

# Long noncoding RNA UCA1 promotes proliferation and metastasis of thyroid cancer cells by sponging miR-497-3p

H. GAO<sup>1</sup>, J.-Y. YANG<sup>2</sup>, L.-X. TONG<sup>3</sup>, H. JIN<sup>1</sup>, C.-Z. LIU<sup>1</sup>

<sup>1</sup>Department of Thyroid Head and Neck Surgery, Tumor Hospital of Jinlin Province, Changchun, China

<sup>2</sup>Department of Anesthesiology, Tumor Hospital of Jinlin Province, Changchun, China

<sup>3</sup>Department of Ultrasonography, Tumor Hospital of Jinlin Province, Changchun, China

**Abstract. – OBJECTIVE:** Recently, long non-coding RNAs (lncRNAs) have attracted much attention for their roles in tumor progression. The aim of this study was to investigate the exact role of lncRNA UCA1 in the development of thyroid cancer (TC) and to explore the possible underlying mechanism.

**PATIENTS AND METHODS:** UCA1 expression in both TC cells and tissues was detected by quantitative Real Time-Polymerase Chain Reaction (qRT-PCR). Colony formation assay, proliferation, and transwell assay were conducted *in vitro*. Subsequent luciferase reporter gene assay was applied to investigate the underlying mechanism. Furthermore, the function of UCA1 *in vivo* was monitored as well.

**RESULTS:** UCA1 expression in TC samples was significantly higher than that of corresponding normal tissues. After UCA1 was knocked down *in vitro* and *in vivo*, the proliferation, migration, and invasion of TC cells were significantly inhibited. Moreover, the expression of miR-497-3p was increased after the knockdown of UCA1. Furthermore, miR-497-3p was directly targeted by UCA1.

**CONCLUSION:** Knockdown of UCA1 could inhibit TC cell proliferation and metastasis via sponging miR-497-3p. Our findings might offer a new therapeutic intervention for TC patients.

**Key Words:** Long noncoding RNA, UCA1, Thyroid cancer (TC), miR-497-3p.

## Introduction

Thyroid cancer (TC) is one of the most prevalent cancers in the world. Over the past decades, the morbidity of TC has greatly increased worldwide. Currently, TC remains the eighth most common cancer in China<sup>1</sup>. In recent years, a huge development has been made in the effective treat-

ment of TC. However, due to its unsatisfied survival rate, it brings a substantial burden to patients and society<sup>2</sup>. Therefore, it is urgent to identify novel biomarkers for an early diagnosis of TC.

Long non-coding RNAs (lncRNA) are a subclass of non-protein-coding RNAs with more than 200 nucleotides in length. Authors have indicated that lncRNAs serve as major contributors in carcinogenesis, including cell apoptosis, cell proliferation, and cell metastasis. LncRNA LINC01133 promotes colorectal cancer metastasis through the regulation of the epithelial-mesenchymal transition (EMT)<sup>3</sup>. By regulating vasculogenic angiogenesis, lncRNA MALAT1 is reported to promote tumorigenicity and cell metastasis in gastric cancer<sup>4</sup>. Meanwhile, lncRNA MALAT1 accelerates the migration and invasion of hepatocellular carcinoma cells *via* targeting miR-204<sup>5</sup>. Besides, lncRNA CRNDE-h<sup>7</sup> has been reported to be a novel serum biomarker for colorectal cancer. However, the clinical role and biological mechanism of lncRNA UCA1 in the development of TC have not been fully elucidated.

In this study, we found that the expression of UCA1 was remarkably up-regulated in TC tissues. The knockdown of UCA1 significantly inhibited the proliferation and metastasis of TC cells. Moreover, our further experiment explored the underlying mechanism of UCA1 function in TC development.

## Patients and Methods

### Cell Lines and Clinical Samples

54 paired TC tissues and adjacent non-tumor tissues were sequentially gathered from TC patients undergoing surgery in the Tumor Hos-

pital of Jinlin Province from February 2017 to December 2018. Written informed consent was obtained from each patient before the operation. No radiotherapy or chemotherapy was applied for any patient before the operation. The tissues obtained from the surgery were stored immediately at  $-80^{\circ}\text{C}$  for use. All collected tissues were analyzed by an experienced pathologist. This investigation was approved by the Ethics Committee of Tumor Hospital of Jinlin Province.

### Cell Culture

K1, TPC-1, SW579 and Nthy-ori 3-1 (normal human thyroid cell line) cells were offered by the Chinese Academy of Science (Shanghai, China). All cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM; Gibco, Rockville, MD, USA) consisting of 10% of fetal bovine serum (FBS; Life Technologies, Gaithersburg, MD, USA) and penicillin. Besides, the cells were maintained in an incubator with 5%  $\text{CO}_2$  at  $37^{\circ}\text{C}$ .

### Cell Transfection

Lentiviral small hairpin RNA (shRNA) targeting UCA1 was synthesized and cloned into pCDH3-EF1a-EGFP-F2A-Puro vector (Biosset, San Diego, CA, USA). Subsequently, synthesized shRNA was transfected into TPC-1 TC cells according to the instructions of Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA).

### Quantitative Real Time Polymerase Chain Reaction (qRT-PCR)

Total RNA in tissues and cells was extracted by using TRIzol Reagent (Invitrogen, Carlsbad, CA, USA). Subsequently, the extracted total RNA was reverse-transcribed into complementary deoxyribose nucleic acids (cDNAs) through the reverse Transcription Kit (Kodak, KaRa Biotechnology Co., Ltd., Dalian, China). Primers used for qRT-PCR were as follows: UCA1 forward 5'-TTTGCCAGGACAGCTTAAT-3', reverse 5'-TTGTC-CCCCTTCCGCTAT-3'; glyceraldehyde 3-phosphate dehydrogenase (GAPDH) forward 5'-CCAGAGATGGCAATGCTGG-3' and reverse 5'-TGGCATGGACTGTGGTCA-3'. The thermo-cycling condition was as follows: 30 s at  $95^{\circ}\text{C}$ , 5 s for 40 cycles at  $95^{\circ}\text{C}$ , and 30 s at  $60^{\circ}\text{C}$ .

### Tumor Formation Assay

TPC-1 ( $1.5 \times 10^3$  cells/well) cells were first seeded into 6-well plates. Ten days later, formed colonies were fixed with 10% of formaldehyde

for 30 min and stained with 0.5% of crystal violet for 5 min. CANON camera was used to take photographs of colonies, and Image J Plus (Silver Springs, MD, USA) was used for data analysis.

### Cell Proliferation Assay

TC cells were first seeded into 96-well plates at a density of  $1 \times 10^3$  cells/well. Subsequently, 10  $\mu\text{L}$  Cell Counting Kit-8 (CCK-8; Dojindo, Kumamoto, Japan) was added into cells at different time points, followed by incubation for 2 h in the dark. Absorbance at 490 nm was measured by a microplate reader (Bio-Rad, Hercules, CA, USA).

### Transwell Assay

For detecting the TC cell migration,  $2 \times 10^5$  transfected cells in 100  $\mu\text{L}$  serum-free DMEM were transformed into the upper chamber of an 8- $\mu\text{m}$  culture insert (Corning, Corning, NY, USA). Meanwhile, 20% of FBS-DMEM was added to the lower chamber of the insert. 24 h later, these inserts were treated with methanol for 30 min and stained with hematoxylin for 20 min. The number of migrating cells was counted by an inverted microscope ( $\times 20$ ). The three fields were randomly selected for each sample. For detecting the TC cell invasion,  $2 \times 10^5$  transfected cells in 100  $\mu\text{L}$  serum-free DMEM were transformed into the upper chamber of an 8- $\mu\text{m}$  culture insert (Corning, Corning, NY, USA) coated with 50  $\mu\text{g}$  of Matrigel (BD Biosciences, Franklin Lakes, NJ, USA). Meanwhile, 20% of FBS-DMEM was added to the lower chamber of the culture insert. 24 h later, these inserts were treated with methanol for 30 min and stained with hematoxylin for 20 min. The number of invaded cells was counted by an inverted microscope ( $\times 20$ ). The three fields were randomly selected for each sample.

### Xenograft Model

For the tumor formation assay, the transfected TPC-1 cells were subcutaneously injected into NOD/SCID mice (4-5 weeks old). Tumor diameters were detected every 5 days after inoculation. Tumor volume was calculated as the following formula: volume = length  $\times$  width<sup>2</sup>  $\times$  1/2. The mice were sacrificed and tumors were extracted after 4 weeks. For the tumor metastasis assay, transfected TPC-1 cells were injected into the tail vein of NOD/SCID mice (4-5 weeks old). The mice were sacrificed and lung tissues were extracted after 4 weeks. The number of metastat-

ic nodules in the lung was then counted. Animal experiments in this study were approved by the Animal Ethics Committee of the Tumor Hospital of Jinlin Province.

### Luciferase Reporter Gene Assay

The 3'-UTR of UCA1 was first cloned into the pGL3 vector (Promega, Madison, WI, USA). Site-directed mutagenesis of the miR-497-3p binding site in UCA1 3'-UTR was performed using the QuickChange site-directed mutagenesis Kit (Stratagene, La Jolla, CA, USA). Subsequently, the cells were transfected with UCA1 WT-3'-UTR or UCA1 MUT-3'-UTR and miR-ctrl or miR-497-3p for 48 h. Finally, the dual-luciferase reporter assay system (Promega, Madison, WI, USA) was utilized to detect the luciferase activity.

### Statistical Analysis

Statistical Product and Service Solutions (SPSS) 20.0 (SPSS, Chicago, IL, USA) was adopted for all statistical analysis. Data were presented as mean  $\pm$  SD (Standard Deviation). Chi-square test and Student's *t*-test were selected when appropriate.  $p < 0.05$  was considered statistically significant.

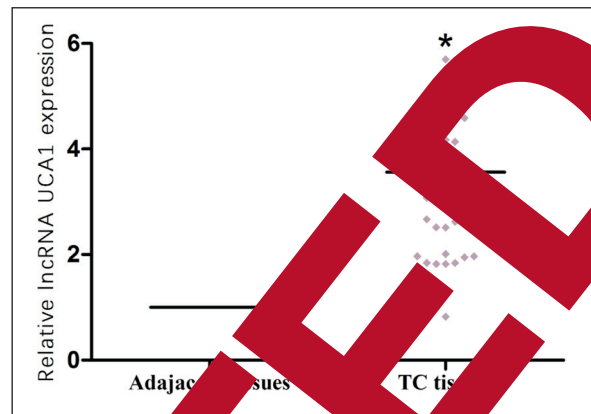
## Results

### UCA1 Expression Level in TC Tissues and Cell Lines

First, the qRT-PCR was conducted to detect UCA1 expression in 5 TC tissues and 5 TC cell lines. As a result, UCA1 was significantly up-regulated in TC tissues when compared with adjacent normal tissues (Figure 1). UCA1 expression in TC cells was significantly higher than that of Nthy-ori2-1 cells as well (Figure 2A).

### Knockdown of UCA1 Inhibited Growth of TC Cells

Following UCA1 expression, TPC-1 TC cell line was chosen for the knockdown of UCA1. QRT-PCR was used to detect UCA1 expression in cells (Figure 2B). Subsequent CCK-8 assay revealed that after UCA1 was knocked down in TC cells, the cell proliferation was remarkably suppressed (Figure 2C). Furthermore, the colony formation assay revealed that after UCA1 was knocked down, the number of cell colonies of TC cells decreased significantly (Figure 2D).



**Figure 1.** Expression level of UCA1 increased significantly in TC tissues. UCA1 expression markedly increased in TC tissues when compared with the corresponding normal tissues. Data were presented as mean  $\pm$  standard error of the mean ( $n = 5$ ).

### Knockdown of UCA1 Inhibited Migration and Invasion of TC Cells

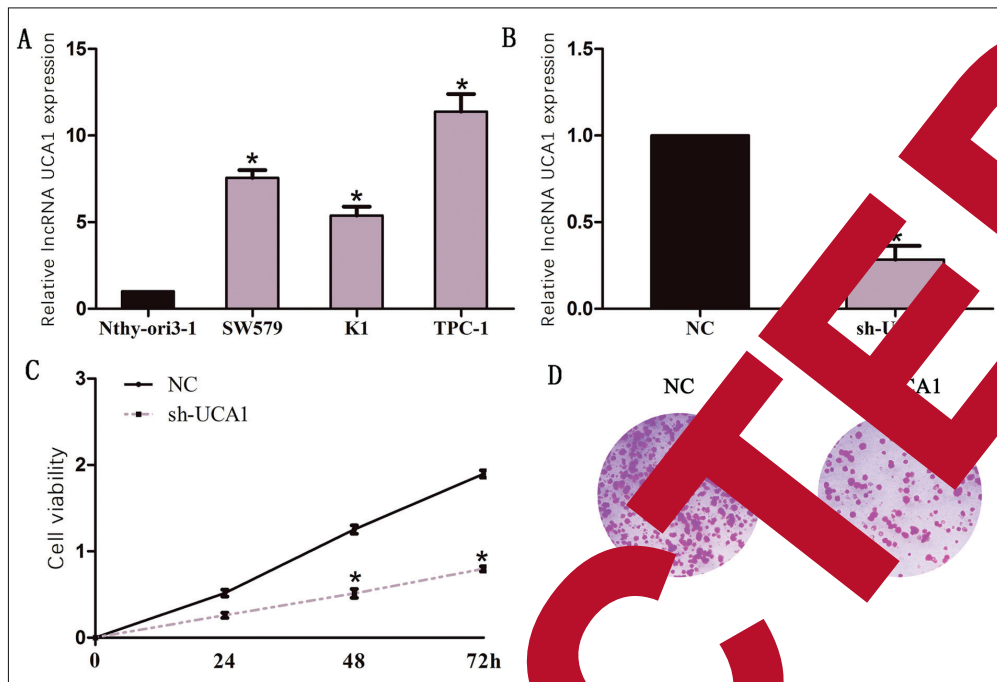
When performed transwell assay and found that the knockdown of UCA1 significantly inhibited TC cell migration (Figure 3A). Moreover, the results of transwell assay demonstrated that the invasion of TC cells was remarkably inhibited after UCA1 knock-down *in vitro* (Figure 3B).

### UCA1 Promoted TC Tumorigenesis via miR-497-3p

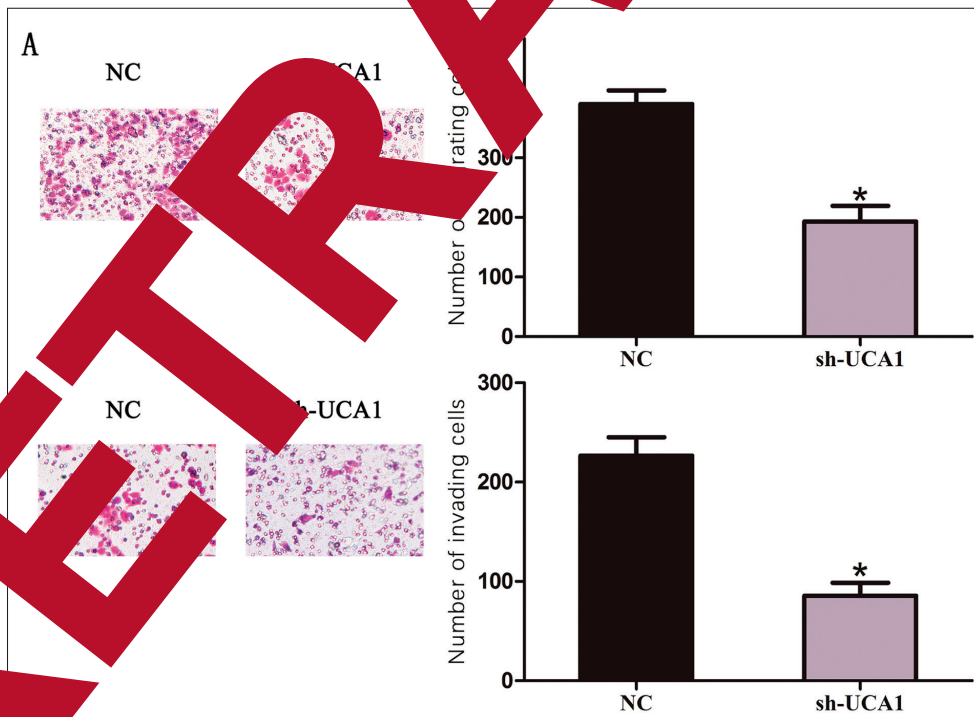
Starbase v2.0 (<http://starbase.sysu.edu.cn/mirLncRNA.php>) was used to search for miRNAs that contained complementary bases with UCA1. Then, we selected miR-497-3p which contained the binding area of UCA1 (Figure 4A), as it was reported to suppress the tumorigenesis of various tumors. QRT-PCR results showed that significantly up-regulated miR-497-3p was observed in UCA1 shRNA group when compared with negative control group (Figure 4B). Furthermore, the luciferase reporter gene assay showed that the luciferase activity was significantly reduced after co-transfection of UCA1-WT and miR-497-3p. However, no significant changes were observed in luciferase activity after the co-transfection of UCA1-MUT and miR-497-3p (Figure 4C). All these data revealed that miR-497-3p was a direct target of UCA1.

### UCA1 Knockdown Inhibited Tumor Formation and Metastasis In Vivo

The ability of UCA1 in tumor formation and metastasis was detected *in vivo*. Results indicated that tumor size in UCA1 shRNA group

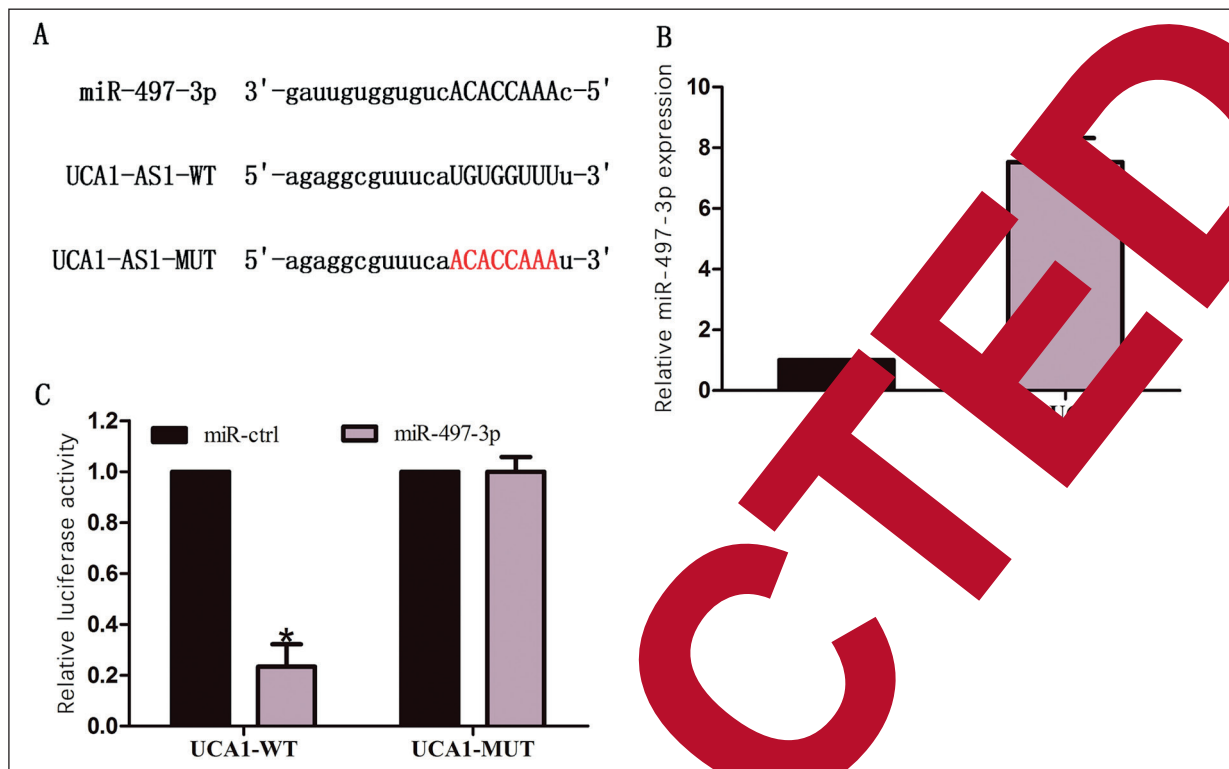


**Figure 2.** Knockdown of UCA1 inhibited TC cell proliferation. **A**, Expression levels of lncRNA UCA1 relative to GAPDH in human TC cell lines and Nthy-ori 3-1 (normal human thyroid cell line) were determined by qPCR. **B**, UCA1 expression in TC cells transduced with UCA1 shRNA (sh-UCA1) and negative control (NC) was detected by qPCR. GAPDH was used as an internal control. **C**, CCK-8 assay showed that the proliferation of TC cells was significantly inhibited via knockdown of UCA1. **D**, Colony formation assay showed that knockdown of UCA1 significantly inhibited the number of formed colonies in TC cells (magnification: 10×). The results represented the average of three independent experiments (mean ± standard error of the mean). \* $p < 0.05$ .



**Figure 3.** Knockdown of UCA1 inhibited TPC-1 TC cell migration and invasion. **A**, Transwell assay showed that knockdown of UCA1 significantly repressed the migration of TPC-1 TC cells (magnification: 40×). **B**, Transwell assay showed that knockdown of UCA1 significantly repressed the invasion of TPC-1 TC cells (magnification: 40×). The results represented the average of three independent experiments (mean ± standard error of the mean). \* $p < 0.05$ , as compared with control cells.





**Figure 4.** The association between UCA1 and miR-497-3p. **A**, The binding sites of miR-497-3p on UCA1. **B**, MiR-497-3p expression increased significantly in sh-UCA1 cells compared with NC group. **C**, Co-transfection of miR-497-3p and UCA1-WT strongly decreased luciferase activity, while co-transfection of miR-control and UCA1-WT did not change luciferase activity. The results represented the average of three independent experiments. Data were presented as mean  $\pm$  standard error of the mean. \* $p < 0.05$ , as compared with NC cells.

was significantly smaller than that of negative control group (Figure 5A). The weight of dissected tumors in UCA1 shRNA group was significantly less when compared with negative control group (Figure 5B). Meanwhile, the number of metastatic nodules in tissues of UCA1 shRNA group was significantly reduced when compared with negative control group (Figure 5C). Subsequently, the expression level of UCA1 in dissected tumor tissues was detected by RT-PCR. Results showed that UCA1 was down-regulated in UCA1 shRNA group when compared with negative control group (Figure 5D). The above results suggested that UCA1 could induce tumor formation and me-

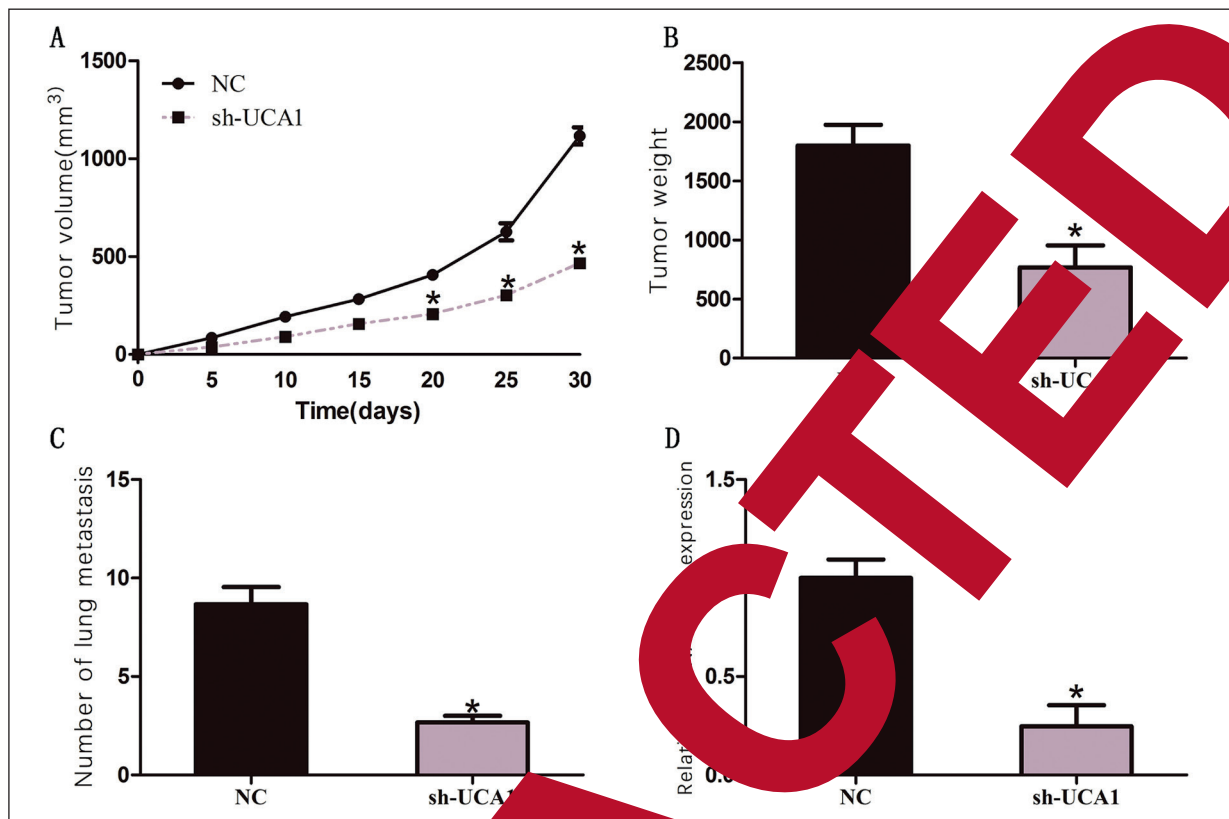
tabolism. Meanwhile, lncRNAs can be applied as potential biomarkers and therapeutic targets for TC<sup>8-10</sup>.

Located on human chromosome 19p13.12 positive strand, urothelial cancer associated 1 (UCA1) was initially discovered and investigated in bladder cancer. Previous studies<sup>11</sup> have indicated that UCA1 functions as an oncogene in bladder cancer by promoting cell proliferation and metastasis. Recently, UCA1 has emerged as a novel regulator in the initiation and progression of various cancers. Through suppressing miR-204-5p expression, UCA1 promotes cell proliferation and drug resistance in colorectal cancer<sup>12</sup>. Induced by SP1, UCA1 facilitates the proliferation of gastric cancer cells through recruiting EZH2 and activating the AKT pathway<sup>13</sup>. Furthermore, UCA1 functions as an oncogene in pancreatic cancer via promoting tumor growth and cell metastasis by sponging miR-135a<sup>14</sup>.

In this study, we found that UCA1 was significantly up-regulated in both TC tissues and cell lines. After UCA1 was knocked down, TC cell

## Discussion

lncRNAs have been reported to play crucial roles in the oncogenesis and progression of TC, including cell proliferation, motility, invasion, and



**Figure 5.** Knockdown of UCA1 inhibited tumor formation and metastasis *in vivo*. **A**, Tumor size in sh-UCA1 group was significantly smaller than that of NC group. **B**, The weight of dissected tumors in sh-UCA1 group was markedly smaller when compared with NC group. **C**, The number of metastatic nodules in lung tissues of sh-UCA1 group was significantly reduced when compared with NC group. **D**, UCA1 was lowly expressed in dissected tumors of sh-UCA1 group compared with NC group. \* $p < 0.05$ , as compared with control group.

proliferation, migration, and invasion were markedly inhibited. The above results indicated that UCA1 acted as an oncogene and promoted tumorigenesis of TC.

In recent years, the interaction between lncRNA/miRNA and protein networks has attracted much attention. Through targeting miR-20b, lncRNA CAMTA1 enhanced cell proliferation and mobility in breast cancer<sup>15</sup>. By targeting miR-211-3p, lncRNA SNHG15 enhances the proliferation, migration, and invasion of human breast cancer cells. By negatively regulating the expression of miR-429, lncRNA ILF3-AS1 facilitates the proliferation, invasion, and migration of human nasopharyngeal carcinoma cells. In this study, bioinformatics software was used to predict the possible targeted miRNAs of UCA1. MiR-497-3p, a highly conserved miRNA on the human chromosome 5.1, was selected from predicted miRNAs. It plays an important role in various cancers<sup>18-20</sup>. Our results indicated that miR-497-3p expression was significantly down-regulated after the knockdown

of UCA1. Subsequent luciferase reporter gene assay showed that miR-497-3p could directly bind to UCA1. By conducting tumor formation and metastasis assays, we found that the knockdown of UCA1 also inhibited tumor formation and metastasis *in vivo*. All the above results suggested that UCA1 might promote tumorigenesis of TC via sponging miR-497-3p.

## Conclusions

We identified that LncRNA UCA1 enhances TC cell proliferation and metastasis through sponging miR-497-3p. Our findings suggest that UCA1 may contribute to therapy for TC as a target candidate.

## Conflict of Interest

The Authors declare that they have no conflict of interests.

# References

- 1) ZHANG X, ZHANG X, CHANG Z, WU C, GUO H. Correlation analyses of thyroid-stimulating hormone and thyroid autoantibodies with differentiated thyroid cancer. *J BUON* 2018; 23: 1467-1471.
- 2) WANG Y, WANG W. Increasing incidence of thyroid cancer in Shanghai, China, 1983-2007. *Asia Pac J Public Health* 2015; 27: P223-P229.
- 3) VIGNERI R, MALANDRINO P, VIGNERI P. The changing epidemiology of thyroid cancer: why is incidence increasing? *Curr Opin Oncol* 2015; 27: 1-7.
- 4) KONG J, SUN W, LI C, WAN L, WANG S, WU Y, XU E, ZHANG H, LAI M. Long non-coding RNA LINC01133 inhibits epithelial-mesenchymal transition and metastasis in colorectal cancer by interacting with SRSF6. *Cancer Lett* 2016; 380: 476-484.
- 5) LI Y, WU Z, YUAN J, SUN L, LIN L, HUANG N, BIN J, LIAO Y, LIAO W. Long non-coding RNA MALAT1 promotes gastric cancer tumorigenicity and metastasis by regulating vasculogenic mimicry and angiogenesis. *Cancer Lett* 2017; 395: 31-44.
- 6) HOU Z, XU X, ZHOU L, FU X, TAO S, ZHOU J, TAN D, LIU S. The long non-coding RNA MALAT1 promotes the migration and invasion of hepatocellular carcinoma by sponging miR-204 and releasing SIRT1. *Tumour Biol* 2017; 39: 1010428317718135.
- 7) LIU T, ZHANG X, GAO S, JING F, YANG Y, DU J, LI G, LI P, LI C, WANG C. Exosomal long non-coding RNA CRNDE-h as a novel serum-based biomarker for diagnosis and prognosis of colorectal cancer. *Oncotarget* 2016; 7: 85551-85563.
- 8) LIU L, ZHOU XY, ZHANG JQ, WANG J, HE J, CHEN YY, HUANG C, LI L, LI SQ. LncRNA SNHG1 promotes non-small cell lung cancer cell proliferation and inhibits the apoptosis by up-regulating phosphoinositide kinase 1 (SIRT1) and its downstream PI3K/Akt pathway. *Chin J Pharmacol Sci* 2018; 22: 8722-8728.
- 9) ZHONG X, LONG W, WU S, XIAO W, HU W. LncRNA-SNHG1 promotes proliferation, apoptosis and invasion of colorectal cancer cells: assurance guidelines. *J BUON* 2018; 23: 776-781.
- 10) KHAITAN M, DINGER ME, MANNING C, CRAWFORD J, SMITH MA, MCKENNA JS, PERERA RJ. The melanoma-upregulated long noncoding RNA PRY4-IT1 modulates apoptosis and invasion. *Cancer Res* 2011; 71: 852-859.
- 11) WANG Z, LI HC, CAI JL, XU QW, LI MC, LI YC, QI J, LU TJ, YU LZ, ZHANG Y, XIN DQ, NA YQ, CHEN WF. Rapid identification of UCA1 as a very sensitive and specific marker for human bladder carcinoma. *Cancer Res* 2006; 12: 4851-4858.
- 12) BIAN Z, JIN L, ZHANG J, YIN Y, CHEN C, HU Y, FENG Y, LIU H, FEI B, MAO Y, ZHOU L, HUANG S, HUANG D, XING C, HUANG Z. LncRNA-UCA1 promotes cell proliferation and 5-fluorouracil resistance in colorectal cancer by inhibiting miR-204-5p. *Cancer* 2016; 6: 23892.
- 13) WANG ZQ, CAI Q, LI J, HE C, JI JF, QUAN ZW, LIU BY, LI C, ZHANG G. Long noncoding RNA UCA1 induced by hypoxia promotes cell proliferation via releasing miR-204 and activating AKT pathway in gastric cancer. *Cell Dis* 2017; 8: e2839.
- 14) ZHANG X, ZHOU L, WANG J, SHI G, TAN X. UCA1 regulates growth and metastasis of pancreatic cancer by sponging miR-135a. *Oncol Rep* 2017; 25: 1529.
- 15) LIU Y, LI L, WANG Y, YANG X, YANG Y. Long noncoding RNA CAMTA1 promotes proliferation and mobility of the human breast cancer cell line MDA-MB-231 by targeting miR-20b. *Oncol Res* 2018; 26: 625-631.
- 16) LIU Q, QIN J. Long noncoding RNA SNHG15 promotes human breast cancer proliferation, migration and invasion by sponging miR-211-3p. *Biochem Biophys Res Commun* 2018; 495: 1594-1600.
- 17) LIU S, ZHAO X, MA X, GAO G, YU L, YAN D, DING H, SUN W. Long noncoding RNA ILF3-AS1 promotes cell proliferation, migration, and invasion via negatively regulating miR-200b/a/429 in melanoma. *Biosci Rep* 2017; 37: pii: BSR20171031.
- 18) CHENG H, XUE J, YANG S, CHEN Y, WANG Y, ZHU Y, WANG X, KUANG D, RUAN Q, DUAN Y, WANG G. Co-targeting of IGF1R/mTOR pathway by miR-497 and miR-99a impairs hepatocellular carcinoma development. *Oncotarget* 2017; 8: 47984-47997.
- 19) SONG J, WU X, LIU F, LI M, SUN Y, WANG Y, WANG C, ZHU K, JIA X, WANG B, MA X. Long non-coding RNA PVT1 promotes glycolysis and tumor progression by regulating miR-497/HK2 axis in osteosarcoma. *Biochem Biophys Res Commun* 2017; 490: 217-224.
- 20) CHAI L, KANG XJ, SUN ZZ, ZENG MF, YU SR, DING Y, LIANG JQ, LI TT, ZHAO J. MiR-497-5p, miR-195-5p and miR-455-3p function as tumor suppressors by targeting hTERT in melanoma A375 cells. *Cancer Manag Res* 2018; 10: 989-1003.