chem* 2010; 29: 2505-2510.