

Propofol suppresses invasion, angiogenesis and survival of EC-1 cells *in vitro* by regulation of S100A4 expression

X.-G. GUO¹, S. WANG¹, Y.-B. XU², J. ZHUANG¹

¹Department of Anesthesia, Guangdong Cardiovascular Institute, Guangdong General Hospital, Guangzhou, Guangdong, China

²Department of Anesthesiology, Provincial-owned Hospital Affiliated to Shandong University, Jinan, Shandong, China

Abstract. – OBJECTIVE: Propofol possess anticancer properties in several cancers. In the present study, we investigate the effect of propofol on the human esophageal squamous cell carcinomas (ESCC) EC-1 cells *in vitro* and its molecular mechanisms of action.

MATERIALS AND METHODS: EC-1 cells were explored to 10-100 $\mu\text{mol/L}$ propofol for 72 h or 100 $\mu\text{mol/L/mL}$ propofol for 24-72 h. EC-1 cells were explored to 100 $\mu\text{mol/L}$ propofol for 24 h, then was transiently transfected into PcDNA3.1-S100A4 cDNA or PcDNA3.1 plasmid for 48 hrs. MTT, TUNEL, ELISA, migration, tube formation and immunoblotting were analyzed.

RESULTS: Propofol inhibits invasion, angiogenesis, proliferation and induces apoptosis in a dose and time-dependence manner, followed by decreased S100A4 expression by Western blot assay. Pre-transfection of PcDNA3.1-S100A4 cDNA inhibits propofol-induced apoptosis and promotes invasion and angiogenesis in EC-1 cells *in vitro*.

CONCLUSIONS: Propofol inhibited invasion, angiogenesis and induces apoptosis of human EC-1 cells *in vitro* through regulation of S100A4 expression. It not only can be an anesthesia agent, but also plays a important role of inhibiting the migration and angiogenesis of ESCC cells in the therapy of ESCC patients.

Key Words:

Esophageal squamous cell carcinomas, Propofol, metastasis; NF- κ B; S100A4.

Introduction

Propofol is a short-acting intravenous anaesthetic agent, which has been widely used in operating rooms as well as in the Intensive Care Unit (ICU). Increasing experimental evidence suggests that propofol has the neuroprotective effect against ischemic neuronal injury in animal mod-

els of cerebral ischemia^{1,2}. The mechanisms by which may be associated with apoptotic inhibition by a consequence of the regulation of Bcl-2, caspase-3 and Bax^{3,4}.

It has recently found that propofol possess anticancer properties in several cancers. In ovarian cancer OVCAR-3 cells and gallbladder cancer GBC-SD cells, propofol inhibited proliferation of OVCAR-3 and GBC-SD cells in a dose- and time-dependent manner. After exposure to propofol for 24-48 hours, these cells showed increased apoptosis and increased invasion⁵⁻⁷. Propofol also has the same anticancer properties in hepatocellular carcinoma⁸, esophageal squamous cell carcinoma⁹, osteosarcoma¹⁰, and lung adenocarcinoma¹¹. However, the propofol's molecular mechanism of action is unclear.

We have previously found propofol inhibited proliferation, invasion and angiogenesis of human esophageal squamous cell carcinomas (ESCC) Eca-109 cells *in vitro* through modulation of ERK-VEGF/MMP-9 signaling⁹. Wu et al¹¹ has reported that propofol inhibits MMP-2 and -9 mRNA and protein expressions, resulting in suppression of lung cancer cell invasion and migration *in vitro*. In breast cancer cells, propofol could inhibit the invasion and migration of breast cancer cells via regulation of NF- κ B/MMPs signal¹².

S100A4 is a member of the S100 family. It has 101 amino acids and with a molecular weight of about 11.6 kDa. S100A4 possesses a wide range of biological functions, such as regulation of angiogenesis¹³⁻¹⁵, cell survival¹⁵⁻¹⁶, motility and invasion¹⁶⁻¹⁸. S100A4 functions through regulation of NF- κ B, MMP-9 and VEGF^{13,14,18}. Previous studies have found that NF- κ B, MMP-9 and VEGF, which were related with invasion and migration of many experimental animal models,

were positively regulated by propofol⁹⁻¹². In the present study, we provide evidence to support the hypothesis that propofol inhibits invasiveness and survival of human ESCC cells through the transcriptional regulation of S100A4.

Materials and Methods

Cell Culture

The human esophageal squamous cell carcinomas (ESCC) EC-1 cells, which has high S100A4 expression¹⁹, was purchased from the American Type Culture Collection (ATCC, Shanghai, China). It was cultured in Dulbecco Modified Eagle's Medium (DMEM) supplemented with 10% fetal calf serum (FCS), nonessential amino acids, sodium pyruvate, penicillin/streptomycin antibiotics, and L-glutamine. Cells were maintained in a humidified incubator containing 10% CO₂ at 37°C. Before propofol treatment, the cells underwent serum starvation for 24 h.

Propofol Treatment

EC-1 cells were explored to 10-100 µmol/L propofol for 72 h or 100 µmol/L/mL propofol for 24-72 h. The cells were collected and processed for analysis of MTT, TUNEL, ELISA, migration, tube formation and immunoblotting.

S100A4 cDNA Transfection

PcDNA3.1-S100A4 cDNA and PcDNA3.1 plasmid was kindly gifted by Dr. Zhang, General Surgery, Affiliated Hospital of Medical College, QingDao University, QingDao¹⁹. EC-1 cells were explored to 100 µmol/L propofol for 24 h, then was transiently transfected into PcDNA3.1-S100A4 cDNA or PcDNA3.1 plasmid for 48 hrs using Lipofectamine 2000 (Invitrogen, Guangzhou, China) in accordance with the manufacturer's protocol. The cells were collected and processed for analysis of MTT, TUNEL, FACS, migration, tube formation and immunoblotting.

Western Blot

Cellular proteins from treated EC-1 cells were prepared according to the manufacturer's protocol. For routine quantitation of proteins, following the manufacturer's protocol (Pierce, Rockford, IL, USA). 40 µg of protein samples were subjected to sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) on either 12% Tris-acetate gradient gels for S100A4 detection. After gel electrophoresis and transfer

to nitrocellulose, the membranes were stained. Membranes were incubated at 4°C overnight in a blocking solution containing 5% bovine skim milk and 0.1% Tween 20 (Fischer Scientific, Pittsburgh, PA, USA) in TBS (10 mM Tris-HCl with 150 mM NaCl, pH 7.6), then probed with specific primary and secondary antibodies conjugated to horseradish peroxidase (HRP). Immunoreactive bands were visualized by chemiluminescence solution and exposure to X-ray film.

TUNEL Assay for Apoptosis

Apoptosis of the EC-1 cells treated with propofol alone or combined with PcDNA3.1-S100A4 cDNA and PcDNA3.1 plasmid transfection was evaluated by the terminal transferase dUTP nick end labeling (TUNEL) assay according to the manufacturer's instructions. TUNEL-positive cells was assessed in three randomly selected fields each section. All assays were performed in quadruplicate.

ELISA for Apoptosis

Apoptosis of the EC-1 cells treated with propofol alone or combined with PcDNA3.1-S100A4 cDNA and PcDNA3.1 plasmid transfection was evaluated by the Cell Apoptosis ELISA Detection Kit according to the manufacturer's protocol. The spectrophotometric absorbance of the samples was determined using ULTRA Multifunctional Microplate Reader (Tecan, San Jose, CA, USA) at 405 nm.

MTT for Cell Growth Inhibition Assay

EC-1 cells treated with propofol alone or combined with PcDNA3.1-S100A4 cDNA and PcDNA3.1 plasmid transfection were seeded at a density of 3×10^3 cells per well in 96-well microtiter culture plates. After overnight incubation, 20 µL of MTT solution (5 mg/mL in phosphate buffered saline – PBS) were added to each well and incubated further for 2 h. Upon termination, the supernatant was aspirated and the MTT formazan formed by metabolically viable cells was dissolved in 100 µL of isopropanol. The plates were mixed for 30 min on a gyratory shaker, and absorbance was measured at 595 nm using a plate reader.

Chicken Chorioallantoic Membrane (CAM) Angiogenesis Assay

EC-1 cells treated with propofol alone or combined with PcDNA3.1-S100A4 cDNA and PcDNA3.1 plasmid transfection for the indicated

time. The conditioned medium was filtered off for future research. White Leghorn eggs were incubated with 70% humidity at 37°C. To investigate the effect of propofol on tumor-induced angiogenesis, the conditioned medium above were placed onto the CAM (10-day-old); After placement of the discs, embryos were transferred back into the incubator. After 3 days, embryos were examined under a stereomicroscope for evaluation of angiogenesis. Blood vessels density was quantified by Image J software and represented as a bar diagram.

Cell Invasion Assay

EC-1 cells treated with propofol alone or combined with PcDNA3.1-S100A4 cDNA and PcDNA3.1 plasmid transfection for the indicated time were trypsinized and resuspended in DMEM and 0.1% bovine serum albumin (BSA) at 6×10^5 cells/ml. A 250 μ l aliquot of the suspension was applied into the inner chamber of a cell invasion plate assembly in a QCM 24-well cell invasion assay kit (Chemicon International, Billerica, MA, USA). The cells were incubated at 37°C for 24 h. Invaded cells were fluorimetrically detected according to the manufacturer's instruction.

Statistical Analysis

All statistical analyses were performed using the SPSS11.0 software (SPSS Inc., Chicago, IL, USA). The results were used as means $\pm$ SD of three replicate assays. Differences between different groups were assessed using ANOVA or also post hoc test. $p < 0.05$ was considered as statistical significance.

Results

Propofol Inhibits S100A4 Expression in EC-1 Cell in a Dose- and Time-Dependent Manner

S100A4 was overexpressed under normal growth conditions in EC-1 cells by Western blot assay. EC-1 cells treated with 10-100 μ mol/L propofol for 72 h significantly reduced the levels of S100A4 in EC-1 cells in a concentration-dependent fashion (Figure 1A). Furthermore, EC-1 cells treated with 100 μ mol/L propofol for 24-72 h also significantly reduced the levels of S100A4 in EC-1 cells in a time-dependent fashion (Figure 1B).

Effect of Propofol on Cell Proliferation

EC-1 cells were exposure to 10-100 μ mol/L propofol for 72 h, and the cell proliferation was detected by MTT analysis. Treatment with 10, 50, and 100 μ mol/L/mL of propofol for 72 h resulted in a dose-dependent inhibition of cell proliferation (Figure 2A). Treated with 100 μ mol/L concentrations of propofol for 24-72 h also resulted in a time-dependent inhibition of cell proliferation (Figure 2B).

Propofol Induces EC-1 cell Apoptosis in a dose and Time-Dependent Manner

EC-1 cells were treated with 0-100 μ mol/L propofol over 72 h, or 100 μ mol/L propofol for 0-72 h. TUNEL and ELISA technique was used to detect cell apoptosis. Treatment with propofol (10-100 μ mol/L/mL) for 72 h resulted in significant apoptosis relative to control by TUNEL assay (Figure 3A). Similarly results were also found when treated with propofol (100 μ mol/L/mL) for 24-72 h (data not shown). The treatment of EC-1 cells with propofol resulted in a dose- and time-dependent promotion of cell apoptosis, suggesting that propofol was an effective promoyion of EC-1 cell death as a single agent. ELISA assay (Figure 3B) has the same results as the TUNEL results.

Propofol Inhibits Invasion and Angiogenesis in EC-1 Cell

We first analyzed the effect of propofol treatment on the invasive capability of EC-1 cells. As

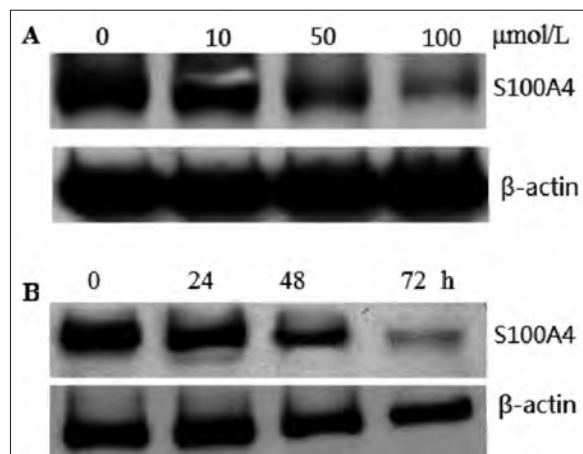


Figure 1. Effect of Propofol on S100A4 expression in EC-1 cells. **A**, EC-1 cells treated with 10-100 μ mol/L propofol for 72 h, Western blot assay was used to detect S100A4 expression in the EC-1 cells. **B**, EC-1 cells treated with 100 μ mol/L propofol for 24-72 h, Western blot assay was used to detect S100A4 expression in the EC-1 cells.

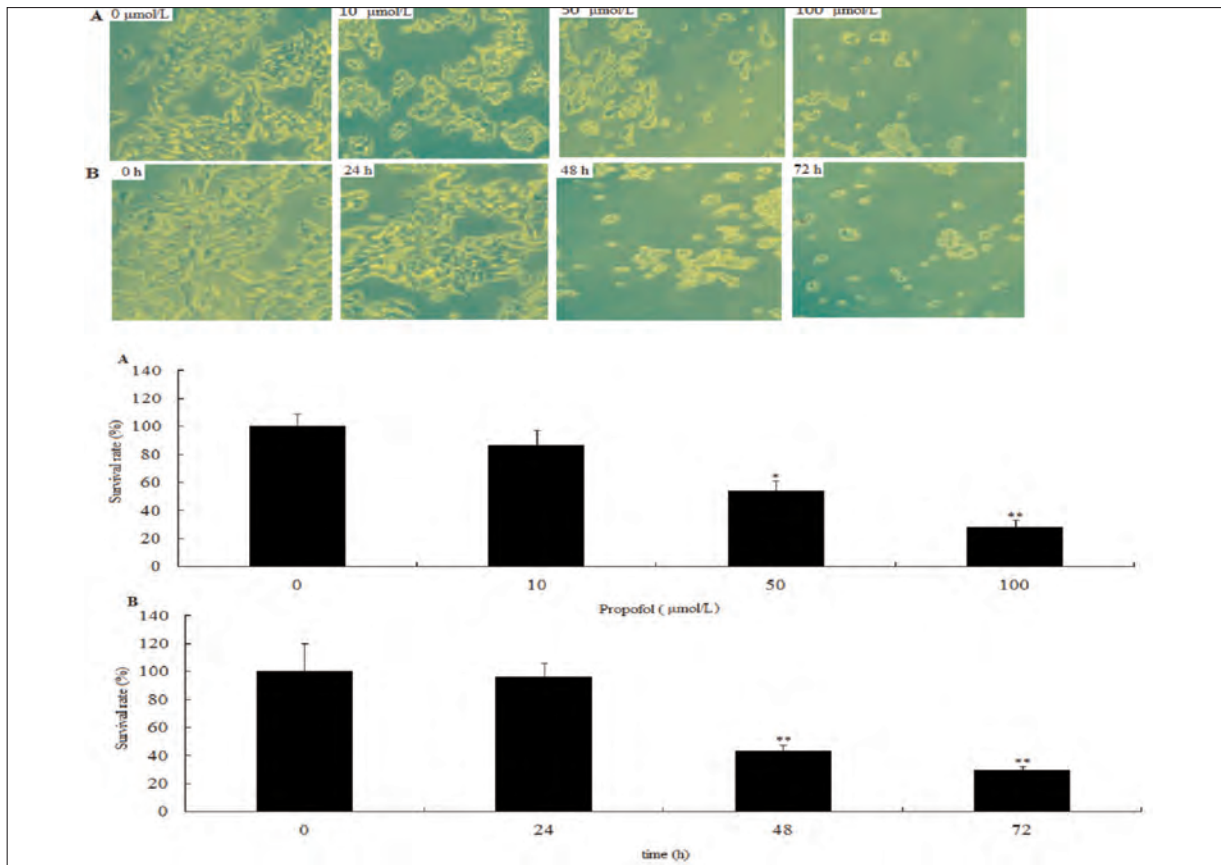


Figure 2. Effect of Propofol on survival in EC-1 cells. **A**, EC-1 cells treated with 10-100 µmol/L propofol for 72 h. **B**, EC-1 cells treated with 100 µmol/L propofol for 24-72 h. MTT assay was used to detect survival in EC-1 cells. vs. control * $p < 0.05$, ** $p < 0.01$ statistical significance.

shown in Figure 4A. treatment with 100 µmol/L propofol significantly decreased ($p < 0.05$) the number of invasive cells. Next we quantified the effect of propofol treatment on the angiogenic ability in CAM (chick chorioallantoic membrane). The images shows the presence of few scattered blood vessels around the sponge when treated with 100 µmol/L propofol in EC-1 cells compared to EC-1 cells alone on CAM. (Figure 4B). These data further support our hypothesis that propofol inhibits the invasive characteristics during human ESCC development.

Propofol-Mediated S100A4-Dependent Growth, Invasion, Angiogenesis and Apoptosis in EC-1 cells

Propofol (100 µmol/L) inhibits angiogenesis and invasion, followed by S100A4 inhibition (Figure 1). When S100A4 cDNA was transfected into the EC-1/propofol cells, S100A4 expression was significantly increased (Figure 4C), and the

effect of propofol on angiogenesis and invasion was increased (Figure 4).

EC-1 cells were treated with 0-100 µmol/L propofol over 72 h, or 100 µmol/L propofol for 0-72 h, resulted in a dose and time-dependent inhibition of cell proliferation (Figure 2), and increase of cell apoptosis (Figure 2), followed by decreased S100A4 expression (Figure 1). When S100A4 cDNA was transfected into the EC-1/propofol cells, propofol-induced apoptosis in the EC-1 cells was significantly decreased (Figure 5C-D), and survival rate was significantly increased (Figure 5A-B).

Discussion

We investigated the effects of propofol on the behavior of human ESCC cells and the role of S100A4 in these effects. We showed that propofol inhibits invasion, angiogenesis, proliferation

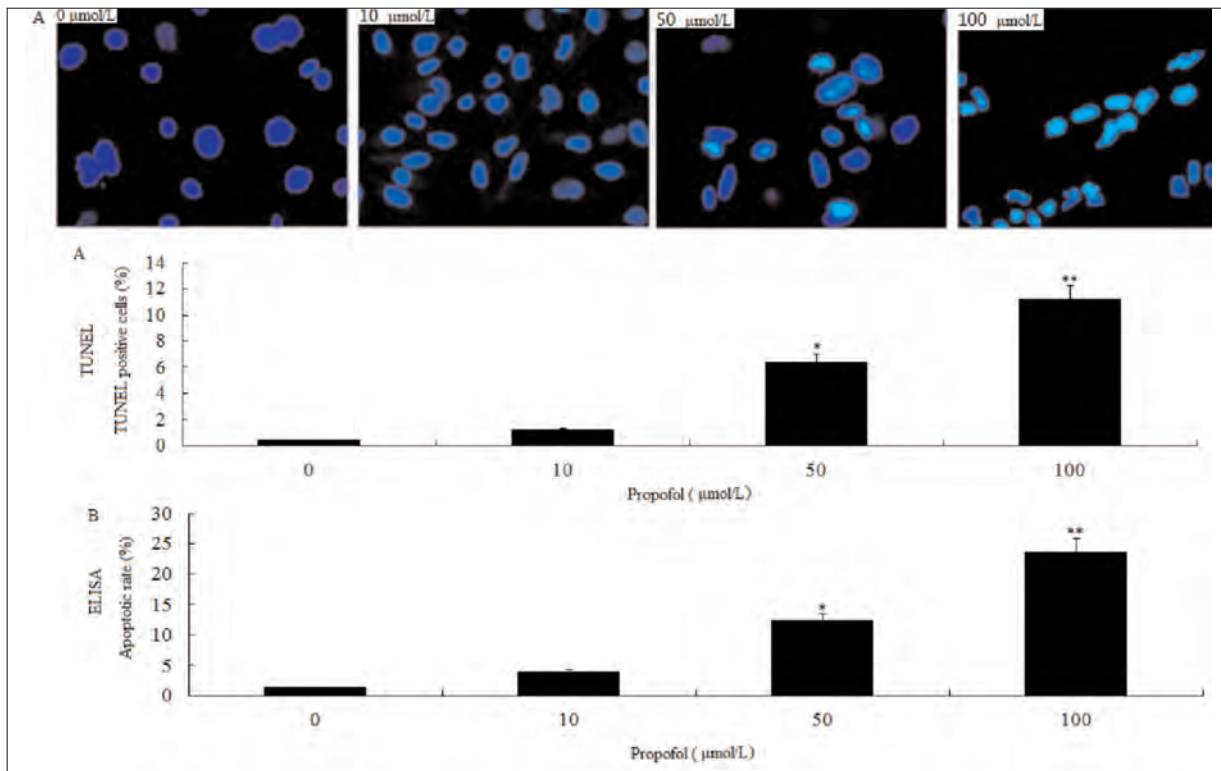


Figure 3. TUNEL and ELISA assay was used to evaluate apoptosis in EC-1 cells. **A**, Propofol-induced apoptotic cell death by TUNEL after 72 h of treatment with 10-100 μmol/L propofol. **B**, Propofol-induced apoptotic cell death by ELISA after 72 h of treatment with 10-100 μmol/L propofol. vs. control, * $p < 0.05$, ** $p < 0.01$.

and induces apoptosis of EC-1 cells through inhibition of S100A4.

Propofol is extensively used as a sedative and general anesthetic for surgery. Additionally, recent studies demonstrated that propofol exerts anti-cancer effects by inducing cancer cell apoptosis²⁰⁻²¹. Propofol may directly and indirectly suppress the viability and proliferation of cancer cells by promoting apoptosis in human lung cancer and neuroblastoma cells²²⁻²³. We have previously found that treating pancreatic cancer cells (MIA-PaCa-2) with propofol in concentrations of 10, 25, 50, and 100 mmol/mL resulted in dose- and time-dependent promotions of apoptosis. It was a single effective promoter of the death of pancreatic cancer cells²⁴. Wang et al⁷ has reported that treatment with propofol induced apoptosis and increased paclitaxel killing of all paclitaxel-sensitive and -resistant ovarian cancer cells followed by significant decrease in the Slug levels. Treatment with propofol inhibits invasion and migration. We also found that propofol inhibited proliferation, invasion and angiogenesis of human Eca-109 cells *in vitro* through modulation of ERK-VEGF/MMP-9 signaling⁹.

Our data showed that propofol-induced apoptosis and survival inhibition in EC-1 cells with high S100A4 expression occurs in a dose and time-dependent manner and significantly rises at the concentration of 100 μmol/L propofol treatment for 72 hours. We also found in the present study that treatment with 100 μmol/L propofol significantly decreased the number of invasive cells, and decreased the angiogenic ability in CAM (chick chorioallantoic membrane).

Studies in rodents²⁵ have provided evidence supporting the direct involvement of S100A4 in tumor progression and metastasis. Based on observations in transgenic mice, S100A4 has been identified as a potent stimulator of angiogenesis²⁶. Otherwise, both extracellular and intracellular S100A4 also participates in the regulation of cell death²⁷.

In the present study, we found that treatment with propofol inhibits S100A4 expression in a dose- and time-dependent manner in the EC-1 cells. Although EC-1 cells treated with propofol resulted in a dose and time-dependent inhibition of cell proliferation, and increase of cell apoptosis. When S100A4 cDNA was transfected into

Propofol, EC-1 cells and regulation of S100A4 expression

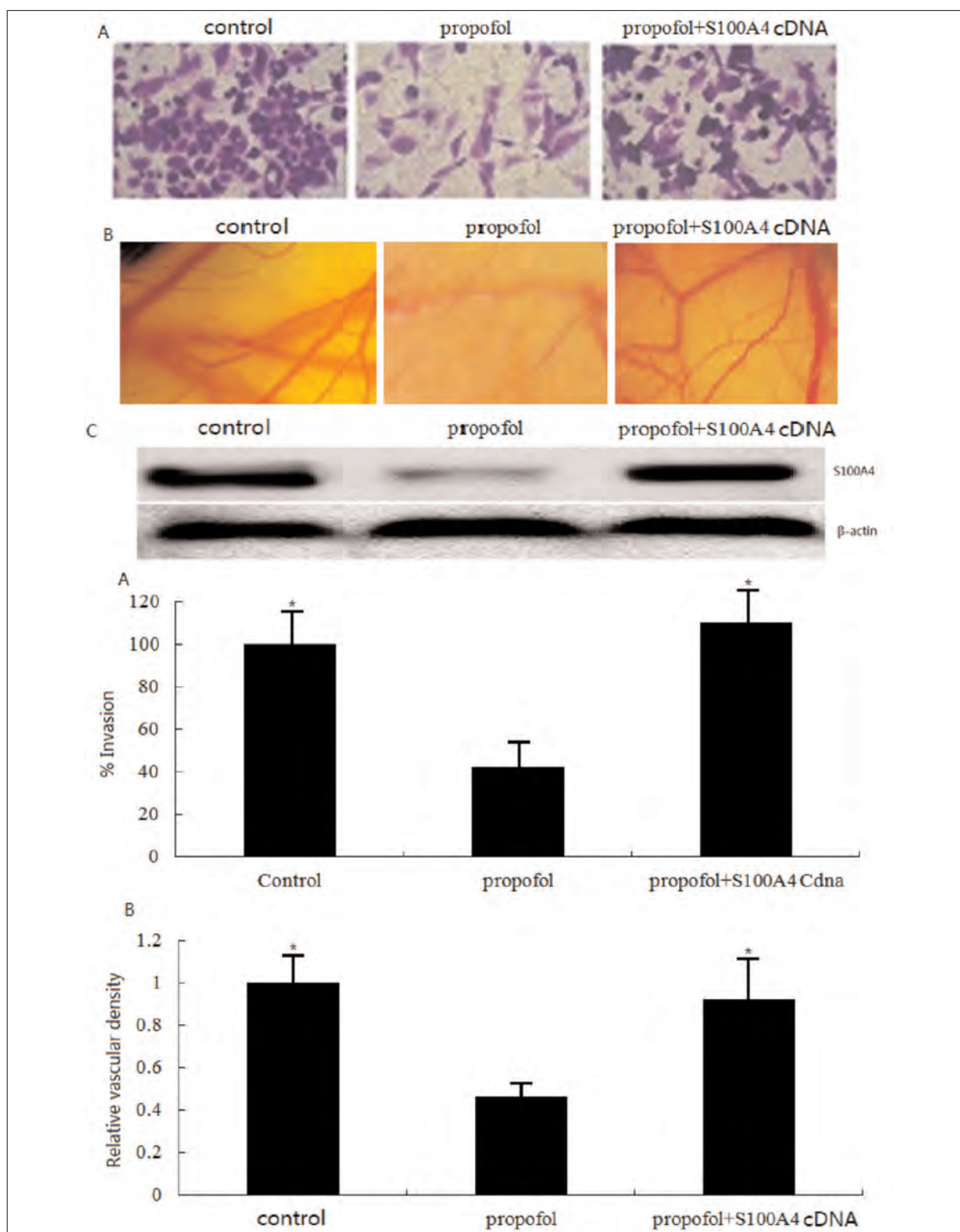


Figure 4. Propofol inhibited angiogenesis by CAM and invasion in matrigel assay. **A**, Propofol inhibits *in vitro* invasion by matrigel assay. **B**, Propofol inhibits *in vivo* angiogenesis through CAM assay. Blood vessels density was quantified. Values are means $\pm$ SD. **C**, Western blot assay for S100A4 in S100A4 cDNA transfected EC-1 cells. Vs. propofol, $*p < 0.05$. Although propofol inhibits angiogenesis and invasion, when S100A4 cDNA was transfected into the EC-1 cells, the effect of propofol on angiogenesis and invasion was restored.

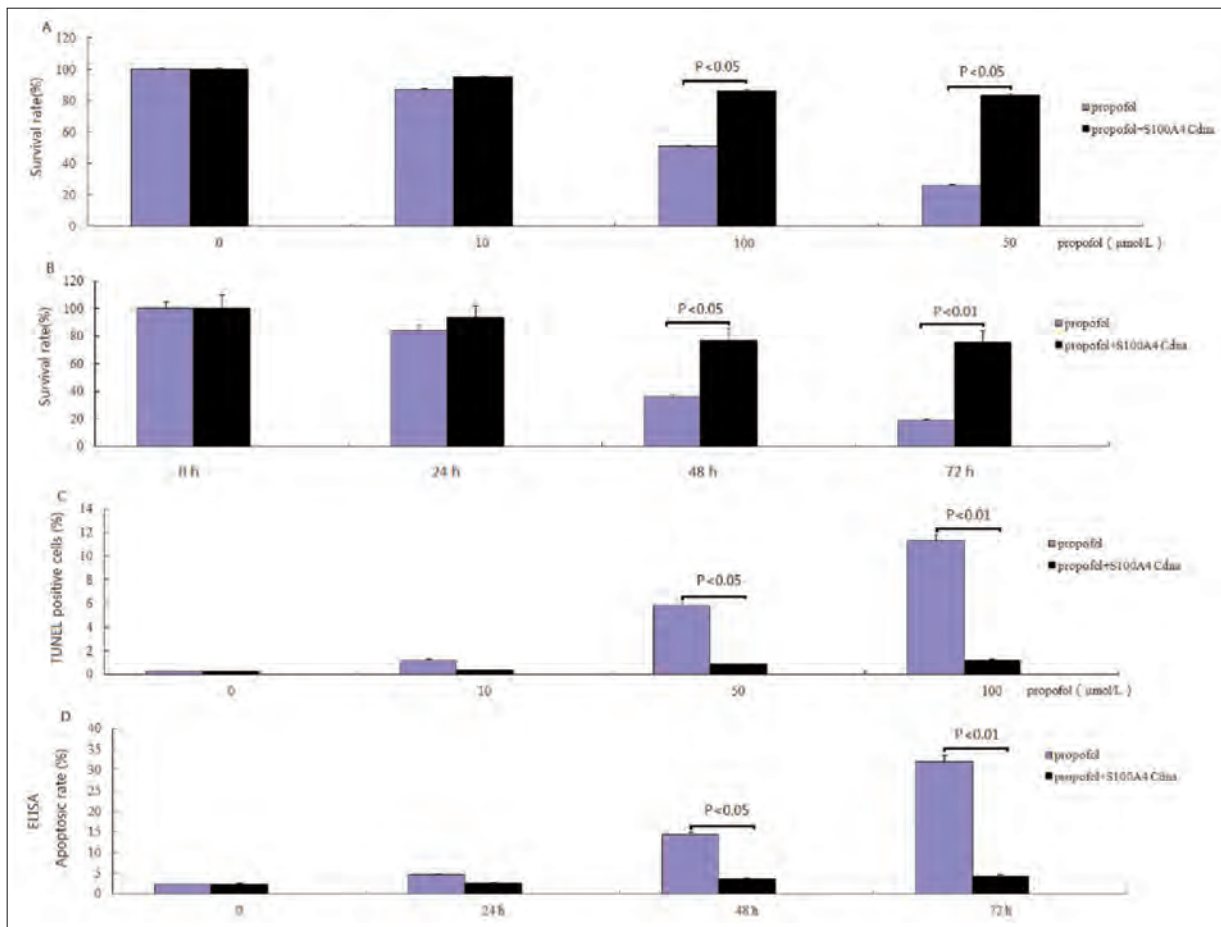


Figure 5. Effect of S100A4 on propofol-induced apoptosis and survival. S100A4 cDNA was transfected into the EC-1 cells, then treated with 0-100 $\mu\text{mol/L}$ propofol over 72 h, or 100 $\mu\text{mol/L}$ propofol for 0-72 h. **A, B,** MTT assay was used to detect survival in EC-1 cells. **C-D:** TUNEL and ELISA assay was used to evaluate apoptosis in EC-1 cells.

the EC-1/propofol cells, propofol-induced apoptosis in the EC-1 cells was significantly decreased, and survival rate was significantly increased. Chicken chorioallantoic membrane (CAM) angiogenesis assay and cell invasion assay also showed that S100A4 cDNA was transfected into the EC-1 cells, the effect of propofol on angiogenesis and invasion was restored.

Conclusions

This study provides new insights into effect of propofol on behavior of ESCC cells and the related mechanism. Our present study suggests that propofol inhibits invasion, angiogenesis, proliferation and induces apoptosis of EC-1 cells through inhibition of S100A4. However, this should be verified in further studies, including animal trials and prospective clinical studies.

Acknowledgements

This work was supported by Science and Technology Planning Project of Guangdong Province, China (No: 2014B031800005).

Conflict of Interest

The Authors declare that there are no conflicts of interest.

References

- 1) FAN W, ZHU X, WU L, WU Z, LI D, HUANG F, HE H. Propofol: an anesthetic possessing neuroprotective effects. *Eur Rev Med Pharmacol Sci* 2015; 19: 1520-1529.
- 2) CUI D, WANG L, QI A, ZHOU Q, ZHANG X, JIANG W. Propofol prevents autophagic cell death following oxygen and glucose deprivation in PC12 cells and cerebral ischemia-reperfusion injury in rats. *PLoS One* 2012; 7: e35324.

- 3) XI HJ, ZHANG TH, TAO T, SONG CY, LU SJ, CUI XG, YUE ZY. Propofol improved neurobehavioral outcome of cerebral ischemia-reperfusion rats by regulating Bcl-2 and Bax expression. *Brain Res* 2011; 1410: 24-32.
- 4) LI J, HAN B, MA X, QI S. The effects of propofol on hippocampal caspase-3 and Bcl-2 expression following forebrain ischemia-reperfusion in rats. *Brain Res* 2010; 1356: 11-23.
- 5) SU Z, HOU XK, WEN QP. Propofol induces apoptosis of epithelial ovarian cancer cells by upregulation of microRNA let-7i expression. *Eur J Gynaecol Oncol* 2014; 35: 688-691.
- 6) ZHANG L, WANG N, ZHOU S, YE W, JING G, ZHANG M. Propofol induces proliferation and invasion of gallbladder cancer cells through activation of Nrf2. *J Exp Clin Cancer Res* 2012; 31: 66.
- 7) WANG P, CHEN J, MU LH, DU QH, NIU XH, ZHANG MY. Propofol inhibits invasion and enhances paclitaxel-induced apoptosis in ovarian cancer cells through the suppression of the transcription factor slug. *Eur Rev Med Pharmacol Sci* 2013; 17: 1722-1729.
- 8) ZHANG J, WU GQ, ZHANG Y, FENG ZY, ZHU SM. Propofol induces apoptosis of hepatocellular carcinoma cells by upregulation of microRNA-199a expression. *Cell Biol Int* 2013; 37: 227-232.
- 9) XU YB, DU QH, ZHANG MY, YUN P, HE CY. Propofol suppresses proliferation, invasion and angiogenesis by down-regulating ERK-VEGF/MMP-9 signaling in Eca-109 esophageal squamous cell carcinoma cells. *Eur Rev Med Pharmacol Sci* 2013; 17: 2486-2494.
- 10) YE Z, JINGZHONG L, YANGBO L, LEI C, JIANDONG Y. Propofol inhibits proliferation and invasion of osteosarcoma cells by regulation of microRNA-143 expression. *Oncol Res* 2013; 21: 201-207.
- 11) WU KC, YANG ST, HSIA TC, YANG JS, CHIOU SM, LU CC, WU RS, CHUNG JG. Suppression of cell invasion and migration by propofol are involved in down-regulating matrix metalloproteinase-2 and p38 MAPK signaling in A549 human lung adenocarcinoma epithelial cells. *Anticancer Res* 2012; 32: 4833-4842.
- 12) LI Q, ZHANG L, HAN Y, JIANG Z, WANG Q. Propofol reduces MMPs expression by inhibiting NF- κ B activity in human MDA-MB-231 cells. *Biomed Pharmacother* 2012; 66: 52-56.
- 13) JIA W, GAO XJ, ZHANG ZD, YANG ZX, ZHANG G. S100A4 silencing suppresses proliferation, angiogenesis and invasion of thyroid cancer cells through downregulation of MMP-9 and VEGF. *Eur Rev Med Pharmacol Sci* 2013; 17: 1495-1508.
- 14) ABU EL-ASRAR AM, NAWAZ MI, DE HERTOIGH G, ALAM K, SIDDIQUEI MM, VAN DEN EYNDE K, MOUSA A, MOHAMMAD G, GEBOES K, OPDENAKKER G. S100A4 is upregulated in proliferative diabetic retinopathy and correlates with markers of angiogenesis and fibrogenesis. *Mol Vis* 2014; 20: 1209-1224.
- 15) OCHIYA T, TAKENAGA K, ENDO H. Silencing of S100A4, a metastasis-associated protein, in endothelial cells inhibits tumor angiogenesis and growth. *Angiogenesis* 2014; 17: 17-26.
- 16) ZHANG W, LIU Y, WANG CW. S100A4 promotes squamous cell laryngeal cancer Hep-2 cell invasion via NF- κ B/MMP-9 signal. *Eur Rev Med Pharmacol Sci* 2014; 18: 1361-1367.
- 17) LI HJ, CHEN YX, WANG Q, ZHANG YG. S100A4 mRNA as a prognostic marker and therapeutic target in Wilms tumor (WT). *Eur Rev Med Pharmacol Sci* 2014; 18: 817-827.
- 18) ZHANG J, ZHANG DL, JIAO XL, DONG Q. S100A4 regulates migration and invasion in hepatocellular carcinoma HepG2 cells via NF- κ B-dependent MMP-9 signal. *Eur Rev Med Pharmacol Sci* 2013; 17: 2372-2382.
- 19) ZHANG M, ZHAO H, YAN B, ZHANG D, LIANG J. S100A4 regulates motility and invasiveness of human esophageal squamous cell carcinoma through modulating the AKT/Slug signal pathway. *Dis Esophagus* 2012; 25: 731-739.
- 20) WANG ZT, GONG HY, ZHENG F, LIU DJ, YUE XQ. Propofol suppresses proliferation and invasion of gastric cancer cells via downregulation of microRNA-221 expression. *Genet Mol Res* 2015; 14: 8117-8124.
- 21) SU Z, HOU XK, WEN QP. Propofol induces apoptosis of epithelial ovarian cancer cells by upregulation of microRNA let-7i expression. *Eur J Gynaecol Oncol* 2014; 35: 688-691.
- 22) CUI WY, LIU Y, ZHU YQ, SONG T, WANG QS. Propofol induces endoplasmic reticulum (ER) stress and apoptosis in lung cancer cell H460. *Tumour Biol* 2014; 35: 5213-5217.
- 23) NAKAJIMA A, TSUJI M, INAGAKI M, TAMURA Y, KATO M, NIYA A, USUI Y, OGUCHI K. Neuroprotective effects of propofol on ER stress-mediated apoptosis in neuroblastoma SH-SY5Y cells. *Eur J Pharmacol* 2014; 725: 47-54.
- 24) DU QH, XU YB, ZHANG MY, YUN P, HE CY. Propofol induces apoptosis and increases gemcitabine sensitivity in pancreatic cancer cells in vitro by inhibition of nuclear factor- κ B activity. *World J Gastroenterol* 2013; 19: 5485-5492.
- 25) LLOYD BH, PLATT-HIGGINS A, RUDLAND PS, BARRACLOUGH R. Human S100A4 (p9Ka) induces the metastatic phenotype upon benign tumour cells. *Oncogene* 1998; 17: 465-473.
- 26) OCHIYA T, TAKENAGA K, ENDO H. Silencing of S100A4, a metastasis-associated protein, in endothelial cells inhibits tumor angiogenesis and growth. *Angiogenesis* 2014; 17: 17-26.
- 27) MAHON PC, BARIL P, BHAKTA V, CHELALA C, CAULEE K, HARADA T, LEMOINE NR. S100A4 contributes to the suppression of BNIP3 expression, chemoresistance, and inhibition of apoptosis in pancreatic cancer. *Cancer Res* 2007; 67: 6786-6795.