

Hsa-miR-337 inhibits non-small cell lung cancer cell invasion and migration by targeting TCF7

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Abstract. – OBJECTIVE: Recent studies have revealed that microRNAs (miRNAs) play a crucial role in the progression of tumorigenesis. Non-small cell lung cancer (NSCLC) is one of the most common malignancies worldwide. The aim of this study was to identify the exact role of hsa-miR-337 in the progression of NSCLC and to investigate the possible underlying mechanism.

PATIENTS AND METHODS: Hsa-miR-337 expression in NSCLC cells and 60 paired tissue samples were detected by Real Time-quantitative Polymerase Chain Reaction (RT-qPCR). Moreover, the functions of hsa-miR-337 were identified by transwell assay and wound healing assay, respectively. Furthermore, the underlying mechanism was explored by RT-qPCR, Western blot assay, and luciferase assay.

RESULTS: The expression level of hsa-miR-337 in NSCLC tissues was remarkably down-regulated when compared with that of adjacent normal samples. Moreover, the expression and function of NSCLC cells were significantly inhibited after overexpression of hsa-miR-337 *in vitro*. Moreover, after overexpression of hsa-miR-337 *in vitro*, the mRNA and protein levels of TCF7 were significantly down-regulated. Besides, the expression of TCF7 in NSCLC tissues was negatively correlated with the expression of hsa-miR-337.

CONCLUSIONS: The above results suggested that hsa-miR-337 could repress the migration and invasion of NSCLC cells through directly targeting TCF7. Furthermore, hsa-miR-337 might offer a new therapeutic intervention for NSCLC patients.

Keywords:
MicroRNA, Hsa-miR-337, NSCLC, TCF7.

Introduction

Lung cancer is one of the most common causes of cancer-related death worldwide, still serving as a threat to public health¹. As the main type of lung

cancer, non-small cell lung cancer (NSCLC) accounts for more than 85% of all lung cancer cases². In the past decades, the diagnosis and treatment of NSCLC have been developed greatly. However, the prognosis of NSCLC patients remains unsatisfactory. The majority of NSCLC patients are diagnosed in an advanced stage, and the 5-year overall survival rate of patients is less than 15%³. Early detection is emerging as the most important step for therapeutic strategy, particularly in NSCLC. The reason for this is that there is a lack of clinical biomarkers to identify novel biomarkers and therapeutic targets for patients with NSCLC.

MicroRNAs (miRNAs) are a subtype of non-coding RNAs with 18 to 22 nucleotides in length. Several studies have showed that miRNAs play a crucial role in the regulation of many biological behaviors, including cell proliferation, apoptosis, and metastasis in diverse malignancies. For example, activated by K-Ras carcinogenic signal, miRNA-155 facilitates the proliferation of pancreatic cancer cells by regulating reactive oxygen species (ROS) stress⁴. MiR-126 plays an important role in breast cancer by interacting with a variety of molecules. This may eventually help to inhibition of breast cancer cell metastasis⁴. Up-regulation of miR-378 represses the proliferation, migration, and invasion of colon cancer cells by inhibiting epithelial-mesenchymal transition⁵. In addition, miR-532 enhances the invasion of gastric cancer cells *via* activating Wnt/beta-catenin pathway and inhibiting naked cuticle 1 (NKD1). Furthermore, miR-532 may provide a novel therapeutic target for gastric cancer⁶. However, no literature has elucidated the role of hsa-miR-337 in NSCLC metastasis as well as its potential molecular mechanism.

In our study, we found that hsa-miR-337 was lowly expressed in NSCLC tissue samples. Meanwhile, hsa-miR-337 significantly inhibited NSCLC metastasis *in vitro*. Furthermore, we explored the underlying mechanism of how hsa-miR-337 functioned in NSCLC metastasis.

Patients and Methods

Tissue Specimens

Totally 60 NSCLC patients who underwent surgical resection at The First Affiliated Hospital of Dalian Medical University were enrolled in this study. Human NSCLC tissues and adjacent non-tumor tissues were collected from patients during the surgery. After surgical resection, all tissue samples were snap-frozen in liquid nitrogen immediately for subsequent use. No radiotherapy and chemotherapy treatment were conducted in any patient before the surgery. The study was approved by the Research Ethics Committee of The First Affiliated Hospital of Dalian Medical University. The written informed consents were obtained from all the patients before the study.

Cell Culture and Transfection

Five NSCLC cell lines (A549, SPCA1, H1975, H1299, and PC-9) and one immortalized human bronchial epithelial cell line (HBE) were bought from American Type Culture Collection (ATCC; Manassas, VA, USA). All cell lines were cultured in Dulbecco's Modified Eagle's Medium (DMEM; Invitrogen, Carlsbad, CA, USA) or Roswell Park Memorial Institute 1640 (RPMI-1640) medium (Invitrogen, Carlsbad, CA, USA). According to the manufacturer's protocol of lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA), miRNA mimics (hsa-miR-337; GenePharma, Shanghai, China) was transfected into cells.

RNA Extraction and Real Time-Quantitative Polymerase Chain Reaction (RT-qPCR)

Total RNA from tissues and cells was extracted according to the instructions of TRIzol reagent (Invitrogen, Carlsbad, CA, USA). Complementary deoxyribose nucleic acids (cDNAs) was synthesized in strict accordance with Reverse qPCR Kit (Takara Biotechnology Co., Ltd., Dalian, China). SYBR Green Real Time-PCR Master Mix (Roche, Basel, Switzerland) and ABI ViiA7 qPCR System (ABI, Foster City, CA, USA) was used to

perform RT-qPCR. The mRNA expression level of hsa-miR-337 was detected. Primers used in this study were as follows: hsa-miR-337, forward 5'-CGCTTCAGCTCCTATATGA-3' and reverse 5'-GTGCAGGGTCCGAGGT-3'; U6, forward 5'-CTCGCTTCGGCAGCACA-3' and reverse 5'-AACGCTTCACGAATTTGCG-3'; TCF7, forward 5'-CTCGAGCCTACCCCAAGT-GACA-3' and reverse 5'-TTTAAAGAG-GCTTTGAAAAACAAA-3'; Glyceraldehyde 3-phosphate dehydrogenase (GAPDH), forward 5'-TGTGGGCATCAATGTTTCT-3' and reverse 5'-ACACCATGTAATCCGTAAT-3'. Specific thermal cycle was 30 sec at 95°C, 5 min at 95°C for 40 cycles, 35 sec at 60°C.

Wound Healing Assay

Six-well plates were used to culture NSCLC cells. miRNA mimics were transfected into cells. When growing to about 90% of confluence, cells were scratched by a sterile 10 µL pipette tip. Then, the cells were incubated in a humidified incubator with 5% CO₂ atmosphere at 37°C. Open wound area was measured at 24 h. These procedures were repeated for three times.

Tube Formation Assays

24 h after transfection, 2×10^5 cells in 100 µL of serum-free DMEM were seeded into the upper chamber of 8-µm culture inserts (Corning, Corning, NY, USA). Then, 20% of fetal bovine serum (FBS)-DMEM (Invitrogen, Carlsbad, CA, USA) was added to the lower chamber of culture inserts. After culturing for 24 h, the cells were fixed with methanol for 30 min and stained with hematoxylin for 20 min. Finally, migrated cells were observed under an inverted microscope ($\times 20$), and the number of cells was calculated. Five fields were randomly selected for each sample.

Western Blotting Analysis

48 h after transfection, cells were first collected. Radioimmunoprecipitation assay (RIPA) protein extraction reagent (Beyotime, Shanghai, China) supplemented with protease inhibitor cocktail (Roche, Basel, Switzerland) and phenylmethylsulfonyl fluoride (Roche, Basel, Switzerland) was used to lyse cells. Subsequently, total proteins were separated by 10% Sodium Dodecyl Sulphate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) and transferred onto 0.22-µm NC membranes. The membranes were incubated with specific primary and secondary antibodies. An-

ti-GAPDH and anti-TCF7 were purchased from Cell Signaling Technology, Inc. (Danvers, MA, USA).

Luciferase Reporter Gene Assay

The 3'-Untranslated Region (3'-UTR) of TCF7 was cloned into a pGL3 vector as wild-type (WT) 3'-UTR. Site-directed mutagenesis of hsa-miR-337 binding site in TCF7 3'-UTR was used as mutant (MUT) 3'-UTR by quick-change site-directed mutagenesis kit (Stratagene, La Jolla, CA, USA). Then, the plasmids were transfected into A549 cells for 48 h. Dual luciferase reporter assay system was used for performing luciferase reporter gene assay.

Statistical Analysis

Statistical Product and Service Solutions (SPSS) 18.0 (SPSS, Chicago, IL, USA) statistical software was used for all statistical analysis. The independent-sample *t*-test was used to compare the difference between the two groups. *p*-values < 0.05 were considered statistically significant.

Results

Hsa-MiR-337 Was Lowly Expressed in NSCLC Tissue Specimens and Cell Lines

First, RT-qPCR was conducted to detect hsa-miR-337 expression in 60 patients' tissues and 4 NSCLC cell lines. Results showed that hsa-miR-337 was significantly down-regulated in tumor tissues than that in adjacent tissues (Figure 1A). Compared with 16HBE cells, expression of hsa-miR-337 in NSCLC cells was significantly lower (Figure 1B).

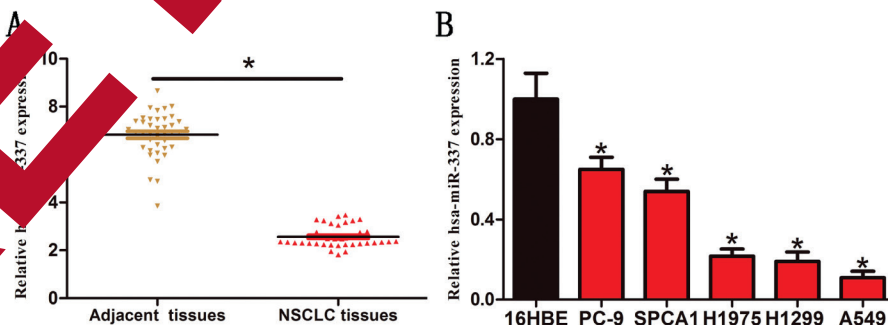


Figure 1. Expression level of hsa-miR-337 was significantly decreased in NSCLC tissues and cell lines. **A**, Hsa-miR-337 expression was significantly decreased in NSCLC tissues when compared with adjacent normal tissues. **B**, Expression level of hsa-miR-337 in human NSCLC cell lines and normal gastric epithelial cell line (A549) was determined by RT-qPCR. Data were presented as mean $\pm$ standard error of the mean. **p* < 0.05.

Overexpression of Hsa-MiR-337 Inhibited Migration and Invasion of NSCLC Cells

According to hsa-miR-337 expression in NSCLC cells, A549 cells were chosen for overexpression of hsa-miR-337. Hsa-miR-337 mimics and negative control (NC) were synthesized and transfected into A549 cells. Subsequently, hsa-miR-337 expression was verified by RT-qPCR (Figure 2A). Wound healing assay results showed that overexpression of hsa-miR-337 inhibited cell migration (Figure 2B). Moreover, the results of the transwell assay demonstrated that the overexpression of hsa-miR-337 significantly inhibited the invasion of NSCLC cells (Figure 2C).

Hsa-MiR-337 Inhibited Tumorigenesis of NSCLC via Targeting TCF7

Starbase v2.0 online database predicted that TCF7 was a possible target of hsa-miR-337 (Figure 3A). Luciferase reporter gene assay showed that significantly less luciferase activity was observed in A549 cells co-transfected with hsa-miR-337 mimics and WT-TCF7-3'-UTR when compared with control group (Figure 3B). Results of Western blot assay showed that the protein expression of TCF7 was remarkably down-regulated after hsa-miR-337 mimics transfection (Figure 3C).

Correlation Between Expression of TCF7 and Hsa-MiR-337

RT-qPCR results further demonstrated that the mRNA expression of TCF7 was significantly up-regulated in NSCLC cells when compared with 16HBE cells (Figure 4A). Meanwhile, TCF7 expression in NSCLC tissues increased markedly when compared with that of adjacent tissues (Figure 4B). Furthermore, the results of linear correlation analysis indicated that in NSCLC tissues,

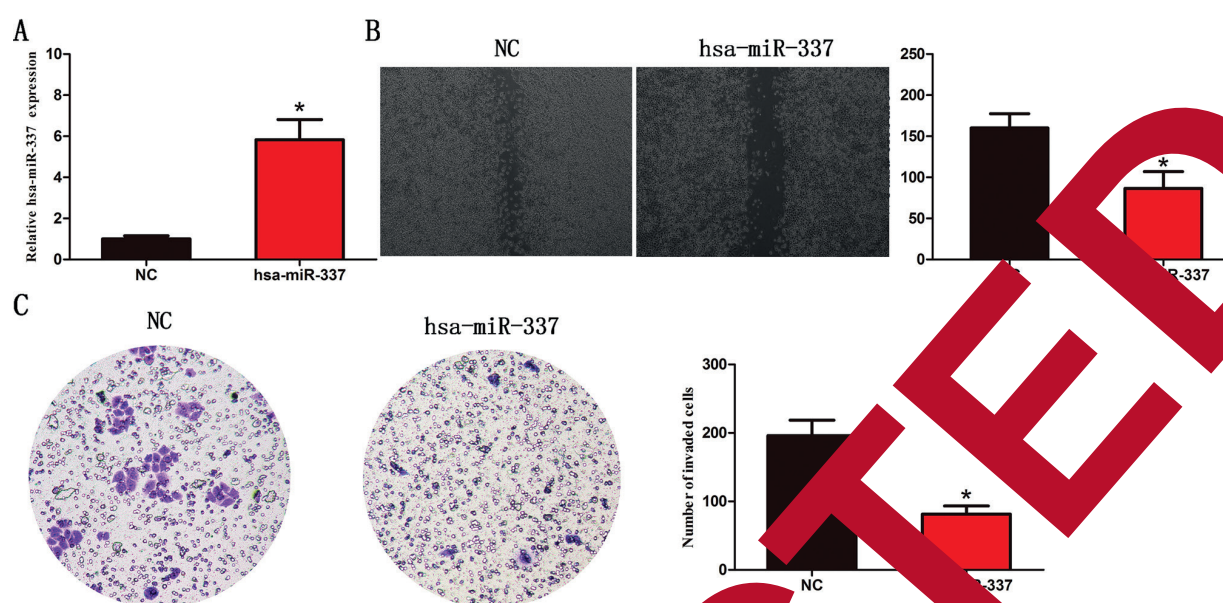


Figure 2. Overexpression of hsa-miR-337 inhibited migration and invasion of A549 NSCLC cells. **A**, Hsa-miR-337 expression in cells transfected with negative control (NC) or hsa-miR-337 mimics (hsa-miR-337) was detected by RT-qPCR. **B**, Wound healing assay showed that the overexpression of hsa-miR-337 significantly suppressed the migration ability of hsa-miR-337 cells compared with NC cells. **C**, Transwell assay demonstrated that the number of invaded cells was significantly decreased in hsa-miR-337 transfected cells compared with NC cells. All results represented the average of three independent experiments (mean $\pm$ standard error of the mean). * $p < 0.05$, as compared with NC cells.

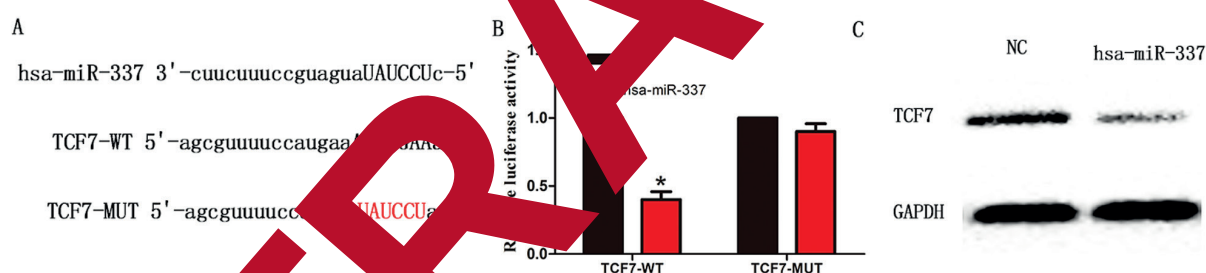


Figure 3. Hsa-miR-337 repressed NSCLC cell metastasis process via inhibiting TCF7. **A**, The binding sites of hsa-miR-337 on TCF7. **B**, Luciferase reporter gene assay showed that cells transfected with hsa-miR-337 mimics exhibited less luciferase activity than those transfected with NC. **C**, Hsa-miR-337 mimics repressed the protein expression of TCF7 in A549 NSCLC cells. * $p < 0.05$.

the expression of TCF7 was negatively correlated with hsa-miR-337 expression (Figure 4C).

Discussion

The regulation of miRNA activity leads to changes in homeostasis and various pathological processes. Researches have proved that miRNAs play important regulatory functions in carcinogenesis of NSCLC. For example, Zhao B. et al⁸ have shown that over-expression of lncRNA PCAT-1 promotes the proliferation and metastasis

of NSCLC. Xue et al⁹ have reported that through down-regulating SOCS1, SOCS6, and PTEN, miR-21 and miR-155 promote the progression of NSCLC. Meanwhile, this is inversely correlated with the prognosis of NSCLC patients. In addition, Zhao L. et al¹⁰ have found that miR-149 suppresses the growth and metastasis of NSCLC cells via depression of FOXM1/cyclin D1/MMP2 axis. Moreover, Wang et al¹¹ have proved that miR-142-5p inhibits the proliferation of NSCLC through targeting PIK3CA.

MiR-337 is reported to play an important role in the progression of diverse carcinomas. For ex-

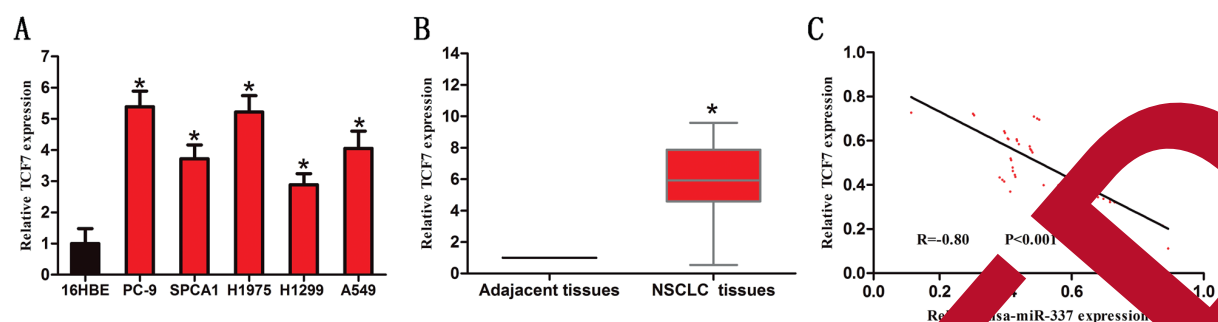


Figure 4. TCF7 expression in NSCLC tissues and NSCLC cell lines. **A**, TCF7 was highly expressed in NSCLC cell lines compared with 16HBE cells. **B**, TCF7 expression in NSCLC tissues was significantly higher than that of corresponding non-tumor tissues. **C**, Correlation analysis revealed that hsa-miR-337 expression was negatively correlated with TCF7 expression in NSCLC tissues. * $p < 0.05$.

ample, miR-337 inhibits the development of melanoma by targeting STAT3. This indicates that miR-337 may serve as a potential therapeutic target for the melanoma¹². MiR-337, functioning as an oncogene, promotes the proliferation of endometrial carcinoma by depressing PTEN expression¹³. MiR-337 acts as a tumor suppressor in the progression of colorectal cancer through targeting KRAS directly and repressing the AKT and Wnt pathways¹⁴. Moreover, miR-337 exerts a suppression effect on pancreatic ductal adenocarcinoma through regulating cell proliferation and invasion by targeting HOXB7¹⁵. In this work, hsa-miR-337 was found significantly down-regulated in both NSCLC tissues and cells. Furthermore, after hsa-miR-337 overexpression, the ability of cell migration and invasion was significantly suppressed. These data indicated that hsa-miR-337 functioned as a tumor suppressor and inhibited the tumorigenesis of NSCLC.

Transcription factor (TCF7) plays an important role in promoting the progression of diverse cancers. For instance, through activation of Wnt signaling and up-regulation of TCF7, miR-143a represses bone metastasis in prostate cancer activated by Ras¹⁶. The overexpression of miR-682 suppresses the proliferation and promotes the apoptosis of human osteosarcoma by targeting TCF7¹⁷. MiR-6852 functions as an anti-oncogene in colorectal cancer. Meanwhile, miR-6852 suppresses tumor progression and metastasis through targeting TCF7, which may serve as a potential prognostic marker and therapeutic target for colorectal cancer¹⁸. In addition, by binding to TCF7 in Wnt signaling pathway, AF1q enhances cell proliferation, migration, and chemoresistance in breast cancer¹⁹. In our study,

results of RT-qPCR and Western blot analysis indicated that TCF7 was significantly down-regulated after hsa-miR-337 overexpression *in vitro*. What's more, TCF7 was remarkably up-regulated in NSCLC samples when compared with that of adjacent tissues. A negative correlation was found between the expression of TCF7 and hsa-miR-337 in NSCLC tissues. The above results revealed that hsa-miR-337 might realize its function *via* targeting TCF7.

Conclusions

Our research indicates that hsa-miR-337 plays a vital role in the carcinogenesis of NSCLC, which can serve as a promising biomarker for NSCLC.

Conflict of interest

The authors declare no conflicts of interest.

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