

Possible mechanistic insights into iron homeostasis role of the action of 4-aminoquinolines (chloroquine/hydroxychloroquine) on COVID-19 (SARS-CoV-2) infection

G.E.-S. BATIHA¹, H.M. SHAHEEN¹, H.M. AL-KURAIHY², J.O. TEIBO³, O.A. AKINFÉ⁴, A.I. AL-GARBEE², T.K.A. TEIBO⁵, S.M. KABRAH⁶

¹Department of Pharmacology and Therapeutics, Faculty of Veterinary Medicine, Damanhour University, Damanhour, AlBeheira, Egypt

²Department of Clinical Pharmacology and Therapeutic Medicine, College of Medicine, Almustansiriyah University, Baghdad, Iraq

³Department of Biochemistry and Immunology, Ribeirão Preto Medical School, University of São Paulo, Ribeirão Preto, Ribeirão Preto, São Paulo, Brazil

⁴Department of Biochemistry, University of Ibadan, Ibadan, Nigeria

⁵Department of Maternal-Infant and Public Health Nursing, College of Nursing, Ribeirão Preto, University of São Paulo, Ribeirão Preto, São Paulo, Brazil

⁶Department of Laboratory Medicine, Faculty of Applied Medical Sciences, Umm Al-Qura University, Kingdom of Saudi Arabia

Abstract. – OBJECTIVE: With the recent direction in drug repurposing, many approved drugs have been evaluated to assess their effect on the coronavirus or SARS-CoV-2 infection (COVID-19). Driving this path, chloroquine (CQ) has been used in the treatment of malaria and hydroxychloroquine (HCQ) in immunomodulatory and anti-thrombotic action, playing a leading role in initial management of the viral infection.

MATERIALS AND METHODS: Literature search was done using Google Scholar, PubMed and Scopus database using keywords “chloroquine” “SARS-CoV-2” “COVID-19” “mechanism of action” and articles of interest were selected providing evidence of the possible role of CQ in viral infection.

RESULTS: In a bid to understand how and if CQ and HCQ would exert their anti-viral property, mechanistic exegesis was done to review various proposed mechanisms of action. This revealed the inhibition of viral attachment and entry, inhibition of enveloped glycoprotein, inhibition of the development and proliferation of new viral particles as the way they perform their action. There is an interplay between iron metabolism and homeostasis with COVID-19 infection and viral reproduction.

CONCLUSIONS: This study aims to show the functional role of CQ and HCQ, as well as to provide possible mechanistic insight on the role of iron on viral infection, iron starvation and its downstream cellular pathways involving hepcidin and proinflammatory cytokines. The overall

aim of providing possible mode of action of CQ and HCQ in the management of COVID-19 infection is exhibited via its anti-viral, anti-inflammatory and anti-thrombotic activities.

Key Words:

COVID-19, Chloroquine/Hydroxychloroquine, Iron homeostasis, Mechanism of action, SARS-CoV-2.

Introduction

In the treatment of malaria in the last few decades, chloroquine (CQ) and various analogs such as hydroxychloroquine (HCQ) and 4-aminoquinolones have played frontline roles (Figure 1). However, this has been replaced with newer antimalarial drugs due to drug-resistant strains of *Plasmodium falciparum*. The immunomodulatory properties as well as anti-thrombotic properties particularly HCQ have been utilized in the treatment of anti-phospholipid syndrome (APS), rheumatoid arthritis (RA), and systemic lupus erythematosus (SLE). It has been clearly shown that CQ and HCQ are good in the management of viral infections because they might potentially inhibit viral entry and spread from various *in vitro* and *in vivo* studies. The application of these drugs has been proposed on several human viruses, espe-

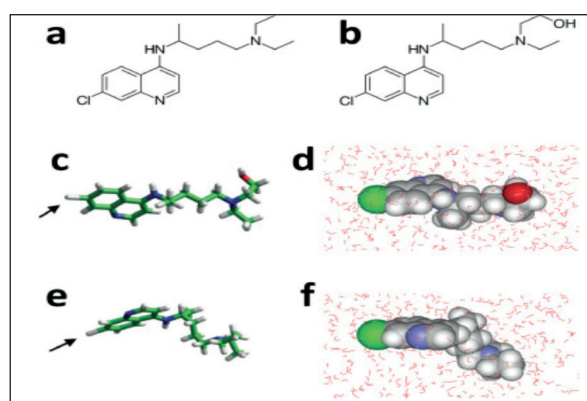


Figure 1. Chloroquine (CLQ) and hydroxychloroquine (CLQ-OH) chemical structures. (a) CLQ. (b) CLQ-OH. (c) CLQ-OH expanded conformer. (d) Aqueous CLQ-OH. (e) Typical dense conformer of CLQ. (f) Aqueous CLQ. The molecules in (c–f) appear in cylindrical or Spherical renderings (carbon, green; nitrogen, blue; oxygen, red; hydrogen, white). In (c) and (e), the chlorine atom of CLQ and CLQ-OH is shown using an arrow³.

cially Coronavirus like SARS-CoV and SARS-CoV-2. At the moment, fact-based findings on their application in human viral infections need elucidation¹.

Translation science from the bench to bedside should be established from comprehensive results, observational results as well as randomized experiments from scientific archives that have looked into the drug potency, duration of use and toxicity. These could be valuable in the prediction of their true efficacy. CQ sulfate and phosphate salts have both been endorsed as medications against malaria. HCQ has equally been utilized as a medication for malaria and auto-immune diseases, like lupus and rheumatoid arthritis. It is noted that CQ or HCQ are relatively safe with moderate and transitory side effects. In any case, there is a thin line between the treatment and toxic dose; the toxicity associated with CQ can lead to life-threatening cardiovascular disorders².

Effect of Chloroquine on Cellular and Biochemical Indices

Weak bases like CQ or HCQ influences acid vesicles by causing loss of enzymes function. Extracellularly, chloroquine or hydroxychloroquine have protonated structures, hence its positive charge and its inability to move across the plasma layer. Notwithstanding, the intracellular compartment permits the entry of the non-protonated part, hence the pH is inversely proportional to its protonation, just like Henderson-Hasselbach law. Therefore, chloroquine or hydroxychloroquine is

abundant in the acidic organelles, for example, CQ or HCQ atoms are positively charged mainly because the Golgi vesicles, lysosomes, and endosome have low pH⁴. CQ or HCQ is expelled to the extracellular space generally by exocytosis and multidrug resistance protein MRP-1 activity, through a cell surface drug carrier ATP-binding cassette family and P-glycoprotein⁵⁻⁷. It has been proven that weak bases restrain post-translational modifications of nascent proteins by interrupting many enzymes such as acid hydrolases through the elevation of lysosomal pH and trans-Golgi network (TGN) vesicles. The rise in endosomal pH regulates iron metabolism inside the cells, reduces ferrated transferrin endosomal iron release and intracellular iron concentration all through the action of CQ. This reduction, influences the capacity of some enzymes activating cellular DNA replication and the gene expression pathways^{8,9}.

Antiviral Activity of the Drug Chloroquine or Hydrochloroquine

Antiviral activity is one of the proposed mechanism for both CQ and/ or HCQ. The previous research explains this activity by pointing out their weak basic nature. HCQ is an analogue of CQ, in the golgi it brings about increased acidic pH in the intracellular organelles, autophagy in vesicles, endosomes, lysosomes, bringing about autophagic flux¹⁰ of the virus. CQ or HCQ could inhibit viral multiplication, replication and entry into various systems^{11,12-15} even in SARS-CoV-2 disease. As far back as 1960, CQ antiviral action has been proven¹⁶ to cause delay in the progression of a wide range of viruses in cell culture¹⁷. The evidence of its activity in mice has been found in many viral infections, including human COVID OC43¹⁸, enterovirus EV-A71¹⁹, Zika infection²⁰ and flu A H5N1²¹.

Endosome-Mediated Viral Entry Interaction

A number of virus use endocytosis as a mechanism to permeate their target cells (Figure 2). This progression marks viral infection at the lysosomal compartment showing low pH enzymes activity. There is disruption of the viral molecule, hence releasing the infected nucleic acid and the enzymes essential for viral replication. CQ blocks viral infections necessitating a pH-dependent phase for entry like in the case of Borna virus²², the minute virus of mice MVMp²³, and the avian

leucosis infection²⁴. Interestingly, a pathologist report confirmed that CQ limits the uncoating of the hepatitis A virus, thereby disrupting the whole replication cycle²⁵.

Replication of Enveloped Viruses Interaction

The enveloped glycoprotein post-translational modification occurs inside the TGN and endoplasmic vesicles and there is a cycling of low pH in glycosyl-transferases and protease. All these processes largely depend on pH, the Mayaro virus progression was evidently inhibited by CQ²⁶ thereby causing the accumulation of herpes simplex virus 1 particle in the TGN²⁷. It was also noticed that in Flaviridae family of protein (flavivirus pr M to M), replication and proteolytic action were inhibited by CQ activity²⁸. Therefore, viral pathogenicity is destabilized. Furthermore, chloroquine prompts the manufacture of non-infectious retrovirus particles, observed in HIV-1 and avian reticuloendotheliosis virus REV-An²⁹. Inhibition appears to be the mode of action used in the glycosylation of enveloped glycoprotein.

Inhibition of Viral Attachment and Entry in the Host Cell

Firstly, coronavirus establishes a viral particle connection with host cells membrane, through specific cell membrane receptors and the structur-

al protein of the virus. CQ or HCQ could repress the entry of the virus by inhibiting the sialic acid biosynthesis, basic players in the recognition of ligand in the virus-cell. CQ *in vitro* activity against coronaviruses is due to the restriction of cellular N-glycosylation receptor on the virus r, SARS-CoV-2 and SARS-CoV angiotensin-converting enzyme 2 (ACE2), and perhaps spike in viral protein (S). This leads to a decreased binding activity between cell viral S protein and ACE2

which causes the glycosylation of SARS-CoV S protein in CQ therapeutic doses³⁰. In SARS-CoV-2 S protein also undergo glycosylation just like SARS-CoV, however this is seen in novel prospective³¹. During the *in silico* examination³², it was pointed out that the S protein of SARS-CoV-2 utilizes ACE2 receptor and the sialic acids linked to gangliosides on the cellular surface, potentially enhancing the cellular attachment to the virus. The *in silico* modelling proposes that CQ or HCQ could have high affinity to attach to host sialic acids and gangliosides, potentially inhibiting the interaction between S protein and the plasma membrane in the host. Given these perceptions, CQ or HCQ action could be in two different ways: decreasing the nascent virion infectivity and/or weakening viral passage. CQ was shown to reduce the phosphatidylinositol expression restricting clathrin assembly protein (PICALM)³³, which is a part of the many proteins

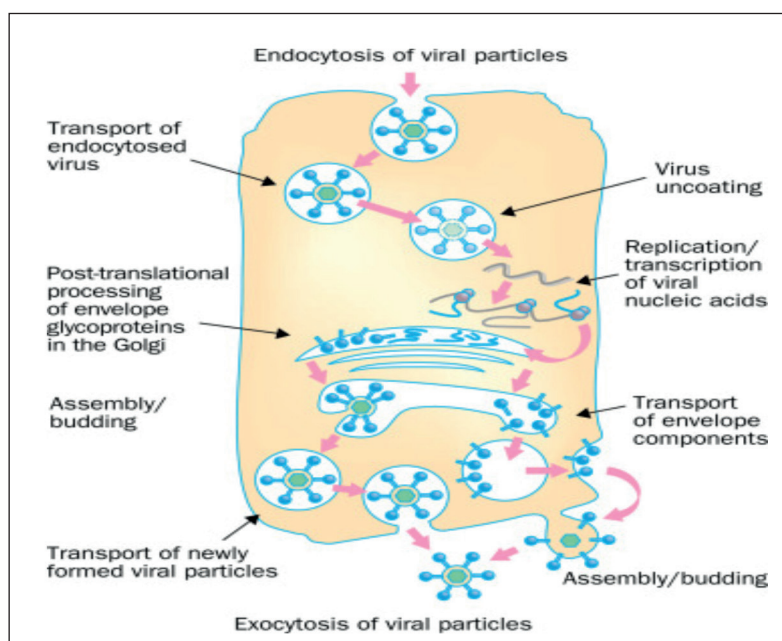


Figure 2. Steps of the replication of various infectious particles that can be targeted by chloroquine (marked by pink rectangles). Chloroquine delays the replication of various viruses either at the early or late phases of viral replication¹¹.

in clathrin-coated pits that manages clathrin-mediated endocytosis (CME) cell, i.e., an adaptor for cargo selecting, involved in entry of SARS-CoV into the mammalian cells³⁴. After the binding of the receptor, coronaviruses S-protein cleaves the endosomal proteases e.g., transmembrane serine protease 2 (TMPRSS2) and cathepsin at the acid dependent site. This allows the endosomal membrane and the viral cell to combine leading to elevated pH thereby causing inhibition. TMPRSS2 and cathepsin move by empowering S protein cleavage of SARS-CoV-2 enzymatic activities³⁵. At that point, CQ or HCQ inhibits the entry of virus and attachment in the cell of the host, perhaps causing obstruction of endocytic vesicles viral infection.

Inhibition of Development and Proliferation of Nascent New Viral Particle

CQ or HCQ have antiviral action, after a viral infection. The effect was found in SARS-CoV-2 and SARS-CoV infections *in vitro*^{13,14}. Another mechanism could therefore be associated with the activity of the antiviral agent. CQ or HCQ may repress or prevent the fusion of endosome-lysosome membrane prompting the recycling viral receptor membrane, the viral genome and viral uncoating is released into the cytosol after endosomal alkalination, as seen in SARS-CoV³⁶. The viral protein developmental processes may be disrupted by CQ/HCQ. This occurs inside the endoplasmic reticulum (ER)-Golgi intermediate compartment (ERGIC) and vesicles of trans-Golgi network (TGN) of the cell when there is low pH. Increase in pH could upset proteolytic handling and glycosylation of viral proteins. Similar to S protein, modifications and glycosylation occur in the coronaviruses envelop membrane protein likewise in ERGIC and TGN vesicles because the structural protein in the viruses is numerous¹². Other cleavage sites apart from TMPRSS2 and cathepsin were recognized in the SARS-CoV-2 S protein that could undergo cleavage by proteases similar to furin. In the TGN there is inhibition of Furin but profound expression is noticed in the lung, its S protein cleavage could be associated with viral progression and proliferation³⁷. This could be delayed by CQ, as seen in Chikungunya viral infection³⁸. Essentially, their basic properties interrupt compartment of the cell vesicles, CQ or HCQ may equally restrain viral growth. This takes place once the encapsulated virion genomes grow into the ERGIC layers because the virus envelope proteins are embedded, estab-

lishing the adult virus³⁹. In respiratory disorders, the virus binds with Toll-like receptors (TLRs), causing the deactivation of mitogen-activated protein kinase (MAPK) pathways, especially p38 MAPK. The respiratory system viruses inhibit the MAPK activation, causing the activation of their downline targets to transmit their proteins for virus progression and spread. The above mentioned effects are very significant in individuals having underlying airways diseases, similar to constant obstructive pulmonary disease (COPD) or asthma. These individuals have viral respiratory disease and CQ was found to interrupt the p38 MAPK activation pathway usually turned on and activated⁴⁰ subsequently inhibiting viral infection^{41,42}. COVID-19 individuals with COPD could benefit from the inhibitory activity because their SARS-CoV-2 infection could be life-threatening. In this way, other than viral connection, passage and uncoating, genome discharge, protein developmental cycle and assembly of nascent viruses development and progression might be restrained by the simple drug, resulting in a diminished infectivity.

Recent Findings on the Activating Mechanisms of CQ or HCQ in SARS-CoV-2 Infection

There is insufficient information about the mechanism of CQ or HCQ activation in the management of SARS-CoV-2 disease. Past publication on the ability of CQ OR HCQ in this infection was proved by *in vitro* studies. Wang et al¹³ revealed the ability of CQ to inhibit SARS-CoV-2 disease (myriad of infection, MOI = 0.05) in Vero E6 cells at decreased micromolar concentration (EC₅₀ = 1.13 μ M and EC₉₀ = 6.90 μ M, 2 days), medically accessible using 500 mg daily administration. Its effectiveness was assessed through the evaluation of virus copies in the cellular surface using Western blot examination, immunofluorescence microscopy, RT-PCR, accessing the nucleoprotein (NP). The scientists noticed that CQ disease inhibited the entry and post-entry points (drug administered 120 mins before the management of the virus, Western blot was done 14 h later). The investigators also analyzed CQ and HCQ efficacies inside a similar cell prototype and noticed that cell toxicity between the two prescriptions showed non significant differences. Estimating antagonistic effect to viral viability at various MOIs (0.80 0.2, 0.02, 0.01) using RT-PCR lower E50 values for CQ at all MOIs (CQ has 7.36, 7.14, 3.81, 2.71 for and HCQ 12.96, 17.31, 4.06, 4.51)

was noticed, affirmed via immunofluorescence examination for NP.¹⁴ These outcomes proposed that CQ is possibly more suitable than HCQ. The researchers also affirmed that HCQ and CQ at the entry and post-entry point brought about double activity in HCQ. another study projected that the cellular barrier of discovered virions and viral particles mostly in early and late endosome-lysosomes (EE, LELs) were under regulated conditions. At the point of cell treatment with CQ and HCQ, the EEs showed more particles, pointing the action of the drugs to block endocytotic vesicle intermediate stage development and possibly slowing down the EEs to LELs viral transportation, noticed a significant advancement in the viral genome movement. The researchers observed that the CQ and HCQ drug causes abnormal development of EEs; however, HCQ extended the number and size of LEL and the administration of CQ did not activate any modifications in LELs number and size. By the way, the structure of the vesicle was disturbed, proposing partially opposite modes of activity in both drugs. The action of CQ and HCQ against SARS-CoV-2 activity *in vitro* was affirmed looking at similar cell model⁴³. These writers discovered EC50 estimates of about 5.47 and 23.90 for CQ and 0.72 and 6.14 mM on days 2 and 1 CQ and HCQ independently. This is however different from the past report. HCQ had a better action. Both drugs have increased activity against viral infection during administration prior to virus infection, EC50 estimates of >100, 6.25 and 5.85 mM for HCQ and 18.01 for CQ individually on days 1 and 2.

The Antagonistic Action of CQ on SARS-CoV-2

Due to the said wide range of its anti-viral activity on most viruses, particularly in SARS-CoV- upon the entry of COVID-19 into the cell occurs via endolysosomal pathway⁴⁴, it seemed appropriate to examine the potential anti-SARS-CoV-2 activity of CQ. Both CQ and remdesivir (antiviral drug) have recently been reported to repress SARS-CoV-2 *in vitro* and in humans experiencing COVID-19¹³. The CQ and HCQ activity was assessed on some enveloped viruses and immune activation. Suggestions were made that the drug could be useful for SARS management in the treatment center. As of now, common anti-viral drugs particularly oseltamivir and ribavirin have not been shown to be effective in SARS treatment. Corticosteroids could be advantageous in controlling the lung inflammatory reactions⁴⁵

but unrestrained immunodepression could cause pulmonary distress. Lately, SARS causative agent has been described as novel corona virus⁴⁶. Recent examinations support the possibility that corona viridae attacks target cells using endocytic pathway and the replication can be restrained by chloroquine⁴⁷. Human Covid HCoV-229E infected cells given nocodazole (a microtubule depolymerising specialist which limits transport from early to late endosomes) secreted HCoV-229E antigens in small quantity⁴⁸. The outcome demonstrates that in HCoV-229E disease, there is a need for transportation in the endosomes. HCoV-229E antigens reduced in chloroquine treated cells⁴⁸. Recently, the China National Center for Biotechnology Development pointed out that CQ among three drugs has a favorable profile against the novel SARS-CoV-2 which causes COVID-19. In provinces around China especially in Beijing Clinic, Guangdong province CQ was employed. According to the initial report^{49,50}, CQ was found to cause reduction in fever including improved lung tomography in about 100 infected patients and required more chances to recover when compared with the control group. Moreover, no antagonistic effects were noticed and therefore the Chinese Medical Advisory Board has recommended that in the treatment of SARS-CoV-2, CQ could be employed. Thus, CQ is probably the primary drug utilized abroad and China is in the front line fight against SARS-CoV-2 disease. Despite the wide use of chloroquine in the treatment of malaria, it is important to take note of the safety dose, the side effect is macular retinopathy⁵¹, another side effect of CQ and HCQ could be cardiomyopathy^{2,53}. A SARS-CoV-2 infected patient profile must take note of the adverse effects of chloroquine treatment. Notwithstanding, chloroquine right now is one of the best drugs used in tackling the severity of SARS-CoV-2 diseases in people. HCQ, an analogue of chloroquine has little observable drug-drug interactions. In the past SARS outbreak, HCQ was thought to have adverse effect on SARS-CoV *in vitro*⁵⁴. It therefore means that HCQ might bring a ray of hope in COVID-19 disease treatment. Nonetheless, until this point, there is no hypothetical proof pointing to HCQ utilization as SARS-CoV-2 disease treatment. Other mechanism of action of HCQ and/or CQ is yet to be completely explained. Past researches discoveries have recommended that HCQ and CQ repress the COVID-19 progression. Medications can alter the pH of the cell layer surface subsequently hindering the viral infection

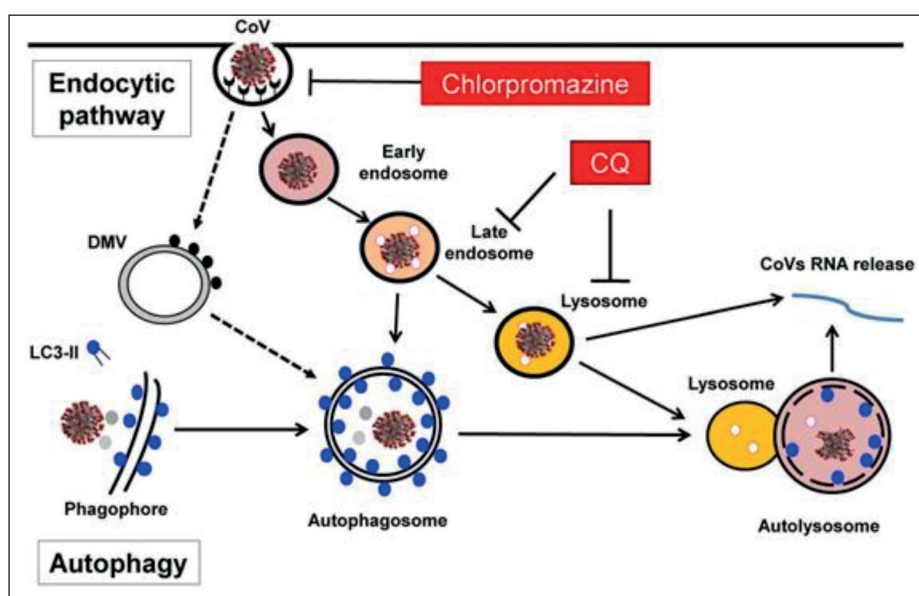


Figure 3. Diagrammatic representation of chloroquine effect on the replication cycle of severe acute respiratory coronavirus 2 (SARS-CoV-2).

from affecting the cell membrane. It also hinders nucleic acid duplication, viral proteins glycosylation, assembling of virus, nascent virus molecule transport, viral discharge, and different cycles to accomplish its antiviral effects⁵⁵. In certain patients reports, the immune response to the SARS-CoV-2 infection results in elevated cytokines interleukin (IL)- 6 and IL-10^{56,57}. This may increase cytokine storm, followed by multiorgan disruption and possibly death. Both CQ and HCQ have immunosuppressive effects and can reduce increased immune response^{11,58}. With this in mind, it is possible that early treatment with both medications may help inhibit the deadly progression of the disease progression. In chronic SARS-CoV-2 patients, corticosteroids administration might be injurious, if used they suppress the immune system and increase infection⁵⁹ (e.g., tocilizumab)⁶⁰.

Functional Principle of Chloroquine

CQ has various operating principles that may vary as shown by the researches conducted on microorganisms. CQ could hinder an initial viral cycle by meddling with viral particles binding to their cell surface receptor (Figure 3). CQ appeared to repress quinone reductase 2⁶¹, an underlying neighbor of UDP-N-acetylglucosamine 2-epimerases⁶² associated with the biosynthesis of sialic acids. The sialic acids are acidic monosaccharides found at the end of sugar chains present on cell transmembrane proteins and are the ba-

sic components of ligand recognition. The possible inference of CQ with sialic acid biosynthesis could be a pointer to the wide antiviral range of that drug since infections, for example, the human HCoV-O43 and the orthomyxoviruses use sialic acid moieties as receptors⁶³. The strength of anti-SARS-CoV-1 effects of CQ *in vitro* was ascribed to a deficiency in the glycosylation of a virus cell surface receptor, the angiotensin-converting enzyme 2 (ACE2) on Vero cells⁶⁴.

CQ could also impede the initial phase of virus replication by meddling with the pH-dependent endosome-mediated viral entry of enveloped viruses, e.g., Dengue or Chikungunya virus^{65,66}. Due to the alkalization of endosomes, CQ was a compelling *in vitro* treatment against Chikungunya infection after additional exposure to Vero cells prior to virus exposure⁶⁷. The mode of inhibition is probably the inhibition of endocytosis as well as fast rise of the endosomal pH and repeal of infection endosome fusion. A pH-dependent mode of entry of COVID-19 into target cells was also revealed for SARS-CoV-1 in the binding of the DC-SIGN receptor⁶⁸. The stimulatory step happens in endosomes at acidic pH resulting in fusion of the viral and endosomal layers prompting the delivery of the viral SARS-CoV-1 genome into the cytosol³⁶. During the lack of antiviral medication, the virus targets the lysosomal compartment with low pH, the enzyme's activity disturbs the viral molecule, subsequently releas-

ing the infectious nucleic acid, vital for its replication⁶⁹. CQ-mediated the inhibition of hepatitis A viral infection was discovered to be related to uncoating, impeding its whole cycle of replication²³. Post-translational modification of viral proteins is also disrupted by chloroquine. These post-translational modifications, which include proteases and glycosyltransferases, occur inside the endoplasmic reticulum or the trans-Golgi network vesicles and may require a low pH. For HIV, the antiretroviral impact of chloroquine is inferably from a post-transcriptional inhibition of glycosylation of the gp 120 envelope glycoprotein and the neosynthesised viral particles are not infectious^{70,71}.

CQ also hinders the replication Dengue-2 infection by influencing the normal proteolytic processing of the flavivirus prM protein to M protein²⁸. Therefore, viral infectivity is weakened. Examining the herpes simplex virus (HSV) model, CQ repressed growth of group of non-infectious HSV-1 particles in the trans-Golgi network²⁷. With non-human coronavirus, it was demonstrated that the intracellular site of COVID-19 spread is dictated by the limitation of its membrane M proteins that gather in the Golgi complex beyond the virion budding site⁷², proposing that the potential activity of CQ was observed at the cycle of replication in SARS-CoV-2. Recently, it has been announced that the MERS-CoV M Protein C-terminal space contains a trans-Golgi network limitation signal⁷³. The action of CQ was also noticed in the disruption of the viral developmental process, viral protein by pH regulation²⁸ and viral antigen recognition by dendritic cells and the necessary endosomal acidification through a Toll-like receptor-dependent pathway⁷⁴. Unexpectedly, other proposed CQ effects on the immune system comprise elevated soluble antigen export into dendritic cell cytosol and the improvement of cytotoxic CD8⁺ T-cell reactions against human antigens in virus⁷⁵. The influenza virus model showed that chloroquine improved non-replicating viral antigen in the cross-presentation of dendritic cells to CD8⁺ T-cells employed to lymph nodes depleting the site of infection, evoking a comprehensively defensive resistant reaction⁷⁶. The action of CQ on the immune system could also be via cell signaling and regulation of pro-inflammatory cytokines.

CQ represses phosphorylation of the p38 mitogen-initiated protein kinase (MAPK) in THP-1 cells just as caspase-1⁷⁷. Activation of cells by means of MAPK signaling is always necessary for infections to accomplish their replication

cycle⁷⁸. In the model of HCoV-229 COVID-19, CQ-induced infection inhibition happens through inhibition of p38 MAPK⁴¹. CQ is a notable immunomodulatory agent fortified as an intermediate for anti-inflammatory reaction¹¹. The use of this drug has also been observed in the treatment of rheumatoid arthritis, a well-known inflammatory disease, including others like⁷⁹⁻⁸¹ lupus erythematosus^{82,83} and sarcoidosis⁸⁴. CQ represses the mRNA expression of interleukin-1 beta (IL-1 β) in THP-1 cells and lessens IL-1 β delivery, macrophage/monocytes IL-1 and IL-6 cytokines tumor necrosis factor-alpha (TNF α) production. Another report⁹⁰ showed the inhibition of the TNF α receptor in U937 monocytic cells during CQ treatment. In the Dengue virus model using U937 cells infected, CQ inhibited TNF α , interferon-alpha (IFN α), IFN β , IFN γ , IL-6 and IL-12 gene expression⁹¹.

SARS-CoV-2, similar to some coronaviruses in humans, contains three envelope proteins, the spike (S) protein (180-220 kDa), the layer (M) protein (25-35 kDa) and the envelope (E) protein (10-12 kDa), necessary for infectious virions entry into target cells. The virus contains the nucleocapsid (N), an important part of replicase complex needed for binding to viral genomic RNA, and nsp3. Hemagglutinin-esterase (65 kDa) is a subset of β -corona viruses used in sialic acids binding at the glycoprotein surface. The host tropism determines S glycoprotein. SARS-CoV-2 also binds to angiotensin-converting enzymes protein 2 (ACE2) expressed on pneumocytes⁹². When ACE2 is bound, conformational changes are triggered on S glycoprotein orchestrating cleavage of the S protein by transmembrane protease TMPRSS2 and S protein release into the cell supernatant leading to antibodies neutralization⁹³. There is movement of cellular virus through the early and late endosomes, cleavage of protease cathepsin L by S protein at low pH, subsequently leading to viral envelope fusion with phospholipidic endosomal membrane bringing about viral genome fusion into the cell cytoplasm. Replication begins, followed by the translation of positive-strand viral genomic RNA to a negative RNA strand needed for virus mRNA synthesis template. There is a rise and then fall in the synthesis of negative MRNA strand relative to that of the positive strand. In infected cells, more positive than negative strands are observed (around 10 and 100 times more). The infected cell ribosome machinery is diverted particularly for the virus, which synthesizes its non-structural proteins (NSPs) which are

collected in the replicase-transcriptase complex to support viral subgenomic mRNA synthesis¹². After replication, there is translation of the envelope proteins, insertion into endoplasmic reticulum followed by transportation into the Golgi compartment. Before the formation of matured virions, there is viral genomic RNA arrangement into the nucleocapsid and fusion of envelope proteins at the budding stage. The M protein interacts with other viral protein causing the localization of trans-golgi network, before viral assembly. After assembly, the nascent viral particles are moved to vesicles cell surface and are delivered by exocytosis. Possibly, chloroquine meddles with ACE2 receptor glycosylation, subsequently stopping the binding between the target cell and SARS-CoV-2. Sialic acids biosynthesis is inhibited by chloroquine. This may be necessary for SARS-CoV-2 to bind to the surface of the cell. The successful binding of viral particles leads to regulation of endosome acidification by chloroquine causing restriction of both autophagosome development and viral replication caused by MAPK cell initiation reduction; chloroquine may likewise restrain viral replication. M protein development, virion assembly and budding can be disrupted by chloroquine. This was also part of the effects of chloroquine seen on the immune system¹¹.

ER-Golgi moderate compartment (ERGIC), the report exhibiting chloroquine exceptional capacity in lessening viral replication, with an Effective Concentration (EC) 90 of 6.90 μ M was attainable with standard dosing, because of its infiltration in tissues, as well as in the lung¹³. The researchers pointed out that chloroquine disrupts viral diseases by elevating endosomal pH and reducing glycosylation of cell receptor in SARS-CoV. The authors inferred that the antiviral effect *in vivo* could be a result of its established immunomodulatory effect¹³.

Studies

The Dutch Center of Disease control (CDC), in a public record on the internet, proposed that the treatment of extreme infections requires admission to the clinic with oxygen treatment or admittance to the ICU with CQ⁹⁴. Notwithstanding, the record also expressed that treating patients just with ideal steady consideration is still a sensible choice, due to the absence of strong evidence. The recommended treatment in adults includes CQ base (6 tablets A-CQ 100 mg; 600 mg) monitored by 300 mg following 12 h on day 1, at that point 300 mg \times 2/ on per of on 2-5 days. This record

also underlined the requirements for halting the treatment at day 5 to diminish the risk of side effects, bearing in mind the drug long half-life (30 h) and the need to separate regimens dependent on CQ phosphate and CQ base since 500 mg of the first set to 300 mg of the second⁹⁴. Another rule recorded by the Italian Society of Infectious and Tropical illness (Lombardy area) suggests the utilization of CQ 500 mg \times 2/ or HCQ 200 mg die for 10 days, despite the fact that the treatment may differ from 5 to 20 days as per clinical gravity. The recommended target populace went from patients with mild respiratory indications and comorbidities to patients with serious respiratory distress⁹⁵.

Chloroquine and Hydroxychloroquine as Antivirals in COVID-19: A Role for Hemin/Hemoglobin

The CQ and HCQ work differently against virus and Plasmodium. The main differential factor is the hemin/hemoglobin. Against virus like SARS-CoV-2, they work by diminishing the efficiency of the virus at entering host cells and decreasing its replication rate. In malaria, they work by poisoning the parasite digestive system (vacuole) via increasing free hemin that is toxic through oxidative stress and enzyme inhibition (cysteine proteases and others). The targeted cells are also different: ACE2-expressing cells such as epithelial ATII cells and other extra pulmonary cells in SARS-CoV-2 infection and red blood cells in malaria. In both circumstances, cell systems where CQ and HCQ accumulate are involved: lysosomes and related acidic compartments of host cells in SARS -CoV-2, and digestive vacuole in malaria parasite. It is thought that CQ and HCQ exert their anti-viral actions mainly via endosomes. However, the main entry point of SARS-CoV-2 is not by the endosome but instead by spike S protein. This may suggest that additional mechanisms could be involved in their antiviral action. Most drug candidates investigated in COVID-19 target key proteins of the host cell or the virus. The host's cell proteins are TMPRSS2 protease or cathepsins (used for virus entrance) and enzymes ACE2 (virus receptor). The targeted viral proteins are the S protein (a glycoprotein), the RNA-dependent RNA polymerase (RdRp), and the two proteases that are essential for viral replication: main protease (3CLpro), a chymotrypsin-like cysteine protease, and papain-like cysteine protein (PLpro). The 3CLpro utilizes a Cys/His catalytic dyad whereas PLpro has a classical Asp/His/Cys catalytic triad⁹⁶. PLpro possesses a C-terminal

zinc-finger like domain bearing four cysteine residues that are critical for catalytic function⁹⁶. Drugs are being tested as inhibitors of these cysteine proteases in order to block viral replication. CQ is not a good inhibitor of cysteine proteases. However, it interacts with heme and increases oxidative reactions (oxidative stress), as well as inhibition of cysteine proteases⁹⁷. These events might hypothetically occur in the host and virus during infection. Cell cultures might contain (methemoglobin) and heme eventually influencing the antiviral action of CQ and HCQ residual hemoglobin. *In vivo*, hemoglobin is confined into red blood cells, which in contrast to *Plasmodium* are not presumably targets of SARS-CoV-2. Recent results suggest that the virus might use CD147 receptor as an additional gate into host cells and the same receptor is used by *Plasmodium* to enter into red blood cells^{98,99}. However, noticeable hemolysis is not stated as a common feature of patients with COVID-19 although ferritin appears to increase. Nevertheless, there are possible sources of extravascular local cell-free hemoglobin and heme that should be considered. Hemoglobin could be locally released from red blood cells breakdown in capillaries close to alveolar cells damaged during infection. Thus, SARS-CoV infections caused hemorrhages in the alveolar space of the lungs¹⁰⁰, and lung hemorrhages in animal models infected with coronaviruses are significantly reduced with antivirals¹⁰¹. SARS-CoV-2 attacks blood vessels and some patients show coagulation dysfunction and develop blood clots while others develop pulmonary embolism. The pathological analysis of the lungs of COVID-19 patients shows noticeable microvascular thrombosis and hemorrhage linked to interstitial inflammation and infected alveolar¹⁰².

Clinical pathological investigations of lung biopsies and dissections of COVID-19 patients demonstrate alveolar discharge along small vessel clots arrangement around the lung periphery^{103,104}. Alveolar Hemorrhage is a typical component of the acute respiratory distress syndrome (ARDS), a condition related with COVID-19. ARDS patients have raised degrees of cell-free hemoglobin inside the alveolar space from red platelet breakdown¹⁰⁵, and it contributes to the intensity of lung pathogenesis. Cell-free hemoglobin caused elevated inflammation, injury to the cell epithelial and loss of integrity of alveolar capillary barrier¹⁰⁵. It converts methemoglobin and delivers free Heme¹⁰⁶ cellularly.

Heme increased alveolar-capillary permeability (barrier dysfunction) but did not cause alveolar inflammation or epithelial cell injury in the

lungs¹⁰⁵. Finally, the drugs CQ and HCQ might also induce erythrocyte break down in some patients¹⁰⁷. Extracellular heme/hemoglobin released from red blood cell breakdown might be a target for the action of CQ and HCQ. On the other hand, hemoglobin is expressed in blood cells other than erythrocyte such as epithelial cells and activated macrophages. Expression of this protein has been observed in several human tissues including the lungs, particularly in alveolar epithelial cells type II (ATII) that produce the pulmonary surfactant essential for lung function¹⁰⁸⁻¹¹⁰. Hypoxia greatly increased hemoglobin expression in ATII cells, and decreased surfactant proteins¹¹⁰. Hemoglobin in these cells could increase the concentrations of oxygen, and oxygen transport facilitation across air-blood barrier¹¹⁰, and it might have a role in ARDS. Remarkably, epithelial ATII cells that are main targets of SARS-CoV-2 infection also express hemoglobin, and this expression highly increases during hypoxia, a phenomenon occurring during viral infection. Hemoglobin/heme expressed in ATII cells might be a target for the action of CQ and HCQ. Local extracellular hemoglobin or hemoglobin expressed in ATII cells could be oxidized to methemoglobin (Fe^{3+}). This oxidation produces reactive oxygen species (ROS) (H_2O_2), in the presence of inflammatory-cell-derived oxidants¹¹¹. Methemoglobin is toxic, generates ROS, and releases free heme¹¹², which can be incorporated into endothelial cells^{111,113} and perhaps other cells as well¹¹⁴. Endothelium protects itself from heme/heme by heme induction, heme oxygenase degradation and production of large amounts of ferritin, iron binding protein¹¹³. CQ and HCQ could modulate those actions. For example, their interaction with free heme could trigger cysteine proteases inhibition and oxidative reactions of the virus and the host, which could result in antiviral actions.

CQ could also limit degradation of extracellular hemoglobin by alveolar macrophages. During disposition of extracellular hemoglobin in hemoglobin scavenger receptor CD163-positive HEK cells, CQ action resulted in intracellular hemoglobin trapping compromising clearance and abolished expression of heme oxygenase-1 (HO-1). CQ blocks the elimination of hemoglobin (or methemoglobin) by interfering with lysosomal hemoglobin degradation, and caused intracellular accumulation of heme and globin¹¹⁴. The results support previous studies stating that chloroquine acts by increasing free heme (and hemoglobin) to potentiate oxidative reactions (oxidative stress)

and inhibition of cysteine proteases⁹⁷ that might affect the host body and virus. However, for effectiveness, this mechanism would need the presence of sufficient local free heme/hemoglobin to interact with chloroquine. It is currently unknown whether this really occurs during infection in COVID-19. In a recent preprint¹¹⁵, it is reported that non-structural proteins of SARS-CoV-2 could attack hemoglobin and heme to release porphyrins used for the virus. This could be interpreted as the virus degraded hemoglobin. However, this is a bioinformatics study and not an experimental work, and does not prove the breakdown of hemoglobin or heme to porphyrins, that would need a kind of heme oxygenase. As described here, CQ and HCQ exert diverse biological effects as shown also by their anti-inflammatory actions in autoimmune (lupus), rheumatoid arthritis diseases and in porphyria¹⁶¹¹⁷. As discussed here, an interaction of these drugs with heme is expected. In this regard, intravenous heme (hydroxyheme or heme arginate) is used for the treatment of acute porphyria attacks. It helps to overcome the relative deficiency of heme in the liver and reduces the supply of porphyrins and precursors needed for heme production. During the current COVID-19 pandemic, porphyria patients could be infected with SARS-CoV-2 and still be treated with heme for-patients/COVID-19-and-porphyria). Harmful drugs are avoided during treatment with heme although CQ and HCQ might probably be safe based on this evidence though they are still insufficient. So far, it is unknown whether possible simultaneous cases of acute porphyria attack and COVID-19 have been treated with heme, CQ and HCQ, but it would be of interest to know the outcome because an interaction of heme with these drugs is expected. Remarkably, heme exhibits antiviral actions against several viruses such as HIV, Zika, influenza, or Ebola. This action could be related with the induction of heme oxygenase enzyme¹¹⁸. Therefore, the antiviral action of heme supports a possible mechanism for antiviral action of CQ and HCQ with involvement of heme, as discussed above. Heme inhibited HIV replication *in vitro* and reduced viral load in mice¹¹⁹. It significantly inhibited replication of Zika virus *in vitro*¹²⁰. Heme showed anti-influenza virus activity *in vitro* and *in vivo*¹²¹. It ameliorated influenza pneumonia and attenuated virus-induced lung injury, lymphocytopenia and local inflammation in a mouse model. Heme protected mice from death and body weight loss caused by influenza A virus infection¹²¹. Heme had an important effect

in the reduction of Ebola virus replication. Treatment of different human cells with heme reduced infection by >90% and showed minimal toxicity to infected cells¹²⁰. This inhibition could be partly due to heme oxygenase (HO-1) activity and expression. The activity of reverse transcriptase in murine leukemia virus was repressed by heme and inhibition increased 1000-times when heme solution was aged¹²². Heme also potentiated the activity of artemisinin against Hepatitis C virus showing synergistic effects¹²³. Taken together, the antiviral effects of heme are noticeable and could be beneficial in the present situation of absence of effective treatments in COVID-19. In this regard, the action of heme, and the combined action of heme and CQ and HCQ may be worth testing in SARS-CoV-2 (COVID-19) studies evaluating antiviral action and effects on the disease. Nevertheless, possible safety and toxic effects should also be considered as heme may induce toxic actions¹²⁴ and CQ and HCQ are not exempted from the undesirable effects such as heart problems and retinopathy^{107,125}.

CQ/HCQ and Iron Metabolism: SARS-CoV-2 Infection and the Role of Iron

The fundamental component for every living being is iron which is also due to its activity in redox reaction I as a basic cofactor in many enzymatic and proteins associated with essential cell function like ATP production, replication and transcription of DNA. Indeed, most viral infections also need iron, as they depend on metabolic mechanism of the host for genome replication and production of mRNAs for translation into useful viral proteins¹²⁶. Subsequently, the cellular repletion of iron increased viral replication and decreased (this term has been moved, please check if it is ok) multiplication or progression can disrupt the life cycle of the virus.

In the course of infections and inflammation, pro-inflammatory cytokines cause iron deficiency. A good number of these directly influence the stability of iron similar to Interleukin-1 β , Interleukin-6 and Tumor Necrotic Factor- α . These cytokines, especially interleukin-6, bring about iron-regulatory hormone hepcidin (HAMP) up-regulation, basically delivered by hepatocytes into the circulatory system direct the activities of iron homeostasis systematically. Iron transportation is blocked in the cell by systemic HAMP blocks through ferroportin 1 (FPN1), bringing about decreased iron absorption in the intestine, elevated macrophages and hepatocytes iron retention and

at last inflammatory/infection anemia¹²⁷. Some cells apart from the hepatic cells have yielded and delivered HAMP that circulate as autocrine and paracrine particles, maintaining a constant internal environment for iron¹²⁷. Not just cells of the immune system like monocytes, lymphocytes, and macrophages (together with alveolar macrophages) but also the airways epithelial cells have been shown to yield HAMP in the course of inflammation, infections and possibly aggravated lung injury¹²⁸. The peptide HAMP is associated with the critical phase protein and natural antimicrobial resistance¹²⁹, lactoferrin (LF), transferrin (Tf), ferritin (FT), hemopexin (HPX) and haptoglobin (HP) are proteins strongly associated with iron, viral diseases. Investigations have been made on viral diseases and iron breakdown in humans¹³⁰.

Incitation of Iron Starvation by CQ OR HCQ in the Cell: A SARS-CoV-2 Duplication Mechanism of Action

Several models of investigation showed that iron entry was restricted by CQ like in eukaryotic model *Saccharomyces cerevisiae*, CQ distorted iron uptake causing competitive inhibition of iron entry and starvation. Yeast that lacked iron showed elevated sensitivity to CQ by the methods of knocking out genes associated with uptake of iron or by utilizing iron chelators¹³¹. In the cells of mammals, CQ acts similarly by inhibiting endocytotic complex of Tf/transferrin receptor 1 (TFR1). The fibroblast obtained from cultured rats gave key evidence that Tf uptake was inhibited by CQ¹³². The major plasma iron transporter is Tf that maintains the cofactor in a passive redox environment by circulating it across human body cells. Two Fe^{3+} firmly binds Tf. TFR1, found on most cell types in the plasma membrane, binds and internalizes Tf into endocytic vesicles through CME¹³³. Similar to the previous ones, CQ has been shown in treated mice macrophages to lessen the expression of PICALM¹³⁴. This protein is expressed ubiquitously with CME and its inadequacy was shown to bring about abnormal metabolism of iron and anaemia in mice¹³⁵ for embryonic fibroblast in murine iron starvation, elevated expression of surface TFR1 with low iron level intracellularly¹³⁶. At that point, CQ OR HCQ administration may bring about cellular iron starvation and inhibition of the uptake of Tf/TFR1 complex. Iron release from Tf after iron-loaded Tf/TFR1 complex CME and its cytosolic movement is a basic stage for cell procurement and

further utilization of iron. The endosomal acidic milieu delays Tf binding to Fe^{3+} , leading to the vesicular delivery¹³³ of CQ or HCQ, elevating endocytic vesicles pH, potentially repressing endocytic vesicle Tf iron expulsion. Inside the vesicle of the endocytes, metalloredutase reduces transported Fe^{3+} to Fe^{2+} a six-transmembrane epithelial antigen of the prostate 3 (STEAP3) moved into the cytosol from the lysosomes by mucopolin 1 (TRPML1/MCOLN1), different carriers and divalent metal-ion transporter 1 (DMT1)¹³³. The transportation of iron from DMT1 is pH dependent, particularly active when the pH is low. Before iron can be moved into the cytoplasm by DMT1 endosomes, an $\text{H}^+/\text{Fe}^{2+}$ symporter needs a positively charged electrochemical potential gradient¹³³.

Na^+ , K^+ and Ca^{2+} are ions that can penetrate TRPML1/MCOLN1 channel also transporting Fe^{2+} . Before the delivery of cation to the cytosol from the lumen, there must be late endosome, acidic vesicles, lysosome and cation delivery¹³⁰. At that point iron delivery from endosomes into the cytosol is inhibited by CQ or HCQ.

CQ or HCQ is used to weaken autophagic movement and the fusion of endosome or lysosome¹³⁷. The basic cellular iron storage is FT which compartmentalizes the non-reactive form of iron until it is used. The delivery of iron from FT mostly happens by protein break-down through a specific lysosome-autophagy pathway called ferritinophagy¹³⁰ in which CQ is inhibited¹³⁸.

Additionally, both TRPML1 and DMT1 have been involved in the iron arrival from ferritinophagy, this occurs in acidic environment because these channels open to allow trapping of autophagic vesicles. The above-mentioned steps cause cellular iron deprivation. The aforementioned condition may perhaps affect the life cycle of SARS-CoV-2, though presently the experiments have not been proven because this pandemic is a strange disease, hence the need for more study. ACE2 receptor is the by which SARS-CoV-2 enters the cell in humans, especially in the lung alveolar epithelial cells, vascular endothelia, skin, kidney, heart, muscle and bronchial and gastrointestinal system¹³⁹. Generally, in human cells iron is an important cofactor for SARS-CoV-2 target cells and many other important life processes involving proteins such as cell development, bioenergetics, nucleic acid synthesis including the virus life cycle. Some viral diseases are found to elevate the uptake of iron in cells, the inhibition of iron starvation during cell cycle of viruses has been proven in various human infections like Hu-

man Deficiency Virus 1 (HIV-1), Human Cytomegalovirus (HCMV) and hepatitis C infection (HCV)¹²⁶. Elevated uptake of iron in cell including FT synthesis has been exhibited in infected mice liver with mouse hepatitis virus type 3 (MHV-3), from the family called Coronaviridae¹⁴⁰.

CQ or HCQ Could Prompt Iron Starvation in the Cell: A Possible Valuable Equilibrium Between Innate And Adaptive Immune Response

Another significant point noticed about CQ or HCQ is their action on cellular immunity through iron starvation, considering the natural and adaptive immune responses against viral infections. For activation, proliferation and proper immune functions, iron is an essential requirement. Excess iron frequently brings about delayed immune response to infections, as seen in hemochromatosis (HH) or thalassemia patients¹⁴¹. Therefore, immune responses that are extreme or dysregulated have specific significance in COVID-19 pathogenesis¹⁴² in the same way as other coronaviruses^{143,144}. Many coronaviruses have shown direct infection to both their inborn and adaptive cellular immunity^{143,149}. At that point iron starvation could equally repress the infections mentioned earlier. Native macrophages can differentiate under the cytokines stimulation in customarily initiated pro-inflammatory macrophages (M1), stimulated by IFN- γ , TNF- α or IL-4 and IL-13, otherwise activated macrophages (M2), associated with microbe clearance, repair of tissue and decreased inflammation. However, M2 macrophages possess low levels of iron; M1 macrophages retain iron, secrete significant levels of pro-inflammatory cytokines, free radicals, HAMP to kill microbes and limit the exit of iron.

Elevated deposition of iron inside the macrophages causes polarization of M1, the pro-inflammatory state elongation as a result early switching to the M2 state¹⁴⁶. The life cycle of viruses can thrive intracellularly by retaining iron in the macrophages during infection advancing the inflammatory process to that eventually bring about antagonistic effects. Furthermore, secondary diseases with different microbes could be caused by excess iron in macrophages. Prolonged administration of CQ brings about reduced iron content in rat models^{9,147} indicating CQ capacity to lessen iron content in liver and spleen and alveolar macrophages of rats groups

in the control, iron-loaded and iron-deprived rats. Also, rats group given lipopolysaccharide (LPS) showed decreased oxidative reaction, estimated by discharge of NO₂, after CQ treatment suggesting that CQ may avert infections, especially those related or caused as a result of iron overload, by restricting iron accessibility in the cells affected by infection similarly in macrophages, thus decreasing inflammation. In any case, homozygous and heterozygous mutations in hemochromatosis gene (HFE) decrease the effect of CQ on iron expulsion in porphyria cutanea tarda¹⁴⁸. Another model is murine, CQ appeared to lessen macrophage activation syndrome especially in hemophagocytic condition by the initiation of pristane, phagocytic functions, by decreasing the penetration of macrophage, production of cytokines and causing a decrease to FT lactate dehydrogenase and triglyceride levels¹⁴⁹.

For activation and proliferation, immune cells need iron but excessive iron is proven to impede the antigen production. The introduction by APCs alters CD4⁺ cells and modifies CD4⁺/CD8⁺ lymphocyte proportion, increases circulating B cell produced by immunoglobulins, decreases natural killer (NK) cell has the ability to lyse cells and disturbs supplement initiation¹⁵⁰; however, iron deficiency produces immunosuppressive effects to T cells¹⁵¹. At that point, CQ or HCQ administration, due to their activity on iron homeostasis, might have a wide function in SARS CoV-2 adaptive response regulation.

CQ OR HCQ Reduces IL-1 β , IL-6 and TNF- α Release: Could Possibly Reduce the Release of Local and Systemic HAMP

CQ and HCQ inhibit cytokine, through their action on iron homeostasis, inflammation reduction and reestablishment of initial iron homeostasis, thus protecting erythropoiesis. Furthermore, IL-1 β , IL-6, TNF- α induce HAMP release both locally and systemically. Iron is fundamental to life, however excess iron is harmful, due to the redox potential that hinders oxidative stress that is detrimental to essential cell constituents. Iron accessibility is then firmly controlled equally systemically and cellularly^{126,130,153,154}. On the other hand, deficiency of iron hinders its appearance, using the hepatic cells and macrophages in the intestine to cause the absorption, maintenance and discharge of iron. Erythropoietin (EPO) as well as hypoxia downregulate HAMP expression through

erythroferrone (ERFE) that permits the recruitment of iron for the formation of red blood cells, but during inflammation all these are upregulated. IL-6 initiates HAMP during inflammation, via the JAK/STAT3 pathway and BMP/SMAD pathway, yet in addition TNF- α and IL-1 β have a direct function on the regulation of HAMP¹⁵⁵⁻¹⁵⁷. As expressed above, CQ or HCQ does not just interfere with iron initiation in the cell causing its starvation in alveolar macrophages^{9,147} bringing about a change to M2 mitigating state, it also restrains, TNF- α , IL-1 β and IL-6 release, perhaps decreasing native HAMP liberated through the macrophages. This decrease can bring about more reduced retention of iron in these cells permitting inflammation resolution. Additionally, cytokines reduction might bring about the decrease of HAMP in the system, through elevated iron absorption in the intestine, enhancing anemic infection. Interestingly, COVID-19 signs have been abated by EPO treatment¹⁵⁸; however, by *in silico* modelling, SARS-CoV-2 now plausibly binds heme causing the dissociation of porphyrin and iron¹⁵⁹.

The Administration of CQ or HCQ for Treatment, Reduces Thrombosis, Anemic Infection by Reducing HAMP and Releasing

Further investigations linked thrombocytic events with thrombocytosis induced by iron deficiency anemia (IDA)¹⁶⁰⁻¹⁶². A new report proved that IDA patients showing thrombocytosis had twice as much tendency of developing thrombosis than IDA patients with normal platelet count¹⁶³. Remarkably, patients diagnosed with COVID-19 with serious pneumonia seem to have elevated platelet count compared to patient with severe pneumonia without COVID-19¹⁶⁴. This kind of elevation is predominant among the non-survivor compared with COVID-19 patients that are survivors¹⁶⁵. Animal models like Sprague-Dawley are used to explain the influence of iron deficiency on thrombotic susceptibility induced by thrombosis. They were fed with an iron-free diet, while the ordinary diet regimen served as control.

Thrombocytosis caused by iron deficiency and platelet count resulted in a significant blood clot. Furthermore, platelet aggregation and adhesion were weakened. Considering the information obtained from this model, the authors presumed that anemic inflammation, brought about by HAMP-mediated iron sequestration in the liver, macrophages and spleen, might be viewed as a functional iron deficiency (ID) and

affected patients should be treated as thrombosis high risk patients especially COVID-19 patients¹⁶⁶. Taken together, the diverse activities of CQ and HCQ as an anti-inflammatory, anti-thrombotic and antiviral medication might also be connected to their effects on iron homeostasis, both systemically and locally. Noticeably, a different regular treatment, routinely utilized in the management of COVID-19 patients with predominantly increased levels of heparin. Similar to CQ and HCQ, heparin is a universal drug, as characterized by Jecko Thachil¹⁶⁷, with potential activity such as anti-inflammatory, anti-viral and anti-coagulant medication. It is noteworthy that CQ or HCQ as well as heparin turned out to control iron metabolism: HAMP expression in human macrophages, FPN1 expression in the plasma-membrane and advance iron export, leading to cellular starvation of iron, antithrombotic activities are part of all the remarkable properties of this drug¹⁶⁸. These findings recommend a possible function of iron in SARS-CoV-2 infection that needs to be investigated in future for clinical studies, it should be equally considered as a possible target for COVID-19 treatment as proposed for other infectious diseases in humans in the past^{126,153,154}.

Safety Considerations

CQ and HCQ have a well-considered profile. This medication has been utilized for over 50 years' in the treatment of malaria showing the safety of the administration of chloroquine to individuals. The utilization of CQ and HCQ in the treatment of rheumatic diseases and for anti-malarial prevention indicated that there is a low rate of unfavorable occurrences during constant administration of this medication for a couple of years. In these circumstances, macular retinopathy is the most serious side effect, which occurs as a result of the cumulative dose instead of daily dose, while permanent injury might be prevented with normal visual examination during management^{51,169,170}. A new report¹⁷¹ gave an encouraging conclusion on what to expect during high dosage authorizing results on the safety of a high dosage of the medication (about 500 mg of chloroquine base every day) even at gestation period. We presume that CQ and HCQ administration offers controlled and limited toxicity and may consequently bring about a generally safe/advantage balance when it is utilized in life frightening conditions. Consequently, we resolve to examine the suitability of this old medication in

the management of dual infectious diseases presenting a danger to global health in the period of globalization, AIDS, as well as severe acute respiratory condition (SARS). These illnesses are both brought about by enveloped RNA viruses, and offer some clinical signs that are probably going to be interfered with by immune responses of the host¹¹. CQ and HCQ are startlingly poisonous when overdose is administered, showing a few signs similar to cyclic anti-depressing toxicity. Persistent or spontaneous overdose prompts quick initiation of central nervous system toxicity (seizures and unconsciousness), cardiac breakdown (with inhibition of the heart sodium and potassium channels bringing about QRS elevations and QT interval extension, respectively) and hypokalemia resulting from intracellular fluctuations¹⁷². Management of overdose strongly requires incorporation of brief therapy of activated charcoal, intravenous benzodiazepines and vasopressors fluctuating, sodium bicarbonate or hypertonic saline for substantial QRS amplification and interrelated arrhythmias, as well as cautious administration of hypokalemia to avoid overcorrection. Strict monitoring from a toxin control center is recommended in every cases.

Could Consumption of These Medicines Possibly Exacerbate COVID-19 Infection?

In spite of the positivity associated with ability of chloroquine and/or hydroxychloroquine in the management of COVID-19, few considerations have been given to the likelihood that the medications may adversely affect the development of infection. For instance, a few proposed that inhibition of T-partner cell expansion and interleukin-2 formation or reaction may coincidentally increase the inflammatory reaction in infected patients, perversely affecting the health of the patients¹⁷³. The likelihood that these medications may present unfavorably effect points out the need for high-quality randomized controlled clinical trials even with an emerging pandemic¹⁰⁷. The immunomodulatory effects of CQ and HCQ are basic to a subset of patients with rheumatoid joint pain, ankylosing spondylitis, systemic lupus erythematosus, sarcoidosis and other chronic infections. Increased treatments based on theory of their functions interactions or therapy in SARS-CoV-2 infection by patients having chronic disorders should be critically examined. In this light, two drug manufacturing companies have declared their plans to increase hydroxychloroquine production fully anticipating this necessity¹⁰⁷.

Conclusions

COVID-19, an infectious disease caused by the novel coronavirus advances through different means to induce infection and its spread¹⁷⁴. The outbreak of COVID-19 has imposed threat to global health and economy characterized by its increased incidence and mortality globally¹⁷⁵. The main proponents of drug repurposing in the proposed management of SARS-CoV-2 infection are CQ and HCQ. Hence, the functional role of CQ and HCQ was analyzed providing possible mechanistic insight into the role of iron on viral infection, iron starvation and its downstream cellular pathways involving hepcidin and proinflammatory cytokines. Overall, the possible mode of action of CQ and HCQ in the management of COVID-19 infection is exhibited via its anti-viral, anti-inflammatory and anti-thrombotic activities which could be considered in viral diseases linked to inflammation or potentially immune initiation. Cytokine inhibition by CQ or HCQ, through their action on iron homeostasis, could reduce inflammation, reestablishing a reasonable basis for iron homeostasis and erythropoiesis. CQ or HCQ have their action on cellular immunity through iron starvation, considering the natural and adaptive immune responses against viral infections.

Conflicts of Interest

The authors declare no conflicts of interest.

References

1. Touret F, de Lamballerie X. Chloroquine and COVID-19. *Antiviral Res* 2020; 177: 104762.
2. Musshoff F, Madea B. Demonstration of a chloroquine fatality after 10-month earth-grave. *Forensic Sci Int* 2002; 125: 201-204.
3. Fantini J, Di Scala C, Chahinian H, Yahi N. Structural and molecular modelling studies reveal a new mechanism of action of chloroquine and hydroxychloroquine against SARS-CoV-2 infection. *Int J Antimicrob Agents* 2020; 55: 105960.
4. Ohkuma S, Poole B. Cytoplasmic vacuolation of mouse peritoneal macrophages and the uptake into lysosomes of weakly basic substances. *J Cell Biol* 1981; 90: 656-664.
5. Pescarmona G, Morra E, Aldieri ED, Bosia A. Movement of vesicles in eucariotic cells: role of intravesicle protons as a fuel and modulation of their concentration by drugs or metabolic changes. *MRS Onli Proc Libr Arch* 1997: 489.
6. Vezmar M, Georges E. Reversal of MRP-mediated doxorubicin resistance with quinoline-based drugs. *Biochem pharmacol* 2000; 59: 1245-1252.

7. Vezmar M, Georges E. Direct binding of chloroquine to the multidrug resistance protein (MRP): possible role for MRP in chloroquine drug transport and resistance in tumor cells. *Biochem Pharmacol* 1998; 56: 733-742.
8. Byrd TF, Horwitz MA. Chloroquine inhibits the intracellular multiplication of *Legionella pneumophila* by limiting the availability of iron: A potential new mechanism for the therapeutic effect of chloroquine against intracellular pathogens. *J Clin Invest* 1991; 88: 351-357.
9. Legssyer R, Josse C, Piette J, Ward RJ, Crichton RR. Changes in function of iron-loaded alveolar macrophages after in vivo administration of desferrioxamine and/or chloroquine. *J Inorg Biochem* 2003; 94: 36-42.
10. Klionsky DJ, Abdelmohsen K, Abe A, Abedin MJ, Abeliovich H, Acevedo AA, Adachi H, Adams CM, Adams PD, Adeli K. Guidelines for the use and interpretation of assays for monitoring autophagy. *Autophagy* 2016; 12: 1-222.
11. Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old drug against today's diseases. *Lancet Infect Dis* 2003; 3: 722-727.
12. Fehr AR, Perlman S. Coronaviruses: an overview of their replication and pathogenesis. *Methods Mol Biol* 2015; 1282: 1-23.
13. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, Shi Z, Hu Z, Zhong W, Xiao G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res* 2020; 30: 269-271.
14. Liu J, Cao R, Xu M, Wang X, Zhang H, Hu H, Li Y, Hu Z, Zhong W, Wang M. Hydroxychloroquine: a less toxic derivative of chloroquine is effective in inhibiting SARS-CoV-2 infection in vitro. *Cell Discov* 2020; 6: 16.
15. Devaux CA, Rolain JM, Colson P, Raoult D. New insights on the antiviral effects of chloroquine against coronavirus: what to expect for COVID-19? *Int J Antimicrob Agent* 2020; 55: 105938.
16. Inglot AD. Comparison of the antiviral activity in vitro of some non-steroidal anti-inflammatory drugs. *J Gen Virol* 1969; 4: 203-214.
17. Keyaerts E, Vijgen L, Maes P, Neyts J, Van RM. In vitro inhibition of severe acute respiratory syndrome coronavirus by chloroquine. *Biochem Biophys Res Commun* 2004; 323: 264-268.
18. Keyaerts E, Li S, Vijgen L, Rysman E, Verbeeck J, Van RM, Maes P. Antiviral activity of chloroquine against human coronavirus OC43 infection in newborn mice. *Antimicrob Agents Chemother* 2009; 53: 3416-3421.
19. Tan YW, Yam WK, Sun J, Chu JH. An evaluation of chloroquine as a broad-acting antiviral against hand, foot and mouth disease. *Antiviral Res* 2018; 149: 143-149.
20. Li C, Zhu X, Ji X, Quanquin N, Deng YQ, Tian M, Aliyari R, Zuo X, Yuan L, Afridi SK. Chloroquine: FDA-approved drug prevents Zika virus infection and its associated congenital microcephaly in mice. *EBioMedicine* 2017; 24: 189-194.
21. Yan Y, Zou Z, Sun Y, Li X, Xu KF, Wei Y, Jin N, Jiang C. Anti-malaria drug chloroquine is highly effective in treating avian influenza A H5N1 virus infection in an animal model. *Cell Res* 2013; 23: 300-302.
22. Paton NI, Lee L, Xu Y, Ooi EE, Cheung YB, Archuleta S, Wong G, Smith AW. Chloroquine for influenza prevention: A randomised, double-blind, placebo-controlled trial. *Lancet Infect Dis* 2011; 11: 677-683.
23. Gonzalez-Dunia D, Cubitt B, de la Torre JC. Mechanism of Borna disease virus entry into cells. *J Virol* 1998; 72: 783-788.
24. Ros C, Burckhardt CJ, Kempf C. Cytoplasmic trafficking of minute virus of mice: low-pH requirement, routing to late endosomes, and proteasome interaction. *J Virol* 2002; 76: 12634-12645.
25. Diaz-Griffero F, Hoschander SA, Brojatsch J. Endocytosis is a critical step in entry of subgroup B avian leukosis viruses. *J Virol* 2002; 76: 12866-12876.
26. Ferreira D, Santo M, Rebello M, Rebello M. Weak bases affect late stages of Mayaro virus replication cycle in vertebrate cells. *J Med Microbiol* 2000; 49: 313-318.
27. Harley CA, Dasgupta A, Wilson DW. Characterization of herpes simplex virus-containing organelles by subcellular fractionation: Role for organelle acidification in assembly of infectious particles. *J Virol* 2001; 75: 1236-1251.
28. Randolph VB, Winkler G, Stollar V. Acidotropic amines inhibit proteolytic processing of flavivirus prM protein. *Virology* 1990; 174: 450-458.
29. Tsai WP, Nara PL, Kung HF, Oroszlan S. Inhibition of human immunodeficiency virus infectivity by chloroquine. *AIDS Res Hum Retroviruses* 1990; 6: 481-489.
30. Al-Kuraishy HM, Hussien NR, Al-Naimi MS, Al-Buhadily AK, Al-Gareeb AI, Lungnier C. Renin-Angiotensin system and fibrinolytic pathway in COVID-19: One-way skepticism. *BBRJ* 2020; 4: 33.
31. Al-Kuraishy HM, Al-Naimi MS, Lungnier CM, Al-Gareeb AI. Macrolides and COVID-19: An optimum premise. *BBRJ* 2020; 4: 189.
32. Al-Kuraishy HM, Al-Gareeb AI. Acute kidney injury and COVID-19. *Egypt J Intern Med* 2021; 33: 1-5.
33. Wolfram J, Nizzero S, Liu H, Li F, Zhang G, Li Z, Shen H, Blanco E, Ferrari M. A chloroquine-induced macrophage-preconditioning strategy for improved nanodelivery. *Sci Rep* 2017; 7: 13738.
34. Batiha GE, Gari A, Elshony N, Shaheen HM, Abubakar MB, Adeyemi SB, Al-Kuraishy HM. Hypertension and its management in COVID-19 patients: The assorted view. *Int J Cardiol Cardiovasc Risk Prev* 2021 Nov 13: 200121.
35. Onohuean H, Al-Kuraishy HM, Al-Gareeb AI, Qusti S, Alshammari EM, Batiha GE. Covid-19 and development of heart failure: mystery and truth. *Naunyn Schmiedebergs Arch Pharmacol* 2021; 394: 2013-2021.
36. Wang H, Yang P, Liu K, Guo F, Zhang Y, Zhang G, Jiang C. SARS coronavirus entry into host cells through a novel clathrin-and caveolae-independent endocytic pathway. *Cell Res* 2008; 18: 290-301.
37. Coutard B, Valle C, de Lamballerie X, Canard B, Seidah NG, Decroly E. The spike glycoprotein of the new coronavirus 2019-nCoV contains a

- furin-like cleavage site absent in CoV of the same clade. *Antiviral Res* 2020; 176: 104742.
38. Ozden S, Lucas-Hourani M, Ceccaldi PE, Basak A, Valentine M, Benjannet S, Hamelin J, Jacob Y, Mamchaoui K, Mouly V. Inhibition of Chikungunya virus infection in cultured human muscle cells by furin inhibitors impairment of the maturation of the E2 surface glycoprotein. *J Bio Chem* 2008; 283: 21899-21908.
 39. Schoeman D, Fielding BC. Coronavirus envelope protein: Current Knowledge. *Virology* 2019; 16: 1-22.
 40. Al-Kuraishy HM, Al-Gareeb AI, Alblihed M, Cruz-Martins N, Batiha GE. COVID-19 and risk of acute ischemic stroke and acute lung injury in patients with type ii diabetes mellitus: the anti-inflammatory role of metformin. *Front Med (Lausanne)* 2021; 8: 110.
 41. Kono M, Tatsumi K, Imai AM, Saito K, Kuriyama T, Shirasawa H. Inhibition of human coronavirus 229E infection in human epithelial lung cells (L132) by chloroquine: Involvement of p38 MAPK and ERK. *Antiviral Res* 2008; 77: 150-152.
 42. Yang M, Huang L, Li X, Kuang E. Chloroquine inhibits lytic replication of Kaposi's sarcoma-associated herpesvirus by disrupting mTOR and p38-MAPK activation. *Antiviral Res* 2016; 133: 223-233.
 43. Yao X, Ye F, Zhang M, Cui C, Huang B, Niu P, Liu X, Zhao L, Dong E, Song C. In vitro antiviral activity and projection of optimized dosing design of hydroxychloroquine for the treatment of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). *Clin Infect Dis* 2020; 71: 732-739.
 44. Al-Kuraishy HM, Al-Gareeb AI, Almulaiky YQ, Cruz-Martins N, Batiha GE. Role of leukotriene pathway and montelukast in pulmonary and extrapulmonary manifestations of Covid-19: The enigmatic entity. *Eur J Pharmacol* 2021 May 15: 174196.
 45. So LKY, Lau ACW, Yam LYC, Cheung, TMT, Poon E, Yung RWH, Yuen KY. Development of a standard treatment protocol for severe acute respiratory syndrome. *Lancet* 2003; 361: 1615-1617.
 46. Al-Kuraishy HM, Al-Gareeb AI, Faidah H, Al-Maiahy TJ, Cruz-Martins N, Batiha GE. The looming effects of estrogen in Covid-19: A Rocky Rollout. *Front Nutr* 2021; 8.
 47. Nauwynck H, Duan X, Favoreel H, Van Oostveldt P, Pensaert M. Entry of porcine reproductive and respiratory syndrome virus into porcine alveolar macrophages via receptor-mediated endocytosis. *J Gen Virol* 1999; 80: 297-305.
 48. Blau DM, Holmes KV. Human coronavirus HCoV-229E enters susceptible cells via the endocytic pathway. *Adv Exp Med Biol* 2001; 494: 193-198.
 49. Gao J, Tian Z, Yang X. Breakthrough: Chloroquine phosphate has shown apparent efficacy in treatment of COVID-19 associated pneumonia in clinical studies. *Biosci Trends* 2020; 14: 72-73.
 50. Jie Z, He H, Xi H, Zhi Z. Expert consensus on chloroquine phosphate for the treatment of novel coronavirus pneumonia. *Zhonghua* 2020; 43: 185-188.
 51. Bernstein H. Ocular safety of hydroxychloroquine. *Ann Ophthalmol* 1991; 23: 292-296.
 52. Ratliff NB, Estes ML, Myles JL, Shirey EK, McMahon JT. Diagnosis of chloroquine cardiomyopathy by endomyocardial biopsy. *N Engl J Med* 1987; 316: 191-193.
 53. Cubero GI, Reguero JR, Ortega JR. Restrictive cardiomyopathy caused by chloroquine. *Br Heart J* 1993; 69: 451-452.
 54. Al-Kuraishy HM, Al-Gareeb AI, Qusty N, Cruz-Martins N, Batiha GE. Sequential doxycycline and colchicine combination therapy in Covid-19: The salutary effects. *Pulm Pharmacol Ther* 2021; 67: 102008.
 55. Fox RI. Mechanism of action of hydroxychloroquine as an antirheumatic drug. *Semin Arthritis Rheum* 1993; 23: 82-91.
 56. Al-Kuraishy HM, Al-Gareeb AI, Al-Niemi MS, Al-Buhadily AK, Al-Harchan NA, Lugnier C. COVID-19 and phosphodiesterase enzyme type 5 inhibitors. *J Microsc Ultrastruct* 2020; 8: 141.
 57. Chen L, Liu H, Liu W, Liu J, Liu K, Shang J, Deng Y, Wei S. Analysis of clinical features of 29 patients with 2019 novel coronavirus pneumonia. *Chinese J Tubercul Resp Dis* 2020; 43: 5.
 58. Schrezenmeier E, Dörner T. Mechanisms of action of hydroxychloroquine and chloroquine: implications for Rheumatology. *Nat Rev Rheumatol* 2020; 16: 155-166.
 59. Shang L, Zhao J, Hu Y, Du R, Cao B. On the use of corticosteroids for 2019-nCoV pneumonia. *Lancet* 2020; 395: 683-684.
 60. Strand V, Ahadiet S, French J, Geier J, Krishnaswami S, Menon S, Checchio T, Tensfeldt TG, Hoffman E, Riese R. Systematic review and meta-analysis of serious infections with tofacitinib and biologic disease-modifying antirheumatic drug treatment in rheumatoid arthritis clinical trials. *Arthritis Res Ther* 2015; 17: 362.
 61. Kwiek JJ, Haystead TA, Rudolph J. Kinetic mechanism of quinone oxidoreductase 2 and its inhibition by the antimalarial quinolines. *Biochemistry* 2004; 43: 4538-4547.
 62. Varki A. Sialic acids as ligands in recognition phenomena. *The FASEB J* 1997; 11: 248-255.
 63. Al-Kuraishy HM, Al-Gareeb AI, Qusti S, Alshammari EM, Gyebi GA, Batiha GE. Covid-19-Induced Dysautonomia: A Menace of Sympathetic Storm. *ASN Neuro* 2021; 13: 17590914211057635.
 64. Al-Kuraishy HM, Al-Gareeb AI, Alzahrani KJ, Alexiou A, Batiha GE. Niclosamide for Covid-19: bridging the gap. *Mol Biol Rep* 2021 Oct 18: 1-8.
 65. Tricou V, Minh NN, Van TP, Lee SJ, Farrar J, Wills B, Tran HT, Simmons CP. A randomized controlled trial of chloroquine for the treatment of dengue in Vietnamese adults. *PLoS Negl Trop Dis* 2010; 4: 785.
 66. Gay B, Bernard E, Solignat M, Chazal N, Devaux C, Briant L. pH-dependent entry of chikungunya virus into *Aedes albopictus* cells. *Infect Genet Evol* 2012; 12: 1275-1281.
 67. Khan M, Santhosh S, Tiwari M, Lakshmana RP, Parida M. Assessment of in vitro prophylactic and therapeutic efficacy of chloroquine against Chikungunya virus in vero cells. *J Med Virol* 2010; 82: 817-824.
 68. Yang ZY, Huang Y, Ganesh L, Leung K, Kong WP, Schwartz O, Subbarao K, Nabel GJ. pH-depen-

- dent entry of severe acute respiratory syndrome coronavirus is mediated by the spike glycoprotein and enhanced by dendritic cell transfer through DC-SIGN. *J Virol* 2004; 78: 5642-5650.
69. Cassell S, Edwards J, Brown DT. Effects of lyso-somotropic weak bases on infection of BHK-21 cells by Sindbis virus. *J Virol* 1984; 52: 857-864.
 70. Savarino A, Gennero L, Sperber K, Boelaert J. The anti-HIV-1 activity of chloroquine. *J Clin Virol* 2001; 20: 131-135.
 71. Savarino A, Lucia MB, Rastrelli E, Rutella S, Gollotta C, Morra E, Tamburrini E, Perno CF, Boelaert JR, Sperber K. Anti-HIV effects of chloroquine: inhibition of viral particle glycosylation and synergism with protease inhibitors. *J Acquir Immune Defic Syndr* 2004; 35: 223-232.
 72. Klumperman J, Locker JK, Meijer A, Horzinek MC, Geuze HJ, Rottier P. Coronavirus M proteins accumulate in the Golgi complex beyond the site of virion budding. *J Virol* 1994; 68: 6523-6534.
 73. Perrier A, Bonnin A, Desmarests L, Danneels A, Goffard A, Rouillé Y, Dubuisson J, Belouzard S. The C-terminal domain of the MERS coronavirus M protein contains a trans-Golgi network localization signal. *J Bio Chem* 2019; 294: 14406-14421.
 74. Diebold SS, Kaisho T, Hemmi H, Akira S, e Sousa CR. Innate antiviral responses by means of TLR7-mediated recognition of single-stranded RNA. *Sci* 2004; 303: 1529-1531.
 75. Accapezzato D, Visco V, Francavilla V, Molette C, Donato T, Paroli M, Mondelli MU, Doria M, Torrisi MR, Barnaba V. Chloroquine enhances human CD8+ T cell responses against soluble antigens in vivo. *J Exp Med* 2005; 202: 817-828.
 76. Garulli B, Di Mario G, Sciaraffia E, Accapezzato D, Barnaba V, Castrucci MR. Enhancement of T cell-mediated immune responses to whole inactivated influenza virus by chloroquine treatment in vivo. *Vaccine* 2013; 31: 1717-1724.
 77. Seitz M, Valbracht J, Quach J, Lotz M. Gold sodium thiomalate and chloroquine inhibit cytokine production in monocytic THP-1 cells through distinct transcriptional and posttranslational mechanisms. *J Clin Immunol* 2003; 23: 477-484.
 78. Al-Kuraishy HM, Al-Gareeb AI, Alqarni M, Cruz-Martins N, El-Saber Batiha G. Pleiotropic Effects of Tetracyclines in the Management of COVID-19: Emerging Perspectives. *Front Pharmacol* 2021; 12: 136.
 79. Fuld H, Horwich L. Treatment of rheumatoid arthritis with chloroquine. *Br Med J* 1958; 2: 1199-1201.
 80. Mackenzie AH. Antimalarial drugs for rheumatoid arthritis. *Am J Med* 1983; 75: 48-58.
 81. Sharma TS, Joyce E, Wasko MCM. Anti-malarials: are there benefits beyond mild disease? *Curr Treatm Opt in Rheumatol* 2016; 2: 1-12.
 82. Lee SJ, Silverman E, Bargman JM. The role of antimalarial agents in the treatment of SLE and lupus nephritis. *Nat Rev Nephrol* 2011; 7: 718-729.
 83. Wozniacka A, Lesiak A, Narbutt J, McCauliffe D, Sysa-Jedrzejowska A. Chloroquine treatment influences proinflammatory cytokine levels in systemic lupus erythematosus patients. *Lupus* 2006; 15: 268-275.
 84. Sharma OP. Effectiveness of chloroquine and hydroxychloroquine in treating selected patients with sarcoidosis with neurological involvement. *Arch Neurol* 1998; 55: 1248-1254.
 85. Jang CH, Choi JH, Byun MS, Jue DM. Chloroquine inhibits production of TNF- α , IL-1 β and IL-6 from lipopolysaccharide-stimulated human monocytes/macrophages by different modes. *Rheumatol* 2006; 45: 703-710.
 86. Picot S, Peyron F, Donadille A, Vuillez J, Barbe G, Ambroise-Thomas P. Chloroquine-induced inhibition of the production of TNF, but not of IL-6, is affected by disruption of iron metabolism. *Immunol* 1993; 80: 127-133.
 87. Jeong JY, Jue DM. Chloroquine inhibits processing of tumor necrosis factor in lipopolysaccharide-stimulated RAW 264.7 macrophages. *J Immunol* 1997; 158: 4901-4907.
 88. Zhu X, Ertel W, Ayala A, Morrison M, Perrin M, Chaudry I. Chloroquine inhibits macrophage tumour necrosis factor- α mRNA transcription. *Immunol* 1993; 80: 122-126.
 89. Weber SM, Levitz SM. Chloroquine interferes with lipopolysaccharide-induced TNF- α gene expression by a nonlysosomal mechanism. *J Immunol* 2000; 165: 1534-1540.
 90. Jeong JY, Choi JW, Jeon KI, Jue DM. Chloroquine decreases cell-surface expression of tumour necrosis factor receptors in human histiocytic U-937 cells. *Immunol* 2002; 105: 83-91.
 91. Chloroquine interferes with dengue-2 virus replication in U937 cells. *Microbiol Immunol* 2014; 58: 318-326.
 92. Wang, PH. Increasing host cellular receptor—angiotensin-converting enzyme 2 (ACE2) expression by coronavirus may facilitate 2019-nCoV infection. *J Med Virol* 2020; 92: 2692-2701.
 93. Glowacka I, Bertram S, Müller MA, Allen P, Soil-leux E, Pfefferle S, Steffen I, Tsegaye TS, He Y, Gnirss K. Evidence that TMPRSS2 activates the severe acute respiratory syndrome coronavirus spike protein for membrane fusion and reduces viral control by the humoral immune response. *J Virol* 2011; 85: 4122-4134.
 94. Gupta N, Agrawal S, Ish P. Chloroquine in COVID-19: the evidence. *Monaldi Arch Chest Dis* 2020; 90.
 95. Cortegiani A, Ingoglia G, Ippolito M, Giarratano A, Einav SA systematic review on the efficacy and safety of chloroquine for the treatment of COVID-19. *J Crit Care* 2020; 57: 279-283.
 96. Al-Kuraishy HM, Al-Gareeb AI, Cruz-Martins N, Batiha GE. Hyperbilirubinemia in Gilbert syndrome attenuates Covid-19 induced-metabolic disturbances: A case-report study. *Front Cardiovasc Med* 2021; 8: 71.
 97. Herraiz T, Guillén H, González-Peña D, Arán VJ. Antimalarial Quinoline Drugs inhibit β -Hematin and increase free Hemin catalyzing peroxidative Reactions and inhibition of cysteine proteases. *Sci Rep* 2019; 9: 15398.
 98. Zhang MY, Zhang Y, Wu XD, Zhang K, Lin P, Bian HJ, Qin MM, Huang W, Wei D, Zhang Z. Disrupting CD147-RAP2 interaction abrogates erythrocyte invasion by *Plasmodium falciparum*. *Blood* 2018; 131: 1111-1121.

99. Wang K, Chen W, Zhou YS, Lian JQ, Zhang Z, Du P, Gong L, Zhang Y, Cui HY, Geng, JJ. SARS-CoV-2 invades host cells via a novel route: CD147-spike protein. *Bio Rev* 2020.
100. Gralinski LE, Baric RS. Molecular pathology of emerging coronavirus infections. *J Pathol* 2015; 235: 185-195.
101. Sheahan TP, Sims AC, Zhou S, Graham RL, Pruijssers AJ, Agostini ML, Leist SR, Schäfer A, Dinnon KH, Stevens LJ. An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice. *Sci Transl Med* 2020; 12: 5883.
102. Moubarak M, Kasozi KI, Hetta HF, Shaheen HM, Rauf A, Al-Kuraishy HM, Qusti S, Alshammari EM, Ayikobua ET, Ssempijja F, Afodun AM. The rise of SARS-CoV-2 variants and the role of convalescent plasma therapy for management of infections. *Life* 2021; 11: 734.
103. Luo W, Yu H, Gou J, Li X, Sun Y, Li J, Liu L. Clinical pathology of critical patient with novel coronavirus pneumonia (COVID-19). *Prepr* 2020; 20200-20407.
104. Al-Kuraishy HM, Al-Gareeb AI. From SARS-CoV to nCoV-2019: Ruction and argument. *Archives of Clinical Infectious Diseases* 2020; 15 (COVID-19).
105. Shaver CM, Upchurch CP, Janz DR, Grove BS, Putz ND, Wickersham NE, Dikalov SI, Ware LB, Bastarache JA. Cell-free hemoglobin: a novel mediator of acute lung injury. *Am J Physiol Lung Cell Mol Physiol* 2016; 310: 532-541.
106. Chloroquine interference with hemoglobin endocytic trafficking suppresses adaptive heme and iron homeostasis in macrophages: the paradox of an antimalarial agent. *Oxid Med Cell Longev* 2013; 2013: 870472.
107. Juurlink DN. Safety considerations with chloroquine, hydroxychloroquine and azithromycin in the management of SARS-CoV-2 infection. *CMAJ* 2020; 192: 450-453.
108. Bhaskaran M, Chen H, Chen Z, Liu L. Hemoglobin is expressed in alveolar epithelial type II cells. *Biochem Biophys Res Commun* 2005; 333: 1348-1352.
109. Ghosh A, Garee G, Sweeny EA, Nakamura Y, Stuehr DJ. Hsp90 chaperones hemoglobin maturation in erythroid and nonerythroid cells. *Proc Natl Acad Sci* 2018; 115: 1117-1126.
110. Choi S, Park YS, Koga T, Treloar A, Kim KC. TNF- α is a key regulator of MUC1, an anti-inflammatory molecule during airway *Pseudomonas aeruginosa* infection. *Am J Respir Cell Mol Biol* 2011; 44: 255-260.
111. Balla J, Jacob HS, Balla G, Nath K, Eaton JW, Vercellotti GM. Endothelial-cell heme uptake from heme proteins: induction of sensitization and desensitization to oxidant damage. *Proc Natl Acad Sci* 1993; 90: 9285-9289.
112. Bunn HF, Jandl JH. Exchange of heme among hemoglobins and between hemoglobin and albumin. *J Bio Chem* 1968; 243: 465-475.
113. Heme and the vasculature: an oxidative hazard that induces antioxidant defenses in the endothelium. *Artif Cells Blood Substit Immobil Biotechnol* 1994; 22: 207-213.
114. Schaer DJ, Buehler PW, Alayash AI, Belcher JD, Vercellotti GM. Hemolysis and free hemoglobin revisited: exploring hemoglobin and heme scavengers as a novel class of therapeutic proteins. *Blood* 2013; 121: 1276-1284.
115. COVID-19: attacks the 1-beta chain of hemoglobin and captures the porphyrin to inhibit human heme metabolism. *Prepr Revis* 2020.
116. Ponticelli C, Moroni G. Hydroxychloroquine in systemic lupus erythematosus (SLE). *Expert Opin Drug Saf* 2017; 16: 411-419.
117. Singal AK, Kormos-Hallberg C, Lee C, Sadagoparamanujam VM, Grady JJ, Freeman DH, Anderson KE. Low-dose hydroxychloroquine is as effective as phlebotomy in treatment of patients with porphyria cutanea tarda. *Clin Gastroenterol Hepatol* 2012; 10: 1402-1409.
118. Chloroquine and hydroxychloroquine as antimalarials and antivirals against SARS-CoV-2: The heme factor. 2020.
119. Hemin activation ameliorates HIV-1 infection via heme oxygenase-1 induction. *J Immunol* 2006; 176: 4252-4257.
120. Huang H, Konduru K, Solovena V, Zhou ZH, Kumari N, Takeda K, Nekhai S, Bavari S, Kaplan G.G, Yamada KM. Therapeutic potential of the heme oxygenase-1 inducer hemin against Ebola virus infection. *Curr Trends Immunol* 2016; 17: 117-123.
121. Esposito S, Tagliabue C, Bosis S, Principi N. Levofloxacin for the treatment of *Mycoplasma pneumoniae*-associated meningoencephalitis in childhood. *Int J Antimicrob Agents* 2011; 37: 472-475.
122. Pellmar T. Electrophysiological correlates of peroxide damage in guinea pig hippocampus in vitro. *Brain Res* 1986; 364: 377-381.
123. Paeshuysse J, Coelmont L, Vliegen I, Vandekerckhove J, Peys E, Sas B, De Clercq E, Neyts J. Hemin potentiates the anti-hepatitis C virus activity of the antimalarial drug artemisinin. *Biochem Biophys Res Commun* 2006; 348: 139-144.
124. Lam HC, Siroky BJ, Henske EP. Renal disease in tuberous sclerosis complex: pathogenesis and therapy. *Nat Rev Nephrol* 2018; 14: 704-716.
125. Guo X, Zhang Y, Zheng L, Zheng C, Song J, Zhang Q, Kang B, Liu Z, Jin L, Xing R. Global characterization of T cells in non-small-cell lung cancer by single-cell sequencing. *Nat Med* 2018; 24: 978-985.
126. Drakesmith H, Prentic A. Viral infection and iron metabolism. *Nat Rev Microbiol* 2008; 6: 541-552.
127. Camaschella C, Nai A, Silvestri L. Iron metabolism and iron disorders revisited in the hepcidin era. *Haematologica* 2020; 105: 260-272.
128. Nguyen NB, Callaghan KD, Ghio AJ, Haile DJ, Yang F. Hepcidin expression and iron transport in alveolar macrophages. *Am J Physiol Lung Cell Mol Physiol* 2006; 291: 417-425.
129. Nairz M, Dichtl S, Schroll A, Haschka D, Tymoszek P, Theurl I, Weiss G. Iron and innate antimicrobial immunity—Depriving the pathogen, defending the host. *J Trace Elem Med Biol* 2018; 48: 118-133.
130. Biasiotto G, Di Lorenzo D, Archetti S, Zanella I. Iron and Neurodegeneration: Is Ferritinophagy the Link? *Mole Neurobiol* 2016; 53: 5542-5574.

131. Emerson LR, Nau ME, Martin RK, Kyle DE, Vahey M, Wirth DF. Relationship between Chloroquine Toxicity and Iron Acquisition in *Item*, *Saccharomyces cerevisiae*. *Antimicrob Agent Chemother* 2002; 46: 787-796.
132. Octave JN, Schneider YJ, Hoffmann P, Trouet A, Crichton RR. Transferrin Uptake by Cultured Rat Embryo Fibroblasts. *Eur J Biochem* 1982; 123: 235-240.
133. Shawki A, Knight PB, Maliken BD, Niespodzany EJ, Mackenzie B. Chapter Five - H+-Coupled Divalent Metal-Ion Transporter-1: Functional Properties, Physiological Roles and Therapeutics. *Curr Top Membr* 2012; 70: 169-214.
134. Wolfram J, Nizzero S, Liu H, Li F, Zhang G, Li Z, Shen H, Blanco E, Ferrari M. A chloroquine-induced macrophage-preconditioning strategy for improved nanodelivery. *Sci Rep* 2017; 7: 13738.
135. Potter MD, Shinpock SG, Popp RA, Popp DM, Godfrey V, Carpenter DA, Bernstein A, Johnson DK, Rinchik EM. Mutations in the Murine Fitness 1 Gene Result in Defective Hematopoiesis. *Blood* 1997; 90: 1850-1857.
136. The possible mechanisms of action of 4-aminoquinolines (chloroquine/hydroxychloroquine) against SARS-CoV-2 infection (COVID-19): A role for iron homeostasis? *Pharmacol Res* 2020; 158: 104904.
137. Klionsky DJ, Abdelmohsen K, Abe A, Abedin MJ, Abeliovich H, Acevedo AA, Adachi H, Adams CM, Adams PD, Adeli K. Guidelines for the use and interpretation of assays for monitoring autophagy. *Autophagy* 2016; 12: 1-222.
138. Kong Z, Liu R, Cheng Y. Artesunate alleviates liver fibrosis by regulating ferroptosis signaling pathway. *Biomed Pharmacother* 2019; 109: 2043-2053.
139. Hamming I, Timens W, Bulthuis MLC, Lely AT, Navis GJ, Van Goor H. Tissue distribution of ACE2 protein, the functional receptor for SARS coronavirus: A first step in understanding SARS pathogenesis. *J Pathol* 2004; 203: 631-637.
140. Tiensiwakul P, Husain SS. Effect of mouse hepatitis virus infection on iron retention in the mouse liver. *Br J Exp Pathol* 1979; 60: 161-166.
141. Al-Kuraishy HM, Al-Gareeb AI. Comparison of deferasirox and deferoxamine effects on iron overload and immunological changes in patients with blood transfusion-dependent β -thalassemia. *Asian J Transfus Sci* 2017; 11: 13.
142. Cao X. COVID-19: Immunopathology and its implications for therapy. *Nat Rev Immunol* 2020; 20: 269-270.
143. Frieman M, Heise M, Baric R. SARS coronavirus and innate immunity. *Virus Res* 2008; 133: 101-112.
144. Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. *Semin Immunopathol* 2017; 39: 529-539.
145. Dandekar AA, Perlman S. Immunopathogenesis of coronavirus infections: implications for SARS. *Nat Rev Immunol* 2005; 5: 917-927.
146. Recalcati S, Locati M, Gammella E, Invernizzi P, Cairo G. Iron levels in polarized macrophages: Regulation of immunity and autoimmunity. *Autoimm Rev* 2012; 11: 883-889.
147. Legssyer R, Ward RJ, Crichton RR, Boelaert JR. Effect of chronic chloroquine administration on iron loading in the liver and reticuloendothelial system and on oxidative responses by the alveolar macrophages. *Biochem Pharmacol* 1999; 57: 907-911.
148. Toll A, Celis R, Ozalla MD, Ercilla MG, Herrero C. Haemochromatosis gene mutations and response to chloroquine in sporadic porphyria cutanea tarda. *Acta Derm Venereol* 2006; 86: 279-280.
149. Ouyang Q, Huang Z, Wang Z, Chen X, Ni J, Lin L. Effects of pristane alone or combined with chloroquine on macrophage activation, oxidative stress, and TH1/TH2 skewness. *J Immunol Res* 2014; 2014: 613136.
150. Martins AC, Almeida JI, Lima IS, Kapitão AS, Gozzelino R. Iron metabolism and the inflammatory response. *IUBMB life* 2017; 69: 442-450.
151. Kemp JD. The role of iron and iron binding proteins in lymphocyte physiology and pathology. *J Clin Immunol* 1993; 13: 81-92.
152. Muriuki JM, Atkinson SH. How eliminating malaria may also prevent iron deficiency in African children. *Pharmaceut* 2018; 11: 96.
153. Drakesmith H, Prentice AM. Hepcidin and the iron-infection axis. *Sci* 2012; 338: 768-772.
154. Ganz T. Iron and infection. *Int J Hematol* 2018; 107: 7-15.
155. Camaschella C, Nai A, Silvestri L. Iron metabolism and iron disorders revisited in the hepcidin era. *Hematologica* 2020; 105: 260-272.
156. Lee P, Peng H, Gelbart T, Wang L, Beutler E. Regulation of hepcidin transcription by interleukin-1 and interleukin-6. *Proc Natl Acad Sci* 2005; 102: 1906-1910.
157. Shu W, Pang Z, Xu C, Lin J, Li G, Wu W, Sun S, Li J, Li X, Liu Z. Anti-TNF- α monoclonal antibody therapy improves anemia through downregulating hepatocyte hepcidin expression in inflammatory bowel disease. *Mediators Inflamm* 2019; 2019: 4038619.
158. Hadadi A, Mortezaazadeh M, Kolahdouzan K, Alavian G. Does recombinant human erythropoietin administration in critically ill COVID-19 patients have miraculous therapeutic effects? *J Med Virol* 2020; 92: 915-918.
159. Wenzhong L, Hualan L. COVID-19: Attacks the 1-beta chain of hemoglobin and captures the porphyrin to inhibit human heme metabolism. *Chem Rev* 2020.
160. Keung YK, Owen J. Iron deficiency and thrombosis: literature review. *Clin Appl Thromb Hemost* 2004; 10: 387-391.
161. Azab SF, Abdelsalam SM, Saleh SH, Elbehedy RM, Lotfy SM, Esh AM, Srea MA, Aziz KA. Iron deficiency anemia as a risk factor for cerebrovascular events in early childhood: a case-control study. *Ann Hematol* 2014; 93: 571-576.
162. Chang YL, Hung SH, Ling W, Lin HC, Li HC, Chung SD. Association between ischemic stroke and iron-deficiency anemia: A population-based study. *PloS one* 2013; 8: 82952.
163. Song AB, Kuter DJ, Al-Samkari H. Characterization of the rate, predictors, and thrombotic complications of thrombocytosis in iron deficiency anemia. *Am J Hematol* 2020; 95: 1180-1186.
164. Yin S, Huang M, Li D, Tang N. Difference of coagulation features between severe pneumonia

- induced by SARS-CoV2 and non-SARS-CoV2. *J Thromb Thrombol* 2021; 51: 1107-1110.
165. Tang N, Bai H, Chen X, Gong J, Li D, Sun Z. Anti-coagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. *J Thromb Haemost* 2020; 18: 1094-1099.
166. Jimenez K, Leitner F, Leitner A, Scharbert G, Schwabl P, Kramer AM, Krnjic A, Friske J, Helbich T, Evstatiev R. Iron deficiency induced thrombocytosis increases thrombotic tendency in rats. *Haematologica* 2021; 106: 782-794.
167. Thachil J. The versatile heparin in COVID-19. *J Thromb Haemost* 2020; 18: 1020-1022.
168. Abreu R, Essler L, Loy A, Quinn F, Giri P. Heparin inhibits intracellular *Mycobacterium tuberculosis* bacterial replication by reducing iron levels in human macrophages. *Sci Rep* 2018; 8: 7296: 1-12.
169. Bernstein HN. Ophthalmologic considerations and testing in patients receiving long-term antimalarial therapy. *Am J Med* 1983; 75: 25-34.
170. Parikh C, Edelhauser H. Ocular surgical pharmacology: Corneal endothelial safety and toxicity. *Curr Opin Ophthalmol* 2003; 14: 178-185.
171. Klinger G, Morad Y, Westall CA, Laskin C, Spitzer KA, Koren G, Ito S, Buncic RJ. Ocular toxicity and antenatal exposure to chloroquine or hydroxychloroquine for rheumatic diseases. *Lancet* 2001; 358: 813-814.
172. De Olano J, Howland MA, Su MK, Hoffman RS, Biary R. Toxicokinetics of hydroxychloroquine following a massive overdose. *Am J Emerg Med* 2019; 37: 2264-2268.
173. Guastalegname M, Vallone A. Could chloroquine/hydroxychloroquine be harmful in coronavirus disease 2019 (COVID-19) treatment? *Clin Infect Dis* 2020; 71: 888-889.
174. Omotoso OE, Awoyemi PP, Wahab VJ, Ragab M, Teibo JO, Akinfe O, et al. Knowledge and adherence to coronavirus disease 2019 preventive measures: A bi-national web-based survey. *Saudi J Health Sci* 2021;10:80-7
175. Teibo JO, Teibo T KA, Omotoso OE, Olagunju AS, Omotoso E. A bi-continental review of the knowledge and adherence to COVID-19 public health guidelines in north and south America. *Infectious Diseases & Tropical Medicine* 2021; 7: e728